



Protein Structure, Function and Methods of Analysis I

Dr. Abel Baerga Ortiz
Bioquímica

Oficina: A626

Teléfono: x1603

Email: abel.baerga@upr.edu



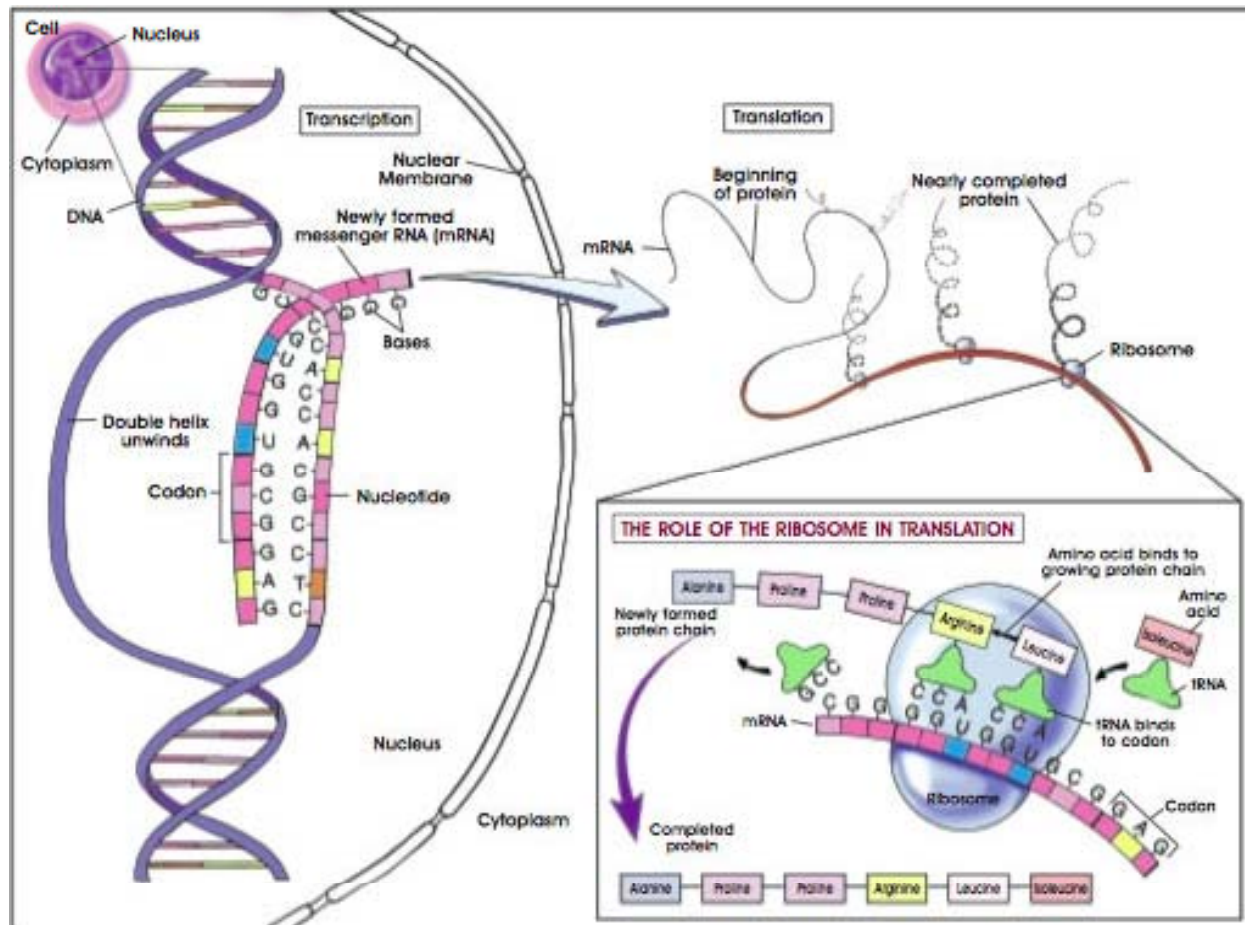
Referencias:

Lieberman, M; Marks, AD. Basic Medical Biochemistry: A Clinical Approach, 3rd Edition, 2009

Devlin, Thomas M. Textbook of Biochemistry with Clinical Correlations, 6th Edition, 2006

Nelson, DL; Cox, MM. Lehninger Principles of Biochemistry, 3rd Edition 2000

Proteins



Prion Disease and Protein Misfolding



- A) Creutzfeldt-Jacob Disease CJD**
- B) Bovine Spongiform Encephalopathy**
- C) Kuru**
- D) Fatal familial insomnia**

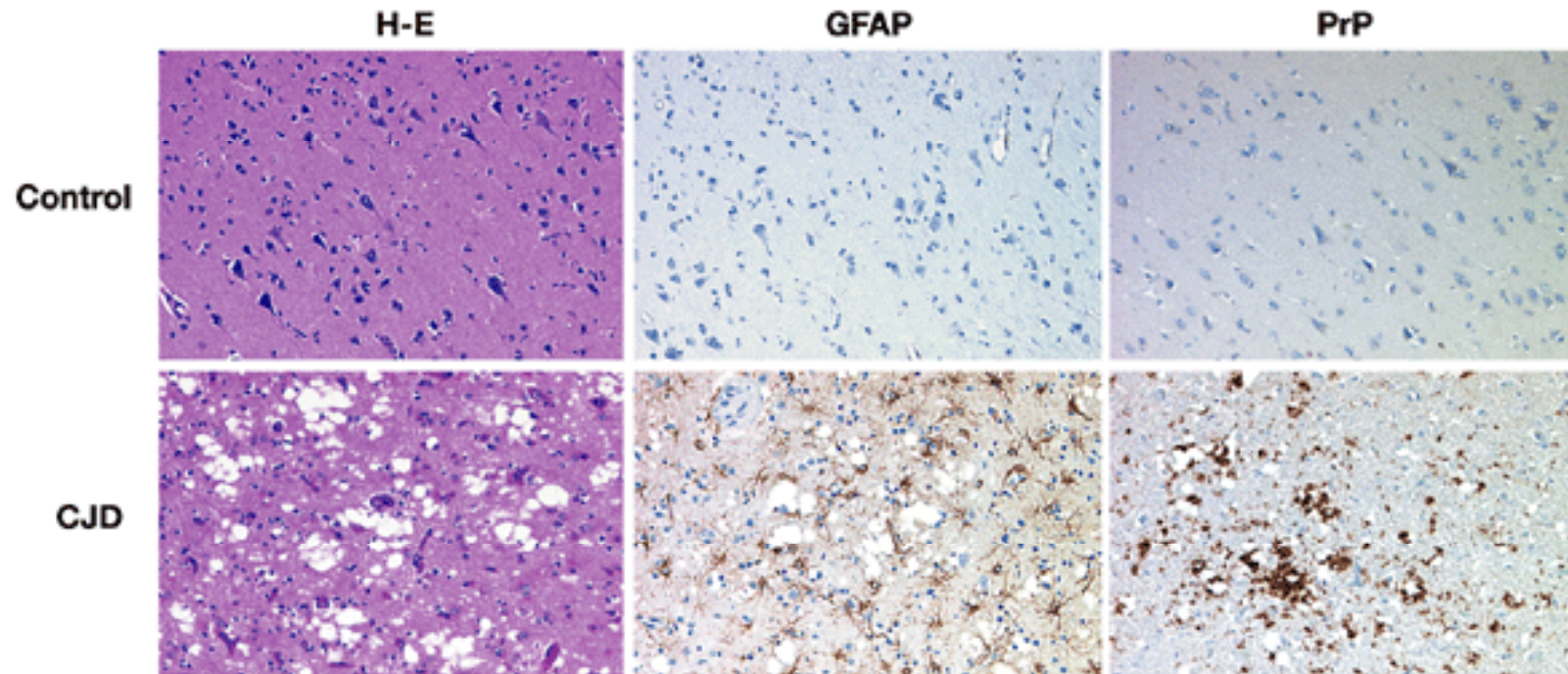
Histopathology:

spongiform changes in the brain

degeneration of neurons

astrocytosis

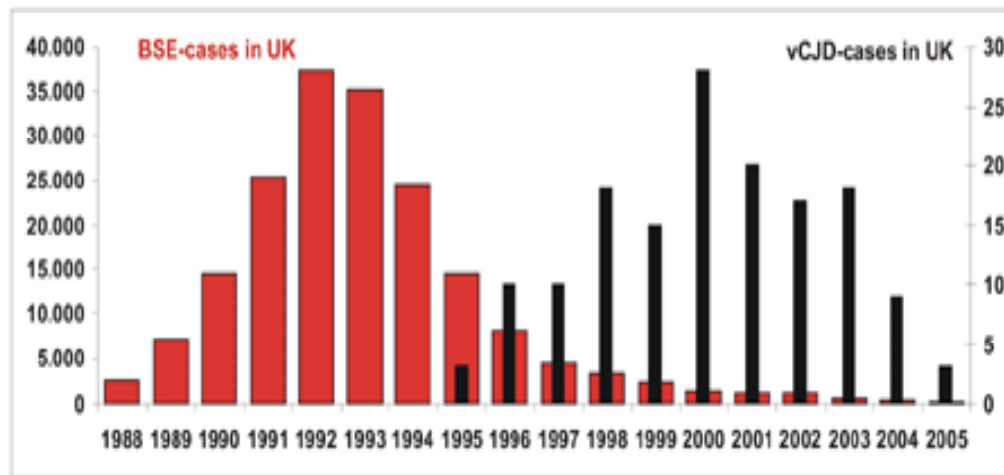
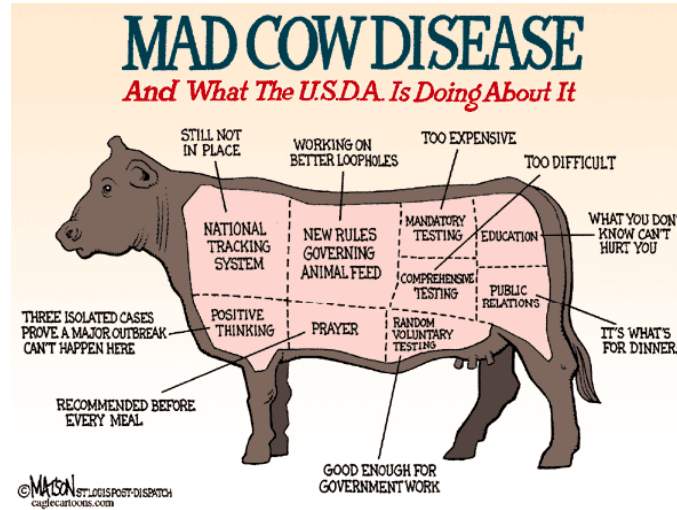
Prion Disease and Protein Misfolding



Nature Reviews | Neuroscience

(pictures Adriano Aguzzi, Markus Glatzel, Fabio Montrasio, Marco Prinz & Frank L. Heppner
Nature Reviews Neuroscience **2**, 745-749)

Prion Disease and Protein Misfolding

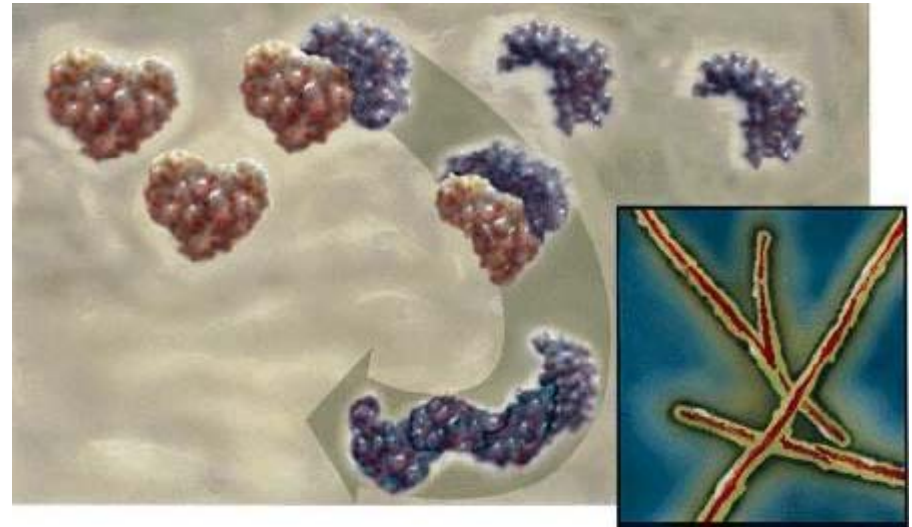


Prion Disease and Protein Misfolding



Stanley B. Prusiner

Nobel Prize in Medicine
For the Prion Hypothesis



Irreversible conformational self –
replication of prion protein

Prion Disease and Protein Misfolding

Cellular prion protein (PrP^c)

non-infectious
monomer
soluble (in mild detergents)
structure: predominantly α -helical
Proteinase K (PK) sensitive

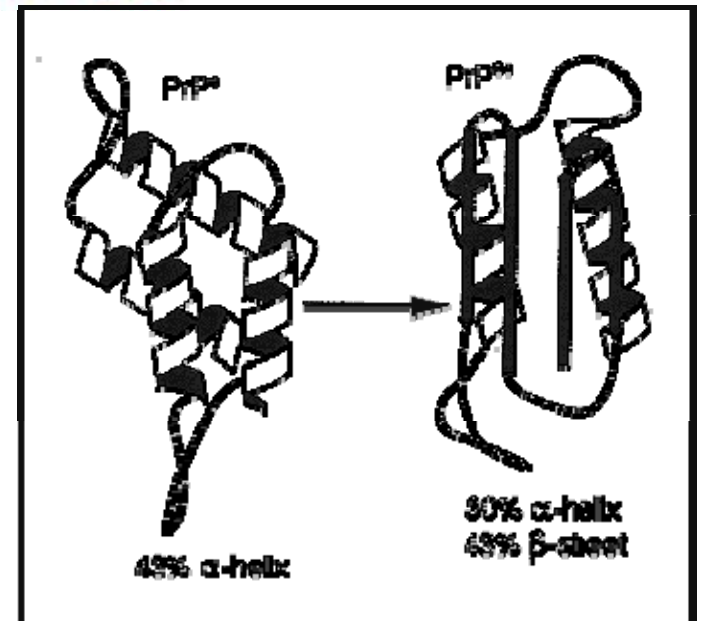


Scrapie-associated prion protein (PrP^{Sc})

infectious
aggregate
insoluble
structure: rich in β -sheets
partial PK-resistant

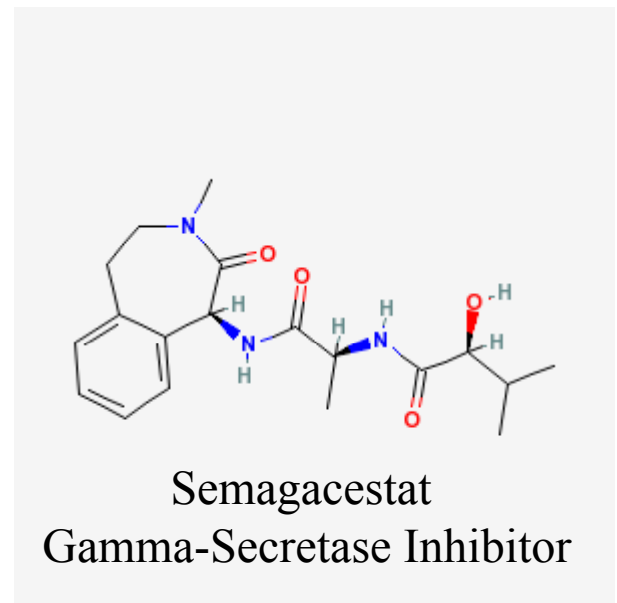
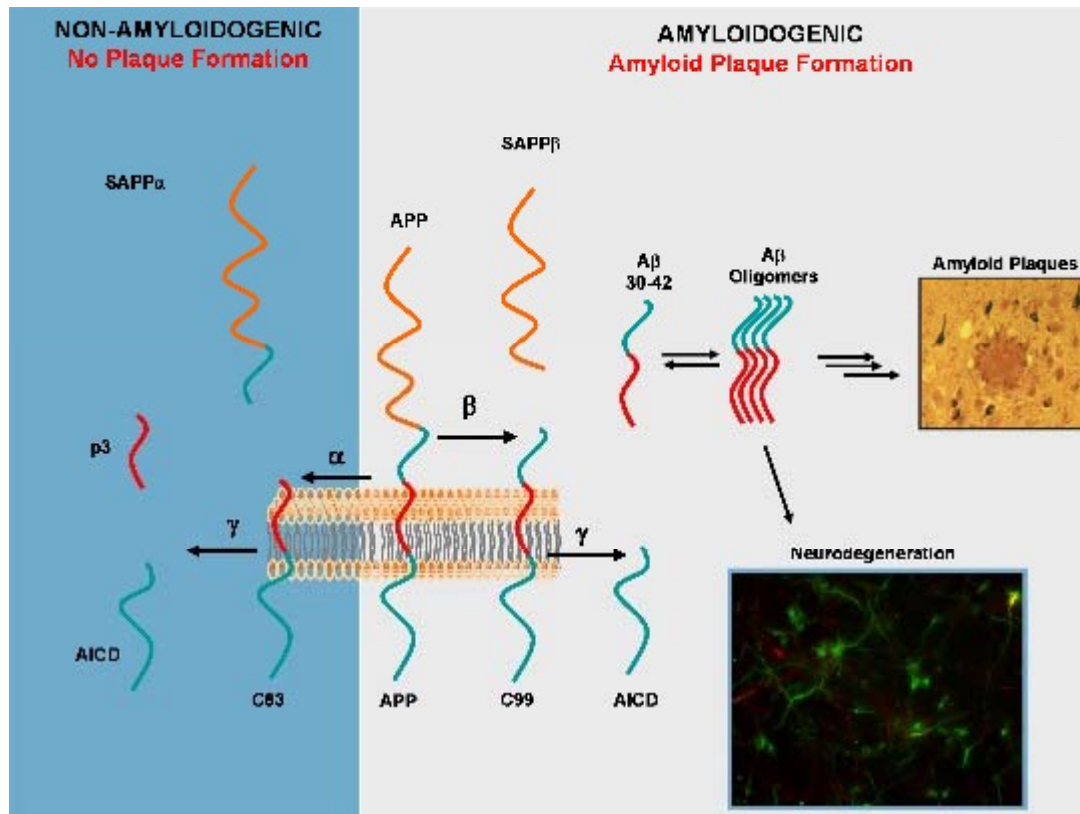


https://www.youtube.com/watch?v=6-Tz8a_vgX0

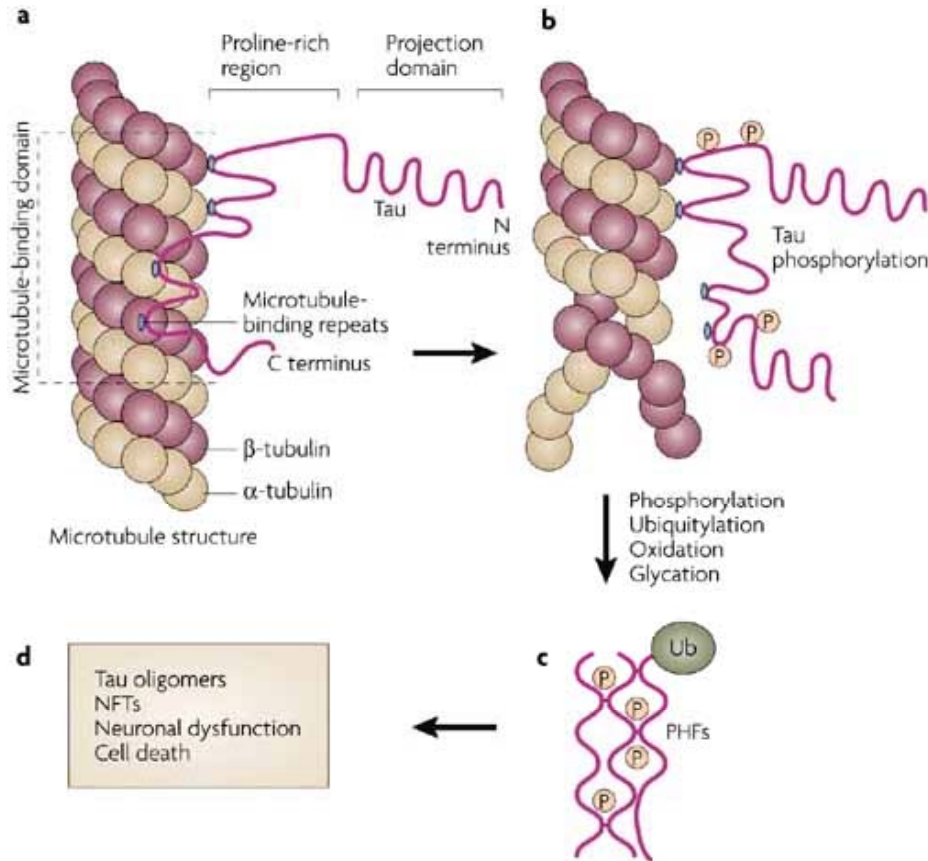


Amyloid Plaques and Alzheimer's Disease

Protein oligomers form the plaque normally associated with Alzheimer's Disease



Neurofibrillary Tangles and Alzheimer's Disease



Tau is a protein present in neurons only.

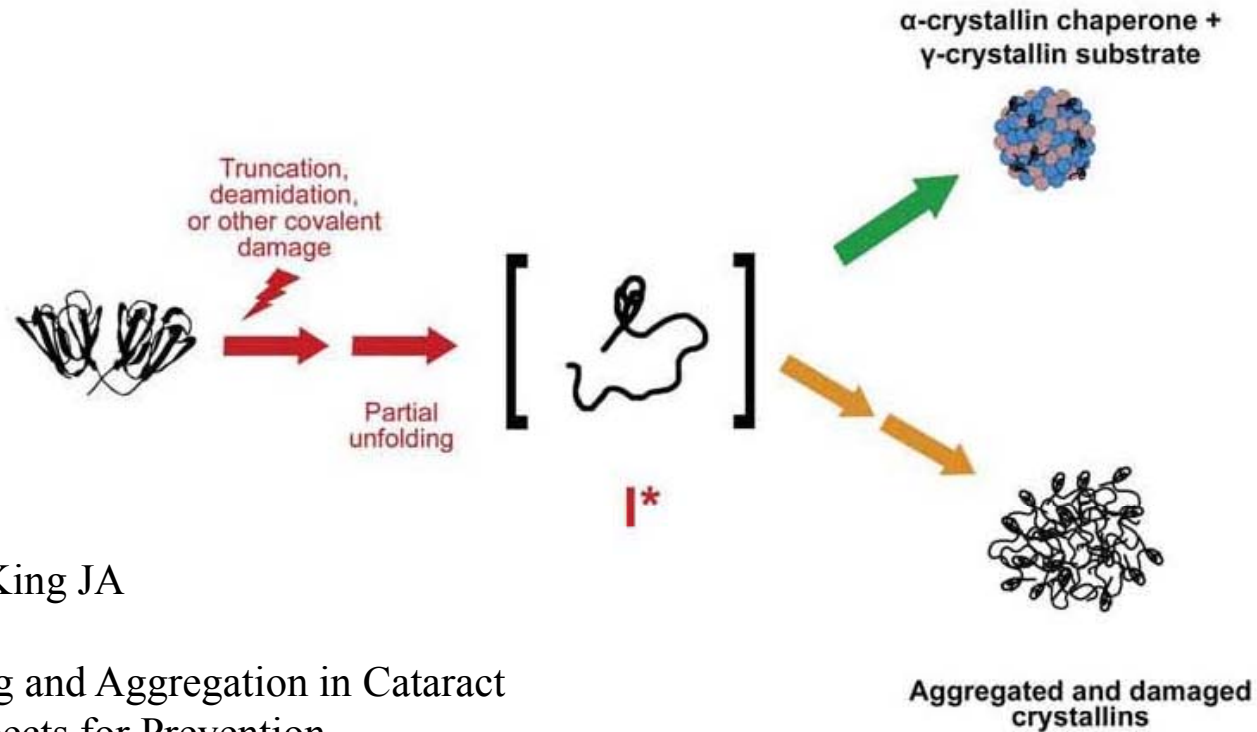
Tau stabilizes microtubules

Hyper-phosphorylation results in aggregation

Tau aggregates are associated with Alzheimer and other dementias.

Lens Proteins and Damage Accumulation

With age, covalent protein damage accumulates through pathways thought to include **UV radiation**, **oxidation**, **deamidation**, and **truncations**.



Moreau KL and King JA

Protein Misfolding and Aggregation in Cataract Disease and Prospects for Prevention

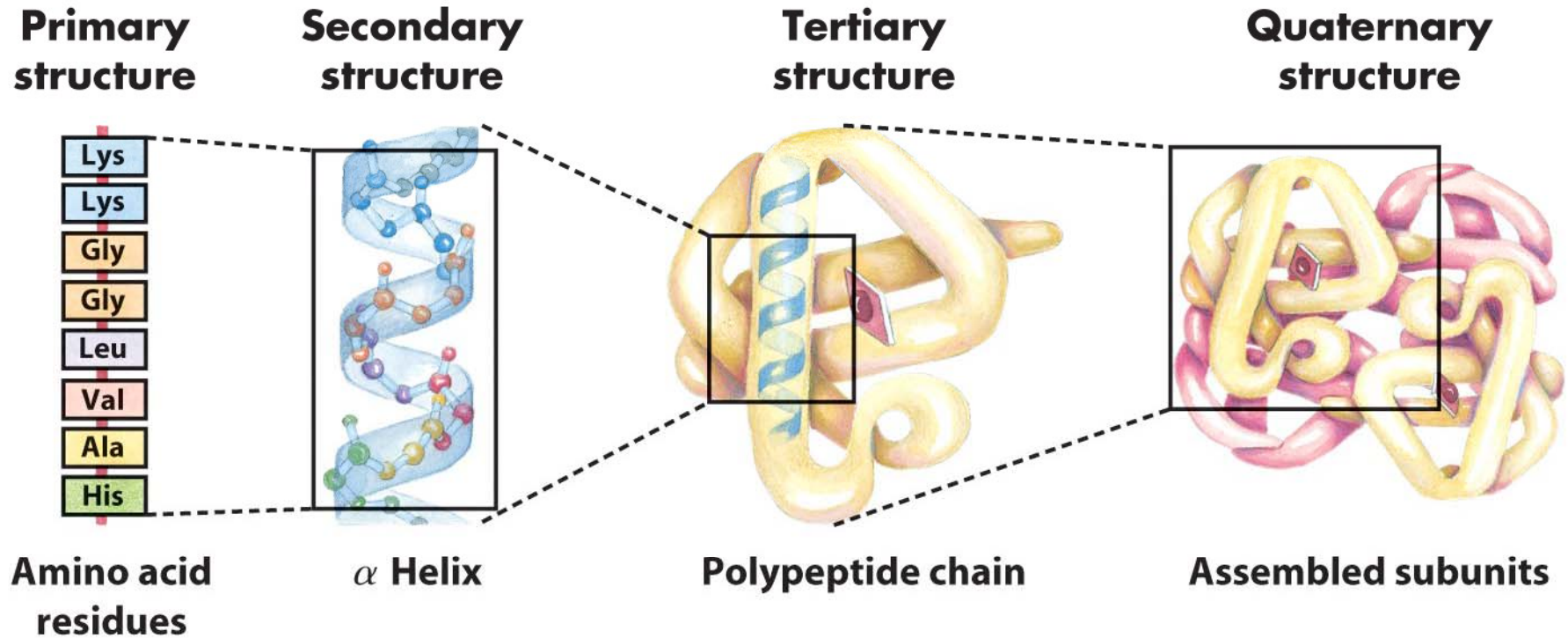
Trends Mol Med 2012 May ; 18(5): 273–282

Protein Structure, Function and Methods of Analysis I

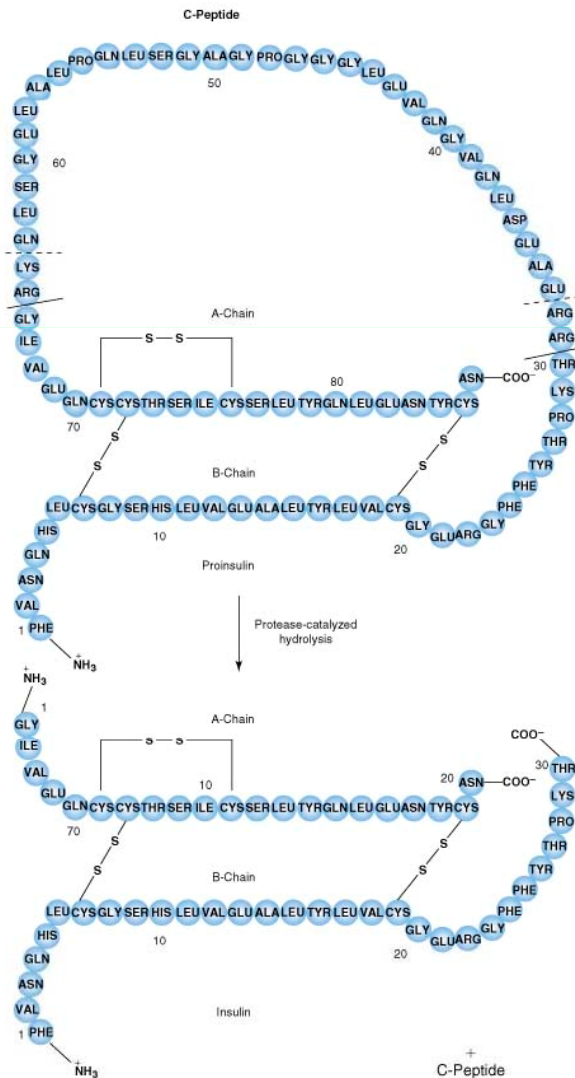


- I. Protein Structure – Primary, Secondary, Tertiary, etc
- II. Chemical Properties of Proteins – pH, UV absorbance
- III. Protein Stabilization and Denaturation
- IV. Post-translational Modifications
- V. Analysis of Proteins
 - i. Electrophoresis – PAGE, IEF
 - ii. Fluorescence – for sub-cellular localization
 - iii. Chromatography
 - iv. Primary Structure Determination – ID
 - v. Glycosylation

The Basics of Protein Structure

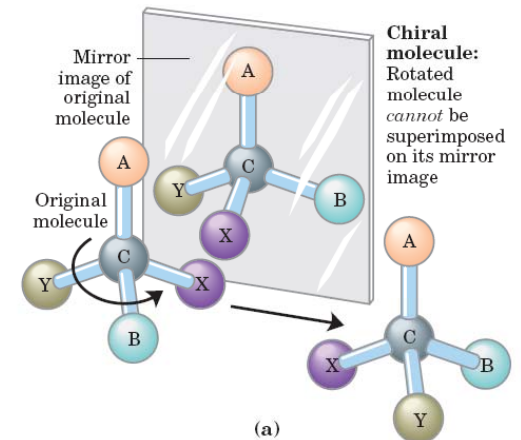
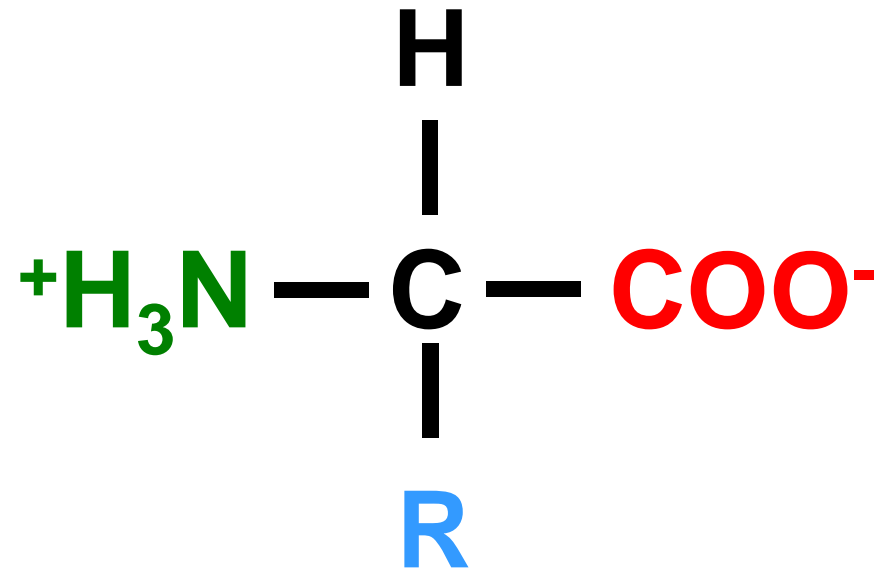


Primary Structure

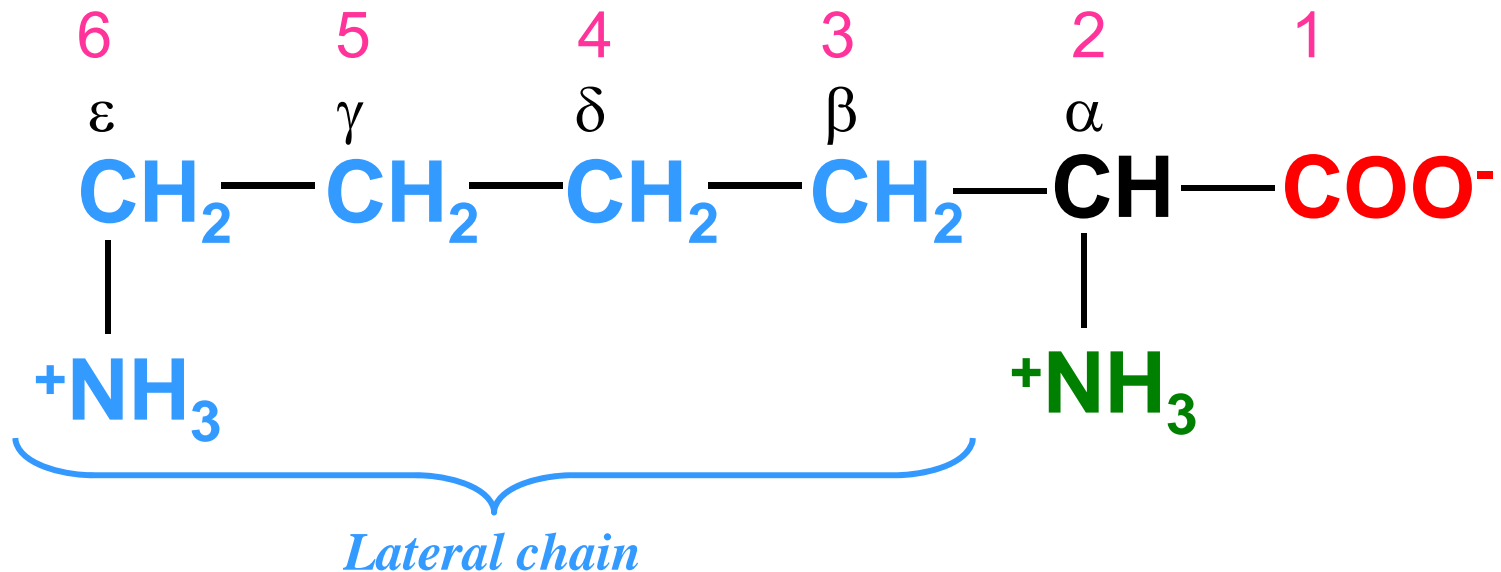


- Refers to the amino acid sequence of the protein
- Stabilized by the peptide bond
- Very stable, to “destroy” need:
 - ✓ 6 N HCl 100 -110°C 18 - 36 hrs.
 - ✓ Problem with acid hydrolysis:
 - Destroys Trp
 - glutamine & asparagine convert into glutamic acid & aspartic acid respectively

Amino Acids - Structure

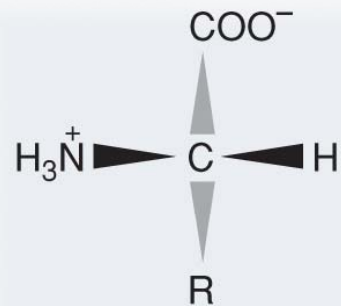
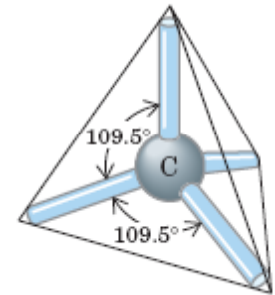


Amino Acids - Nomenclature

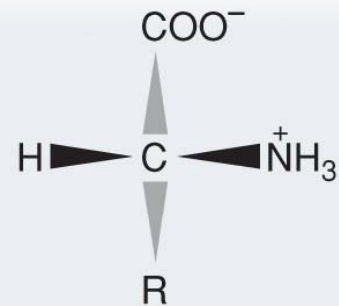


lysine

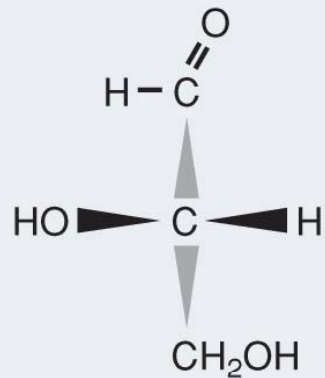
Absolute Configuration



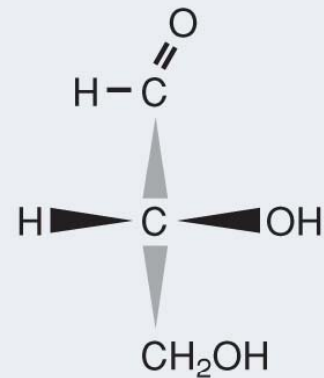
L-amino Acid



D-amino acid



L-glyceraldehyde



D-glyceraldehyde

Amino Acids - Nomenclature

1 Unique first letter:

C ysteine	=	Cys	=	C
H istidine	=	His	=	H
I soleucine	=	Ile	=	I
M ethionine	=	Met	=	M
S erine	=	Ser	=	S
V aline	=	Val	=	V

2 Most commonly occurring amino acids have priority:

A lanine	=	Ala	=	A
G lycine	=	Gly	=	G
L eu c ine	=	Leu	=	L
P roline	=	Pro	=	P
T hreonine	=	Thr	=	T

3 Similar sound names:

A rginine	=	Arg	=	R	("a R ginine")
A sparagine	=	Asn	=	N	(contains N)
A spartate	=	Asp	=	D	("aspar D ic")
G lutamate	=	Glu	=	E	("glut E mate")
G lutamine	=	Gln	=	Q	("Q-tamine")
P henylalanine	=	Phe	=	F	("Fenylalanine")
T yrosine	=	Tyr	=	Y	("t Y rosine")
T ryptophan	=	Trp	=	W	(double ring in the molecule)

4 Letter close to initial letter:

A spartate or asparagine	=	Asx	=	B	(near A)
G lutamate or glutamine	=	Glx	=	Z	
L ysine	=	Lys	=	K	(near L)
Undetermined amino acid	=		=	X	

Amino acids found in proteins

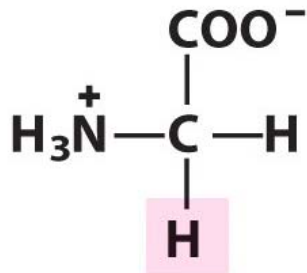


- C α is α -carboxyl & α -amino
- configuration L-
- Only 20 can be incorporated into proteins.
 - ✓ Proteins can have amino acids derivatives, but those modifications are integrated after protein synthesis.

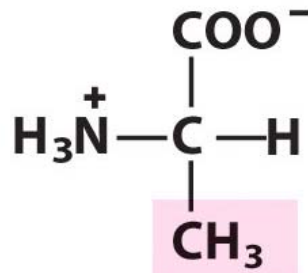
Classification of amino acids

- Properties of group R
 - ✓ Non-polar
 - ✓ Polar, uncharged
 - ✓ Polar, charged
 - Basic
 - Acids
- Synthesis
 - ✓ Essentials
 - ✓ Non- Essentials

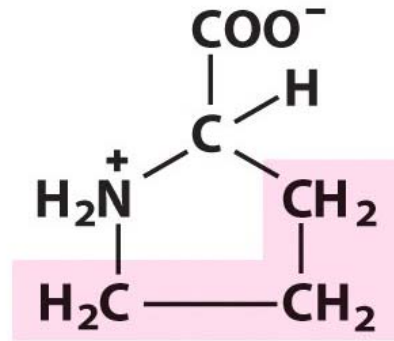
Nonpolar, aliphatic R groups



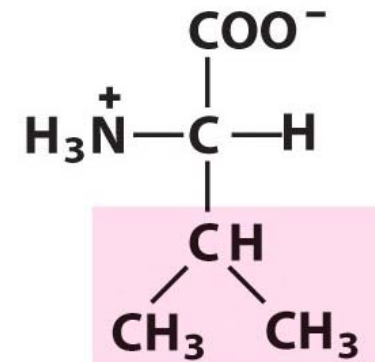
Glycine



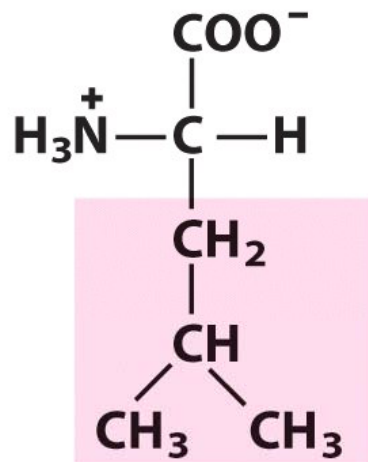
Alanine



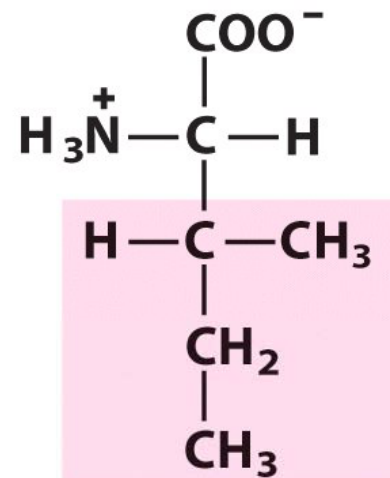
Proline



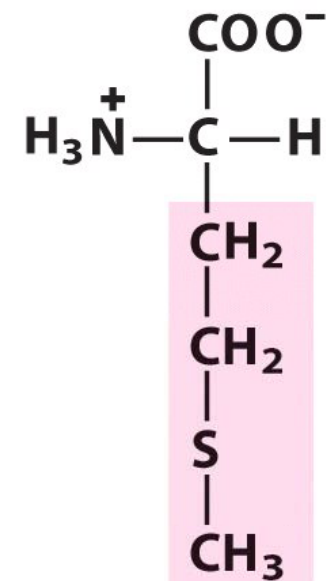
Valine



Leucine



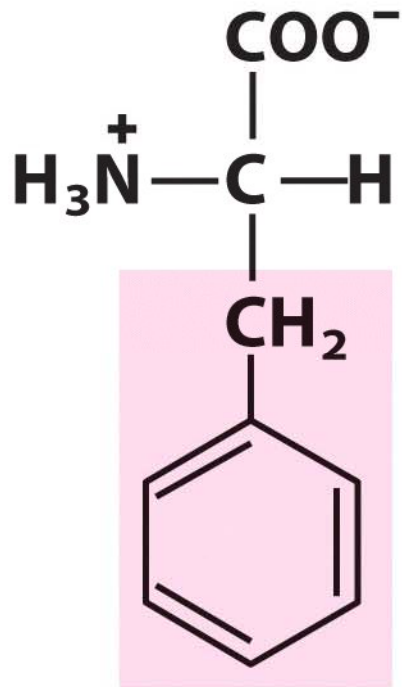
Isoleucine



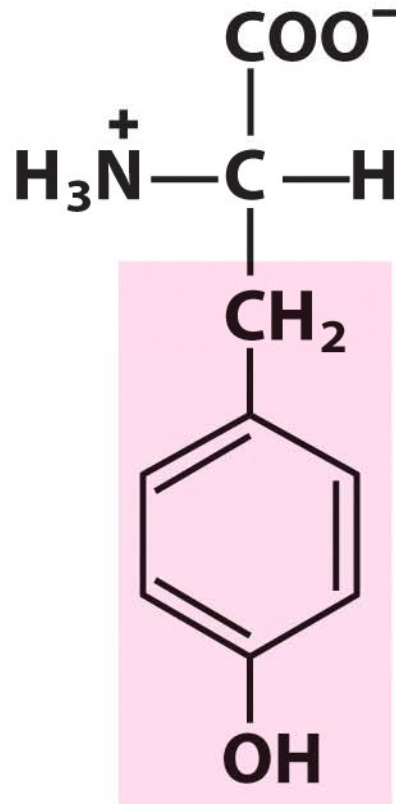
Methionine

*Devlin:
Fig 3.3
Page 78*

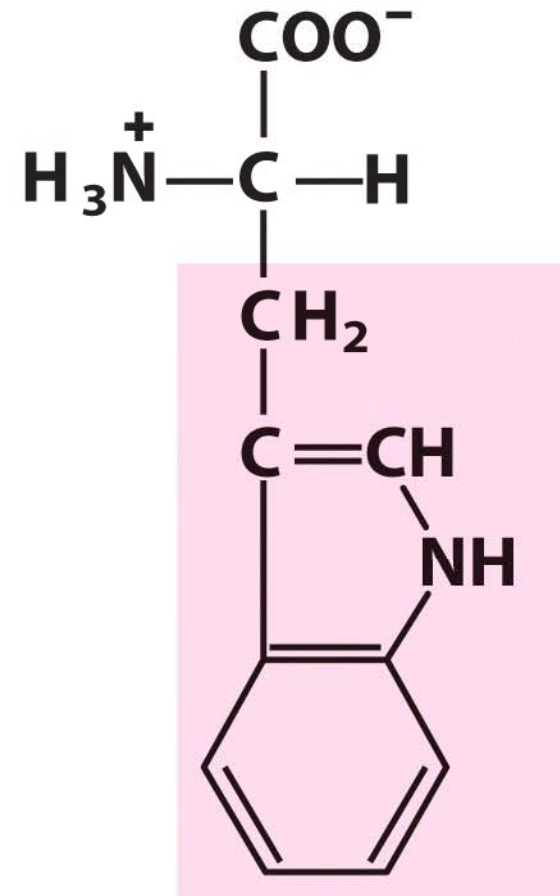
Aromatic R groups



Phenylalanine

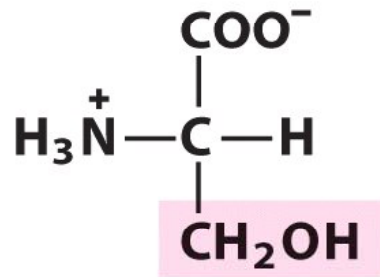


Tyrosine

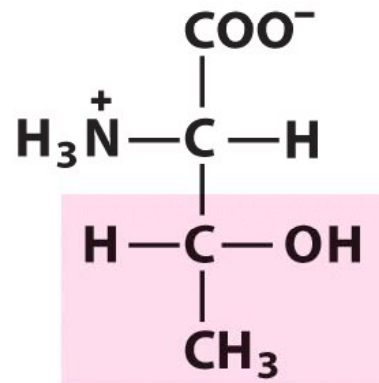


Tryptophan

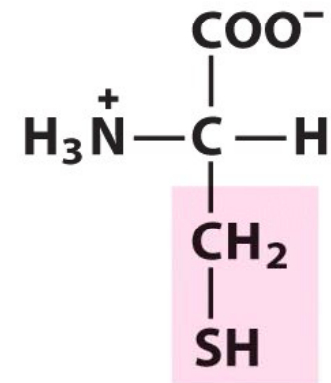
Polar, uncharged R groups



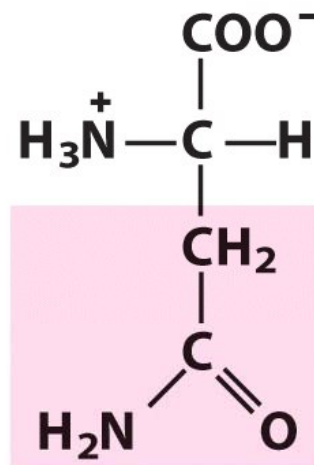
Serine



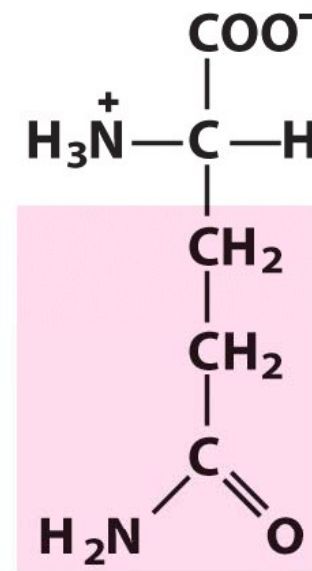
Threonine



Cysteine

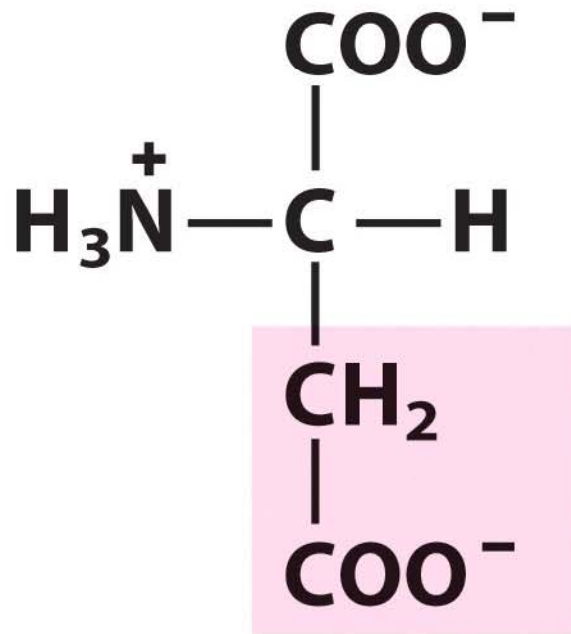


Asparagine

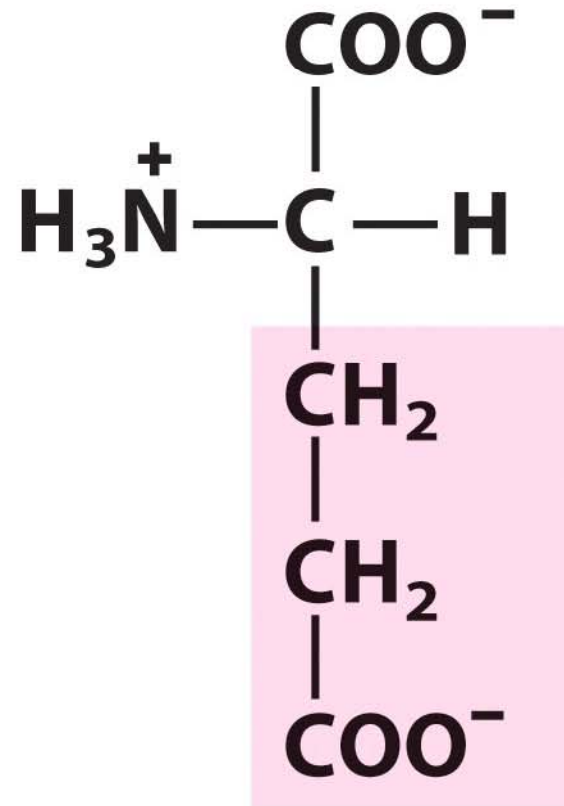


Glutamine

Negatively charged R groups

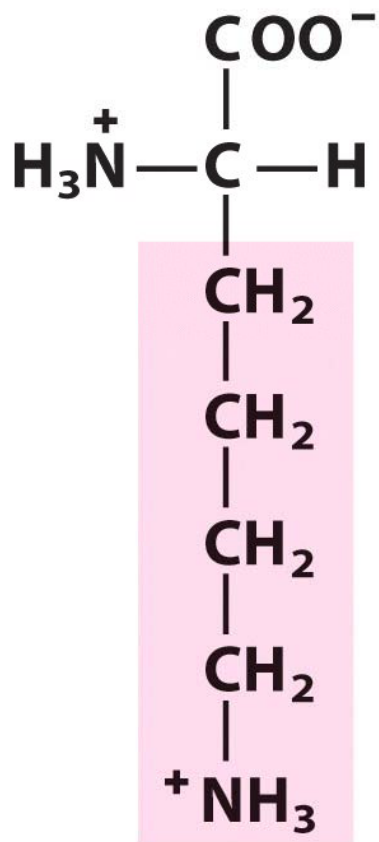


Aspartate

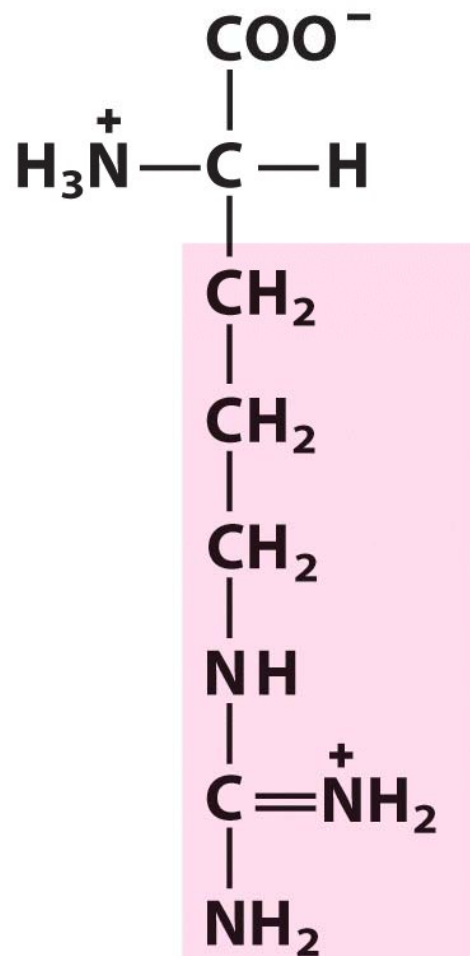


Glutamate

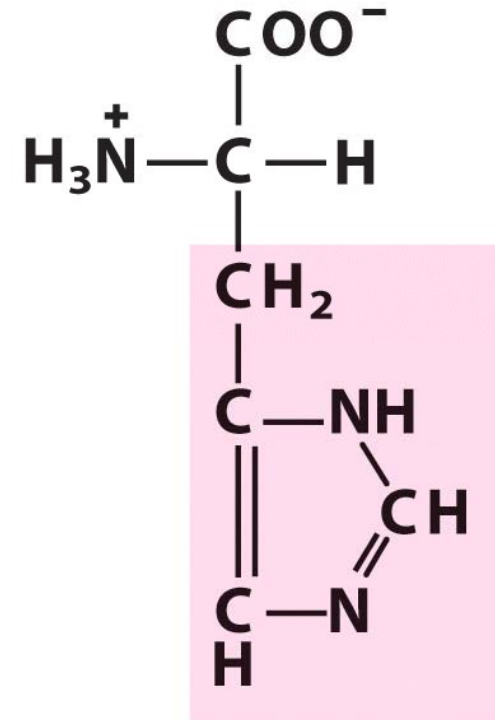
Positively charged R groups



Lysine

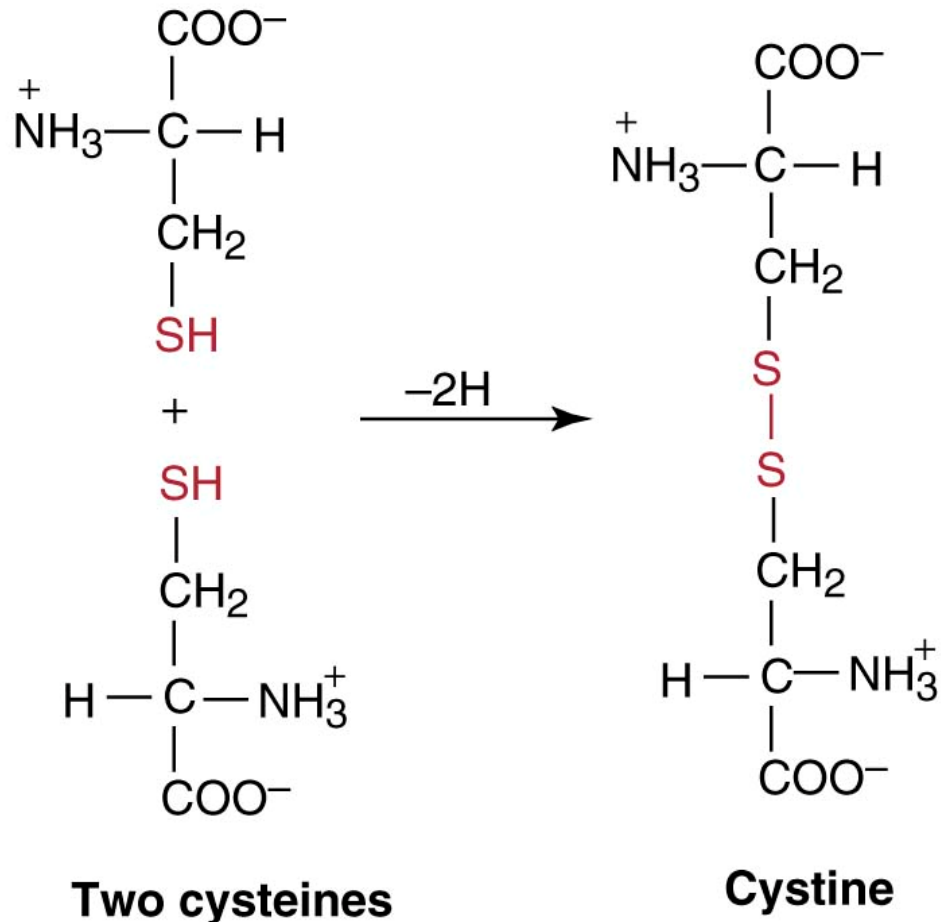


Arginine



Histidine

Modified amino acids

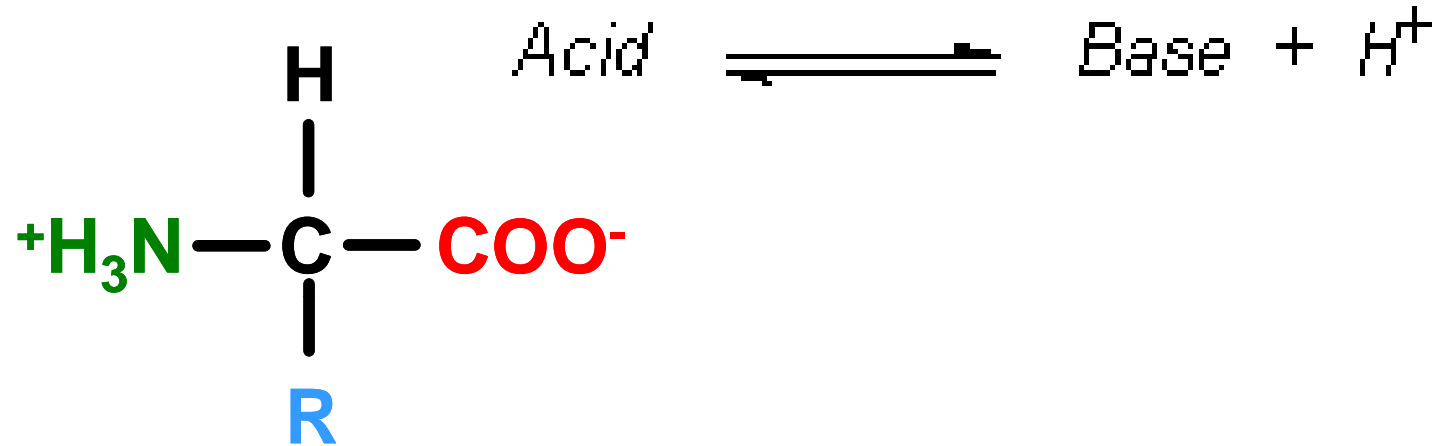


Cystinuria: incomplete reabsorption of cysteine during filtration in the kidney results in high cysteine and cystine in urine.

Excess cystine forms stones.

Figure 3.7. Cystine bond formation.

Acid- Base Properties



Acid- Base Properties

Zwitterion

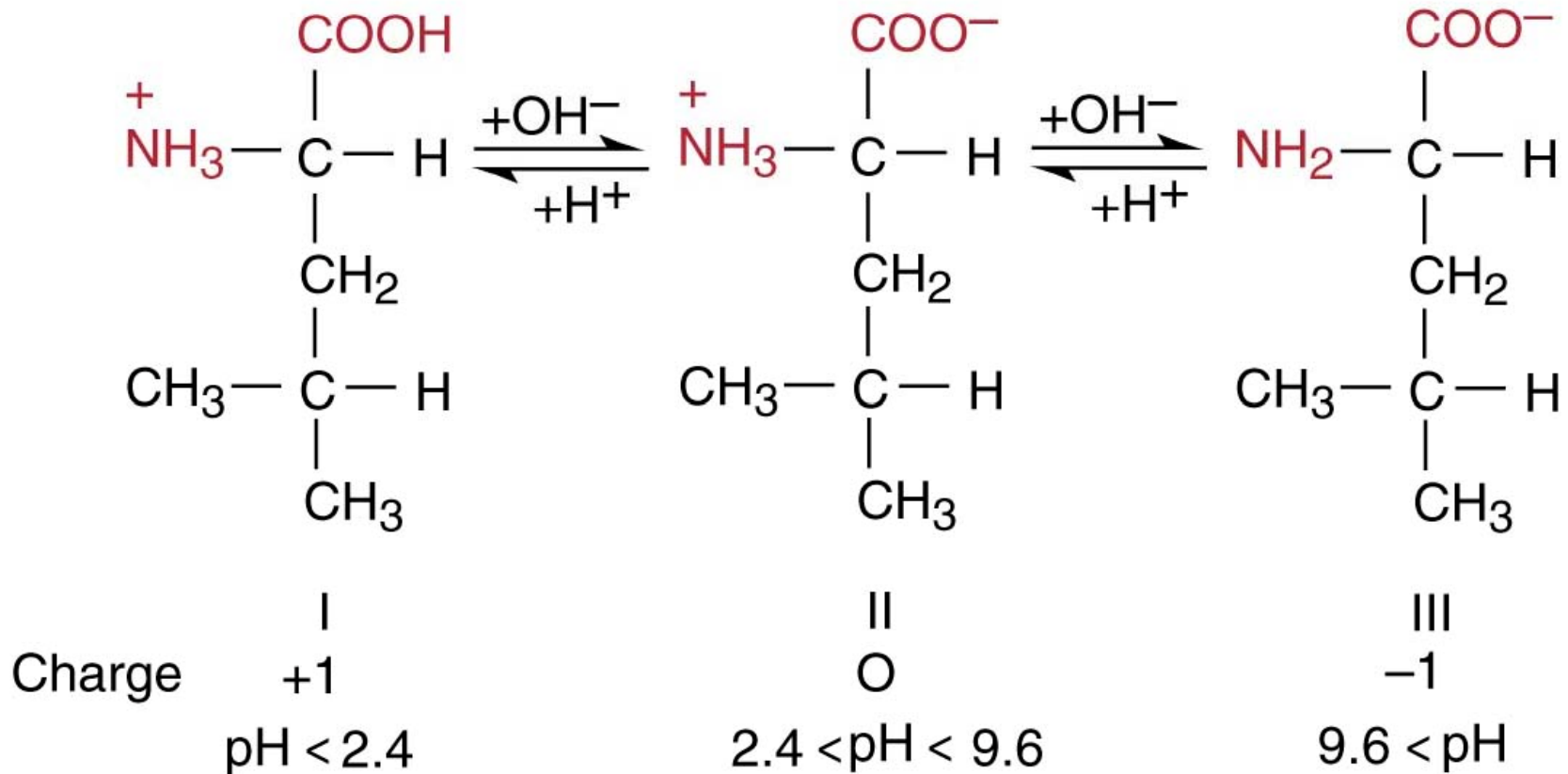


Figure 3.14. Ionic forms of leucine.

Acid- Base Properties

Henderson-Hasselbalch

$$\text{pH} = \text{pK}_a + \log \frac{[\text{conjugate base}]}{[\text{conjugate acid}]}$$

- $\text{pK}_a = \text{pH}$ acid is 50% deprotonated
- isoelectric point- (pI)
 - ✓ pH where the net charge of a molecule equals zero

$$\text{pI} = \frac{\text{pK}_a \text{COOH} + \text{pK}_a \text{NH}_3^+}{2}$$

Acid-Base Properties

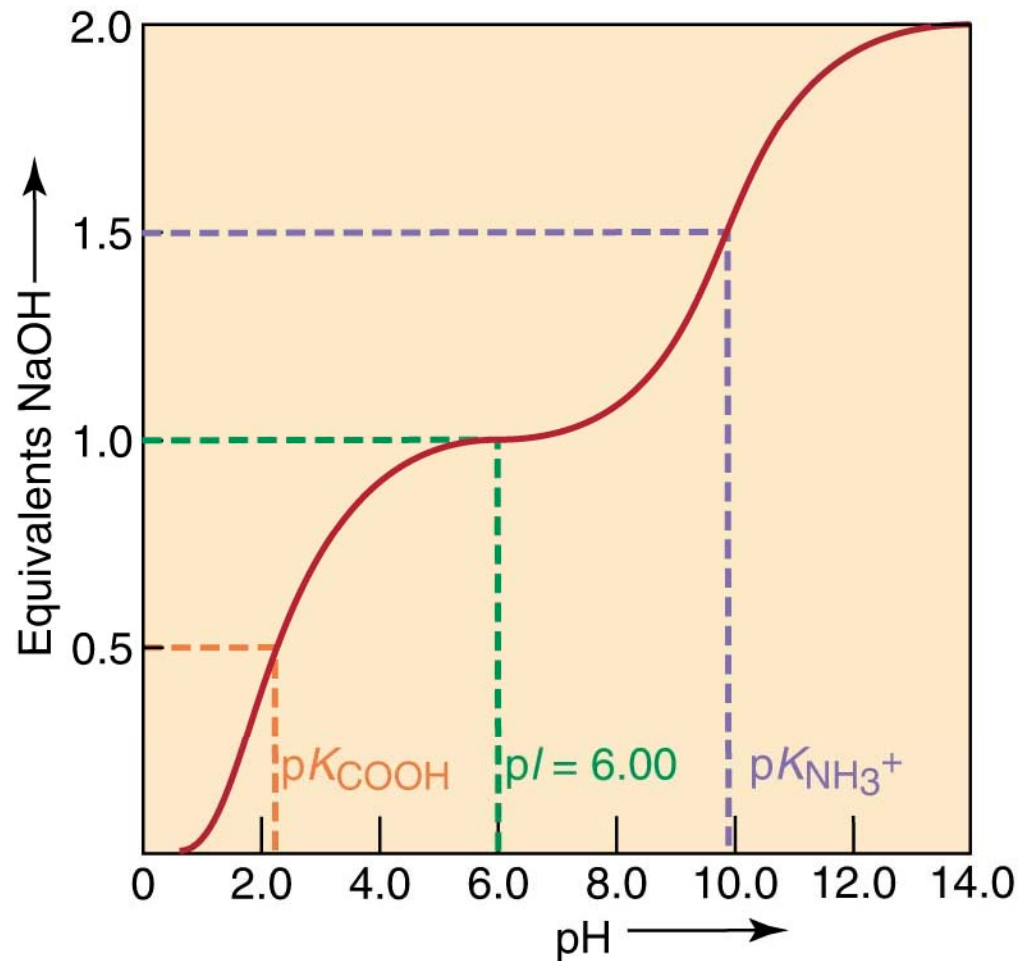
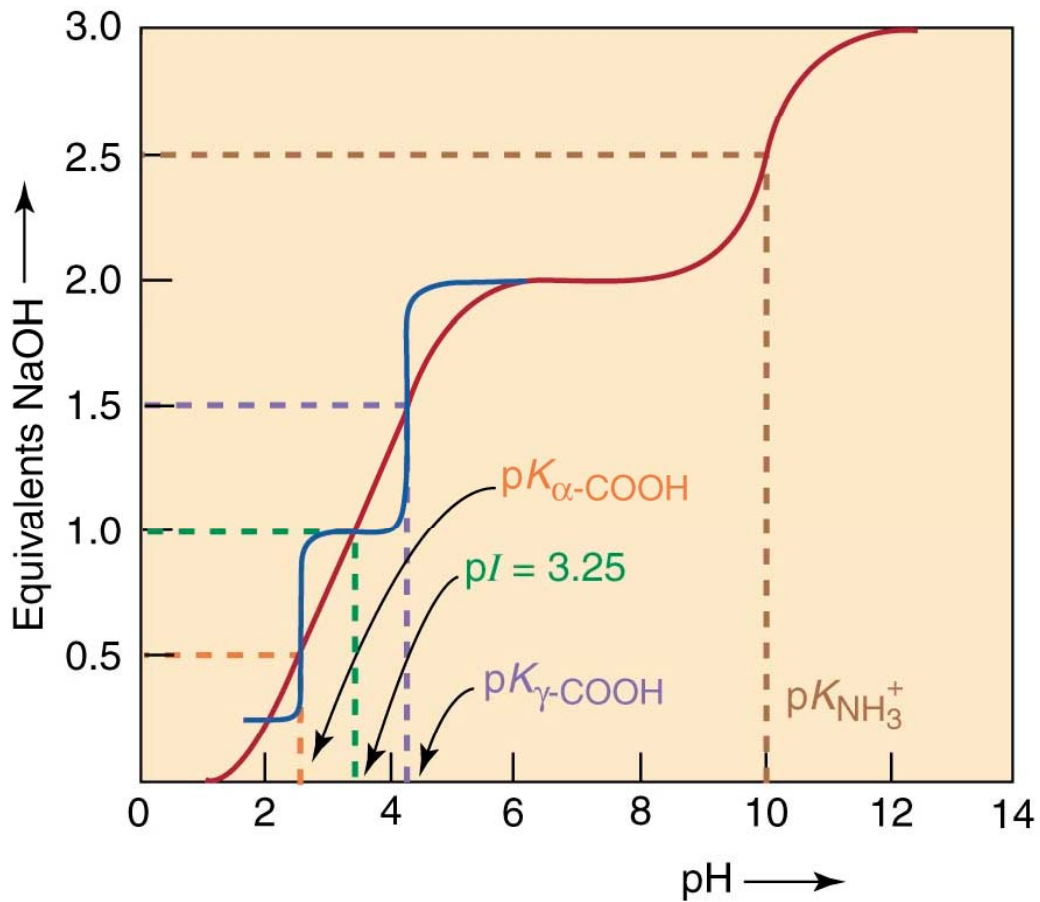
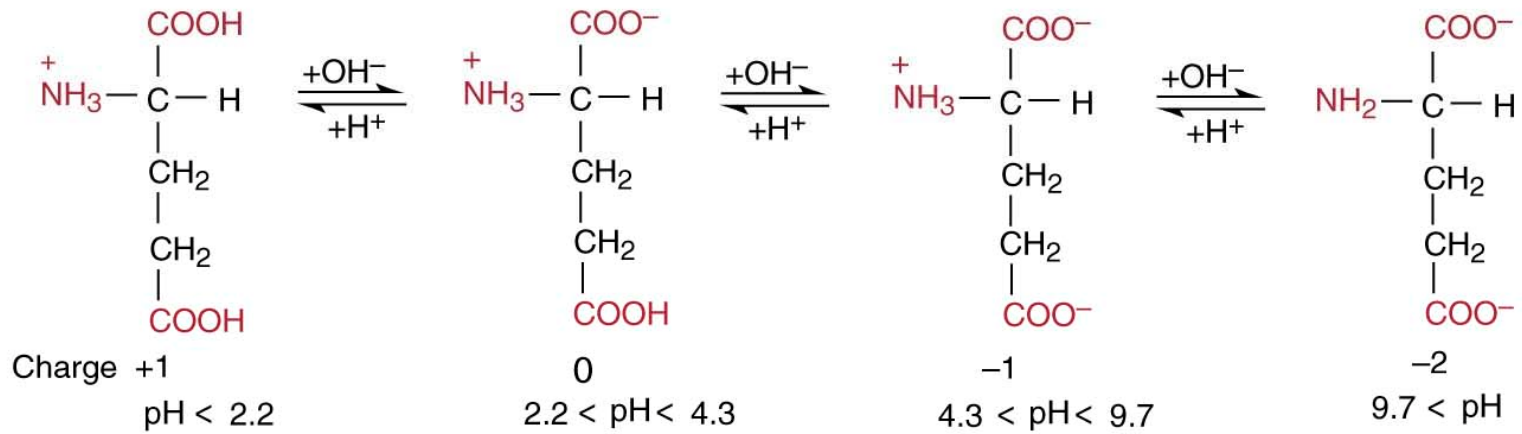


Figure 3.15. Titration curve of leucine.



Glutamic Acid

Acid-Base Properties

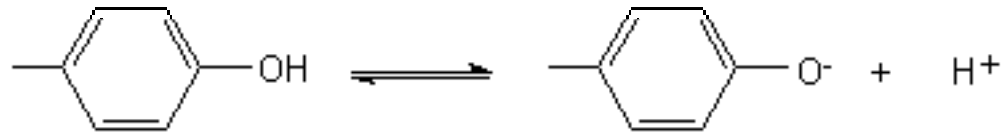
Aspartate / Glutamate



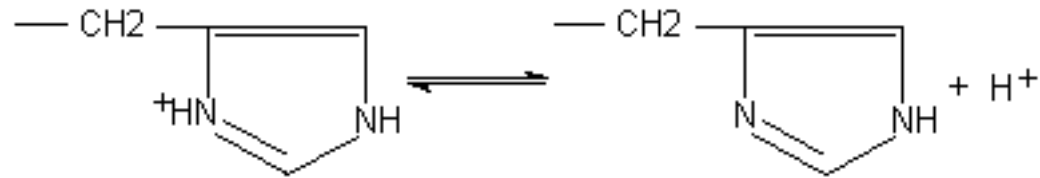
Cysteine



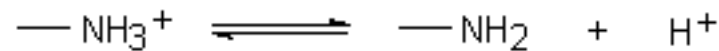
Tyrosine



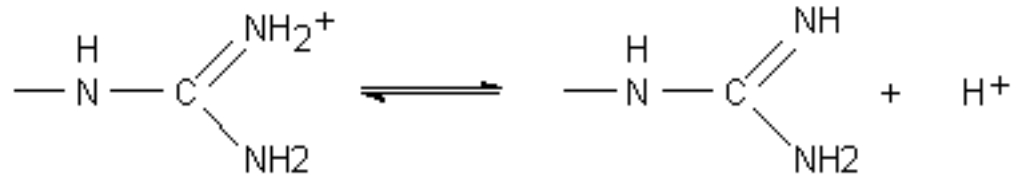
Histidine



Lysine

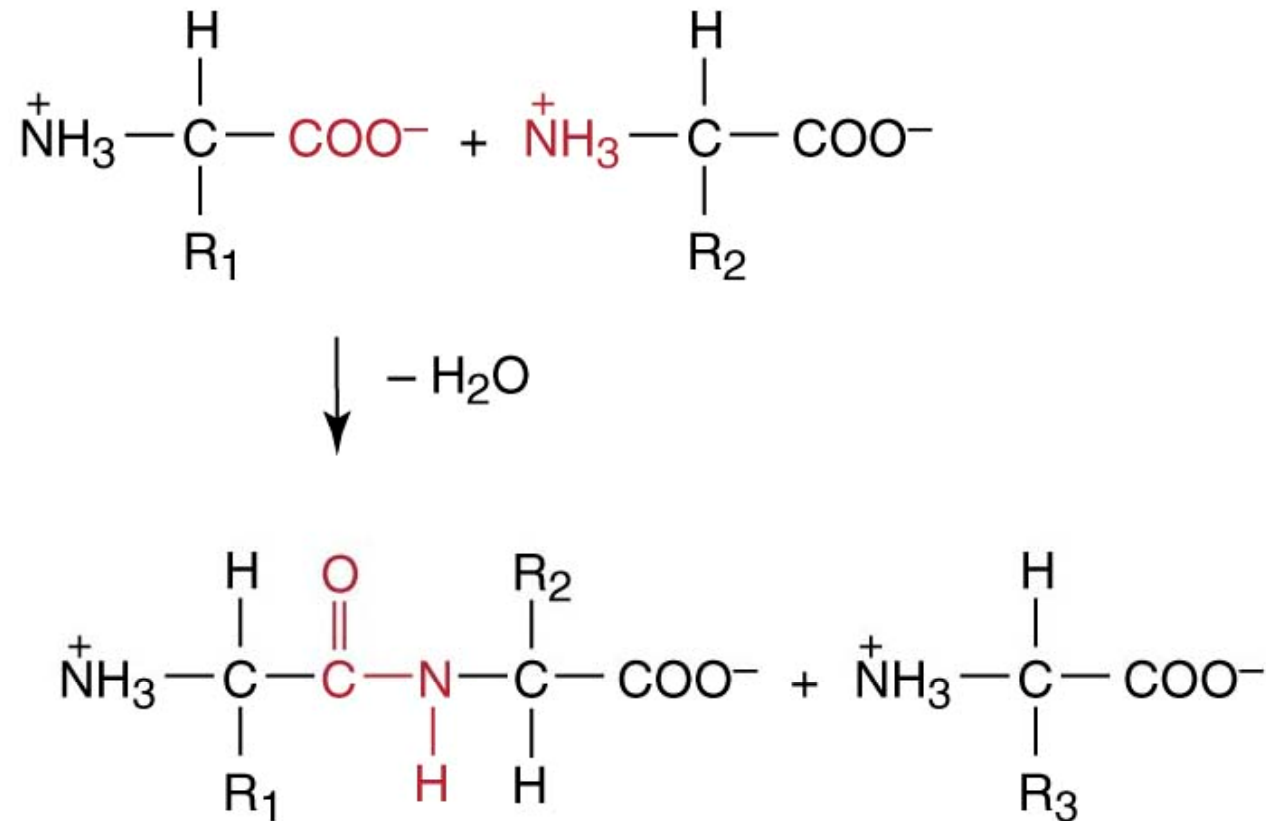


Arginine



Devlin:
Table 3.3

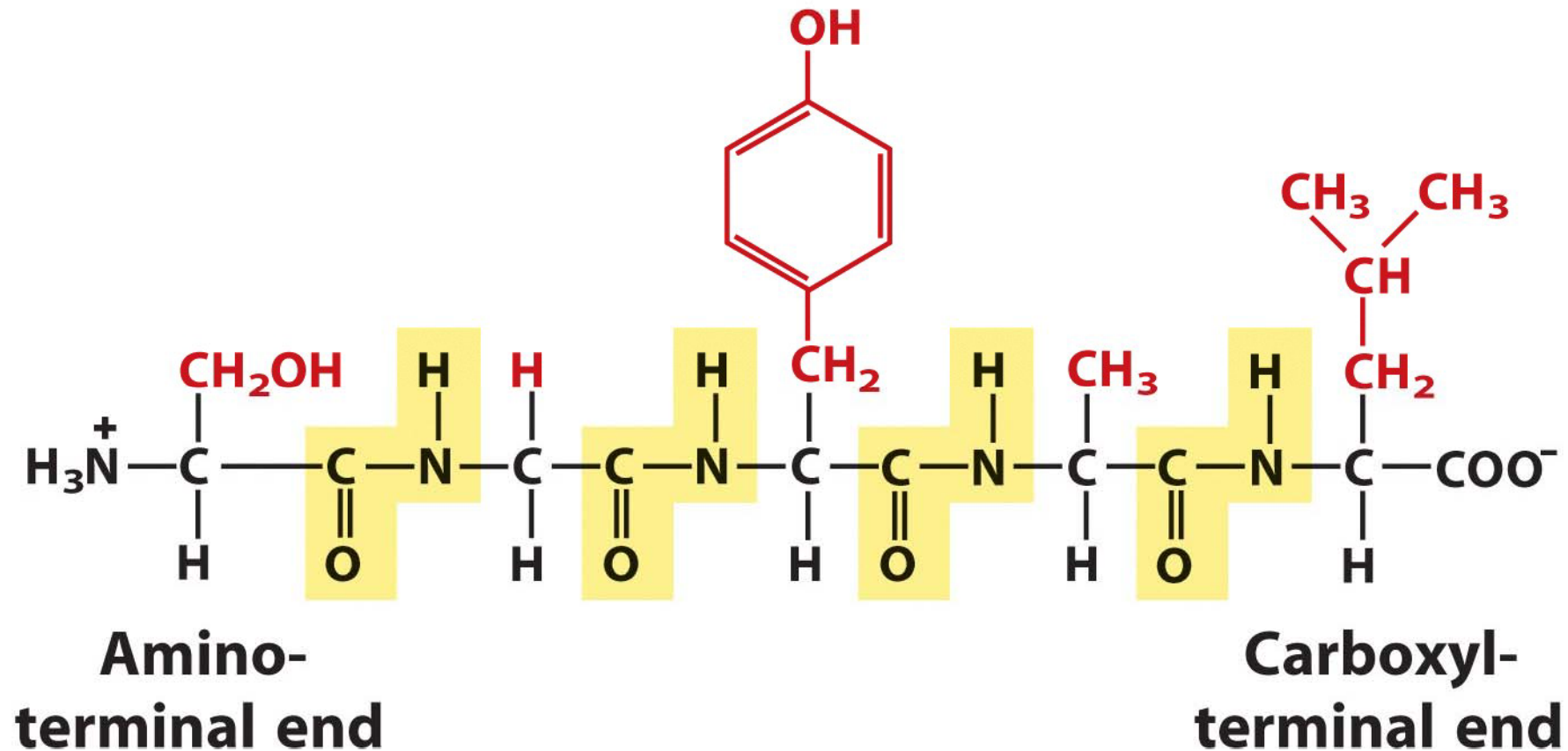
Peptide bond



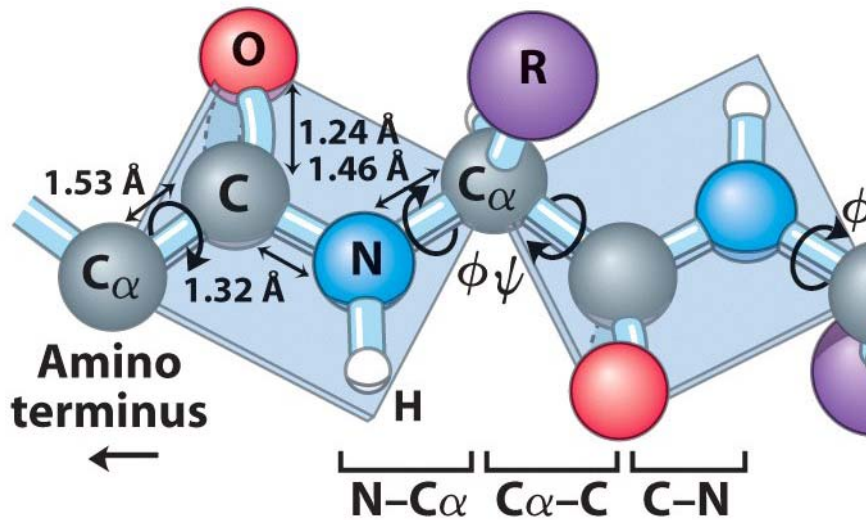
Dipeptide

Peptide bond

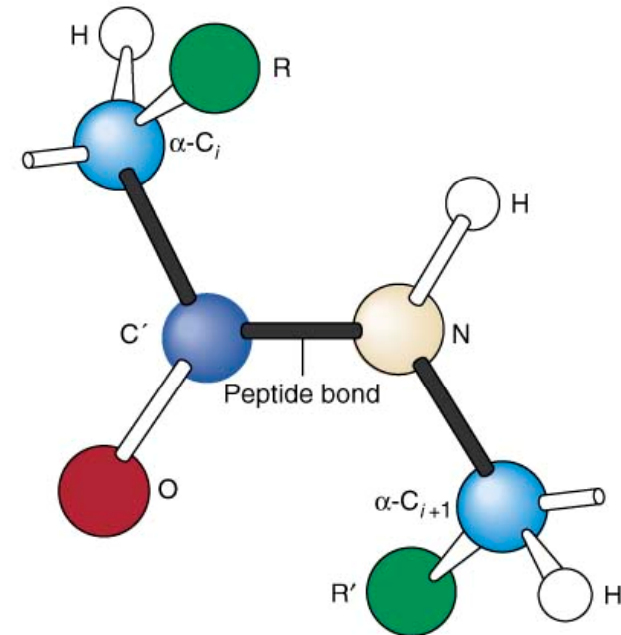
serylglycyltyrosylalanylleucine



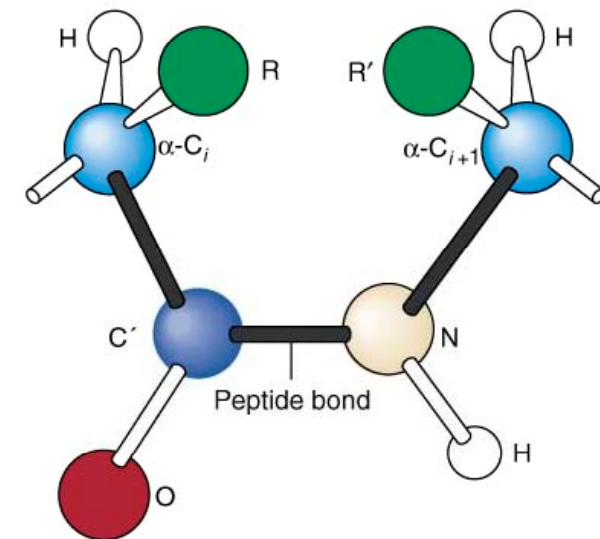
Peptide bond



- Partial double bond character
- Planar
- trans- configuration

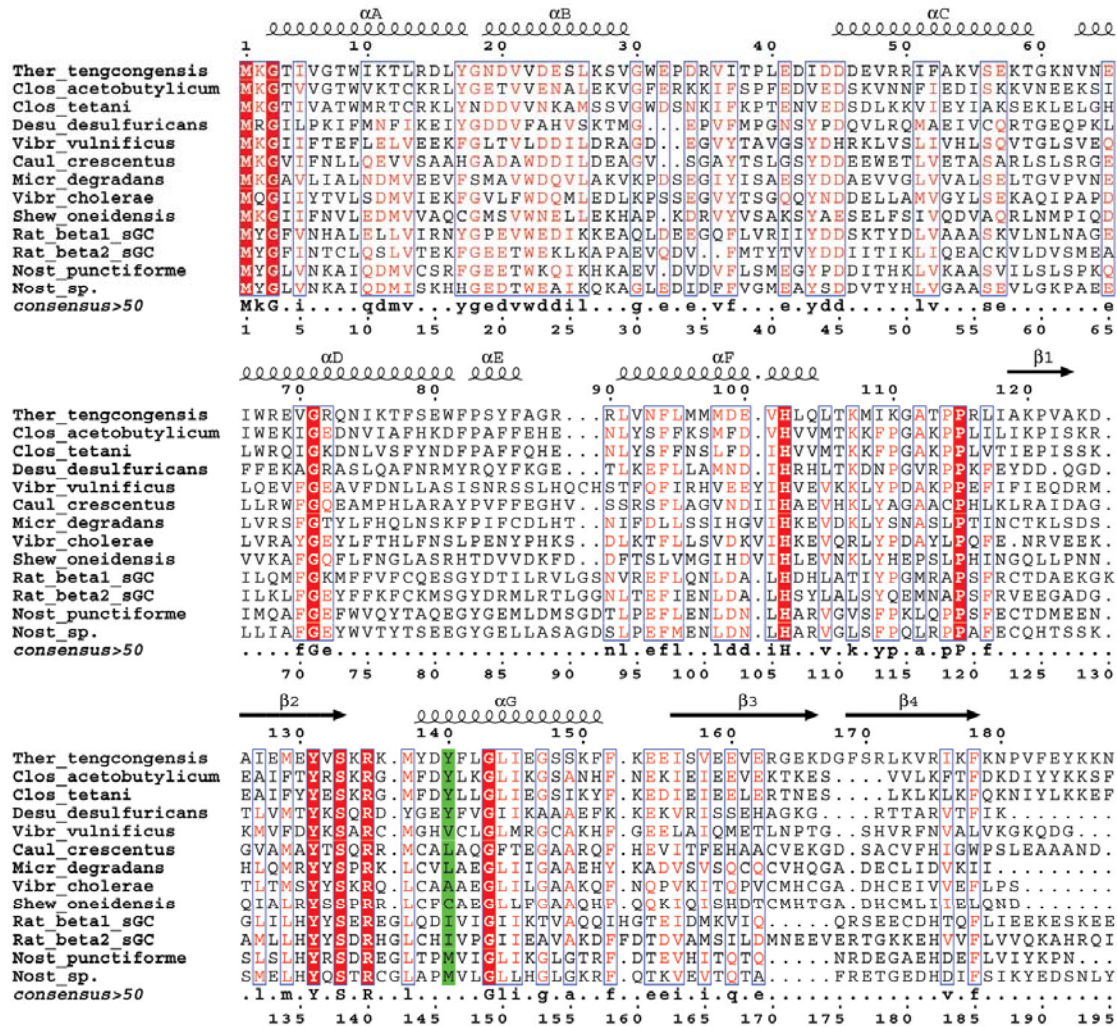


(a) *trans* configuration



(b) *cis* configuration

Primary Structure Sequence Alignment



- Analogy – seq. that are structurally similar but no evolutionary relationship has been demonstrated.
- Homology (homologous proteins) – aa sequences are highly alignable (proteins belong to the same family) – evolve from same gene and have similar functions
 - ✓ Paralog – two proteins within a family are present in the same species.
 - ✓ Ortholog – homologs from different species

Human and chimp myoglobin are ~100 % Identical

```

          10      20      30      40      50      60
Chimpa MGLSDGEWQLVLNVWGKVEADIPGHGQEVLRIRLFGKHPETLEKFDKFKHLKSEDEMKASE
.....
Human  MGLSDGEWQLVLNVWGKVEADIPGHGQEVLRIRLFGKHPETLEKFDKFKHLKSEDEMKASE
          10      20      30      40      50      60

          70      80      90      100     110     120
Chimpa DLKKHGATVLTALGGILKKKGHHEAEIKPLAQSHATKHKIPVKYLEFISECIIQVLHSKH
.....
Human  DLKKHGATVLTALGGILKKKGHHEAEIKPLAQSHATKHKIPVKYLEFISECIIQVLQSKH
          70      80      90      100     110     120

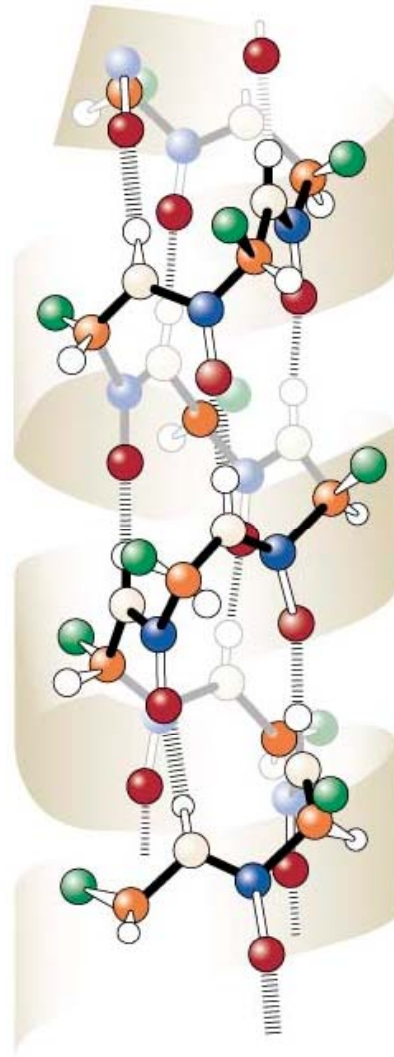
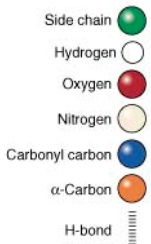
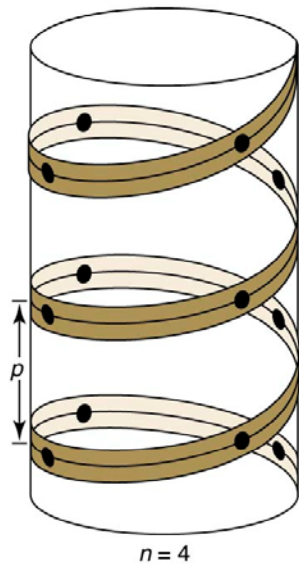
          130     140     150
Chimpa PGDFGADAQGAMNKALELFRKDMASNYKELGFQG
.....
Human  PGDFGADAQGAMNKALELFRKDMASNYKELGFQG
          130     140     150
  
```

95% of their DNA sequence, and 99% of coding DNA sequences are in common

Secondary Structure

- Refers to local conformation of some part of the polypeptide
- Due to the partial double character of the peptide bond
- Stabilized mainly by H-bonds
- α -helix
- β -sheet
- β -turn

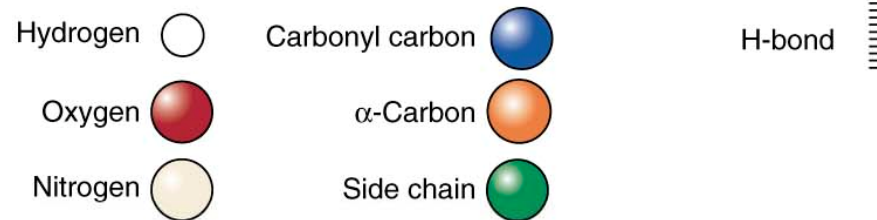
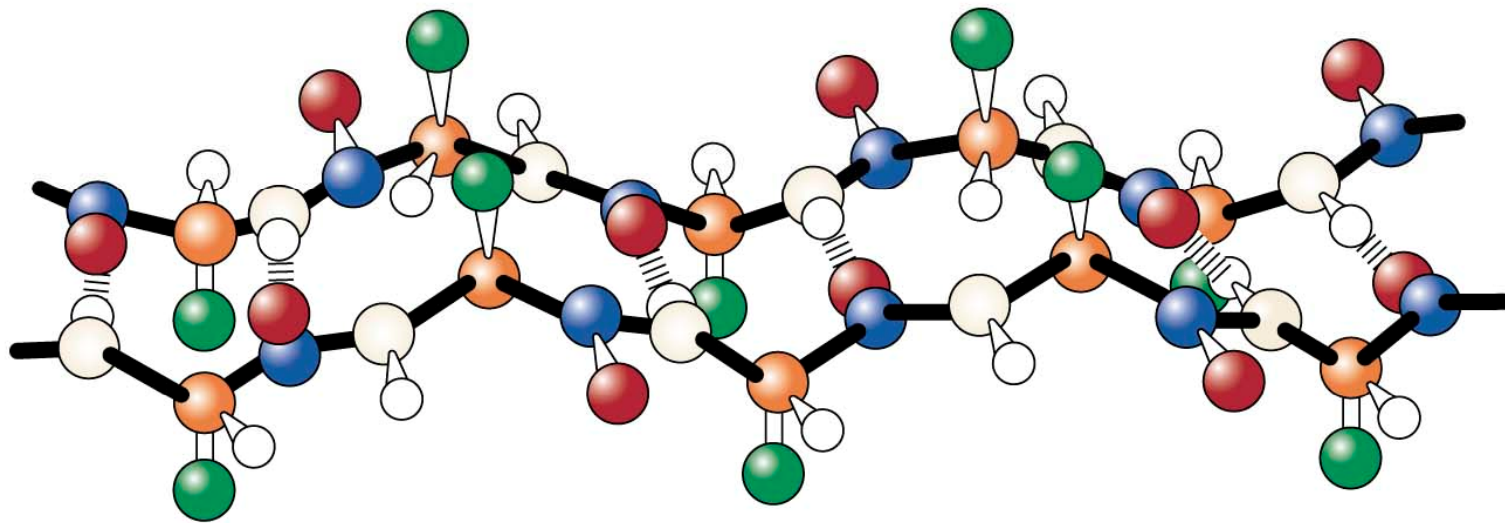
Secondary Structure



- alpha - helix
 - ✓ 3.6 aa per turn
 - ✓ Stabilized by H-bonds
 - ✓ Right handed

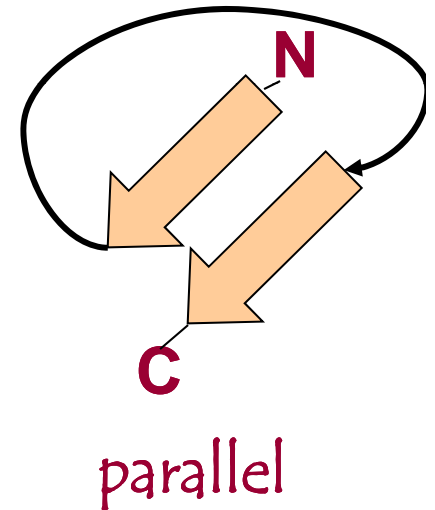
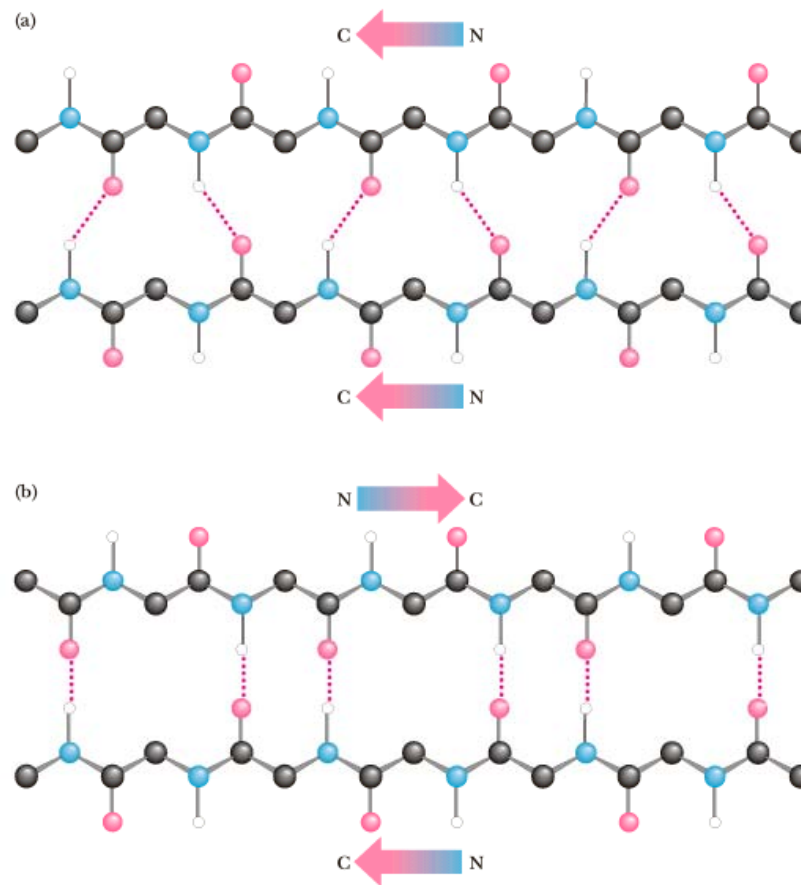
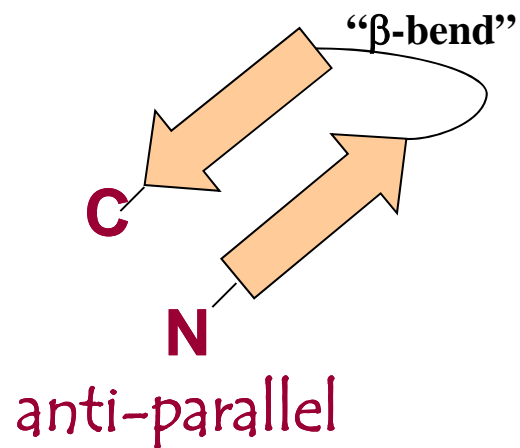
Secondary Structure

- β -pleated sheets



Secondary Structure

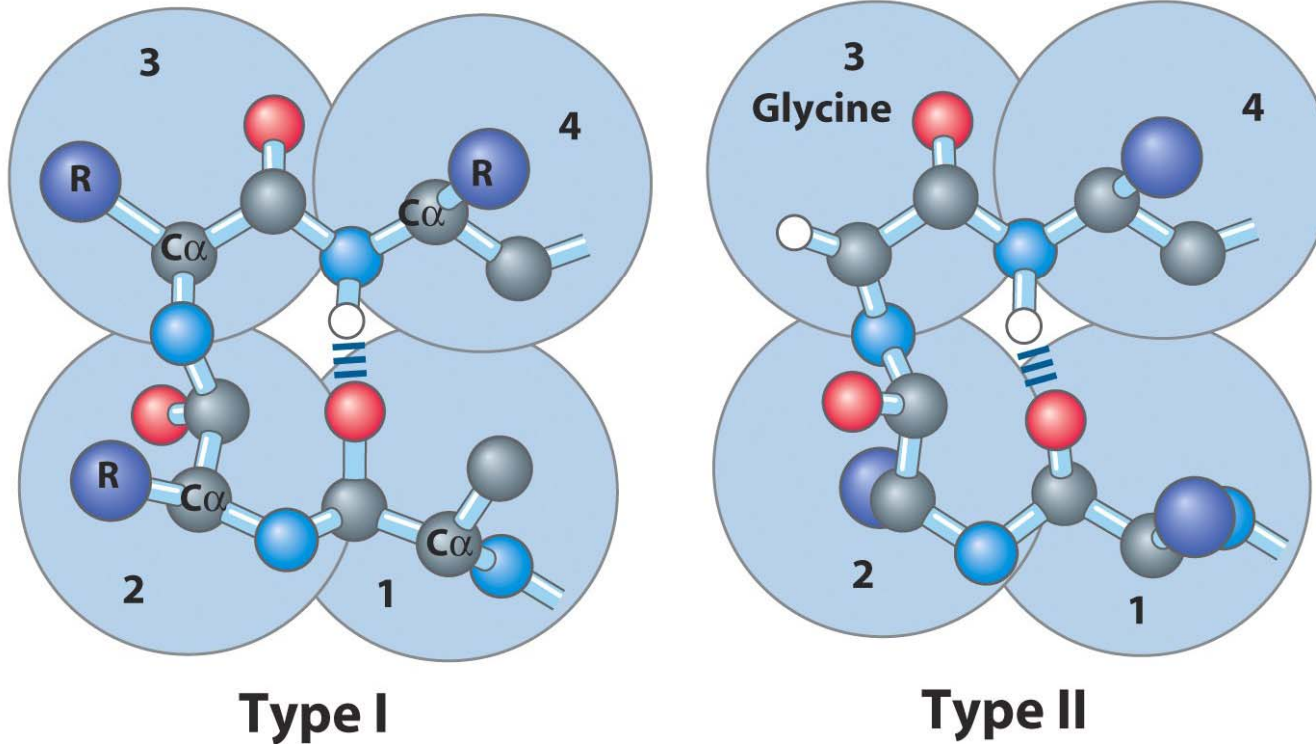
- β -pleated sheets



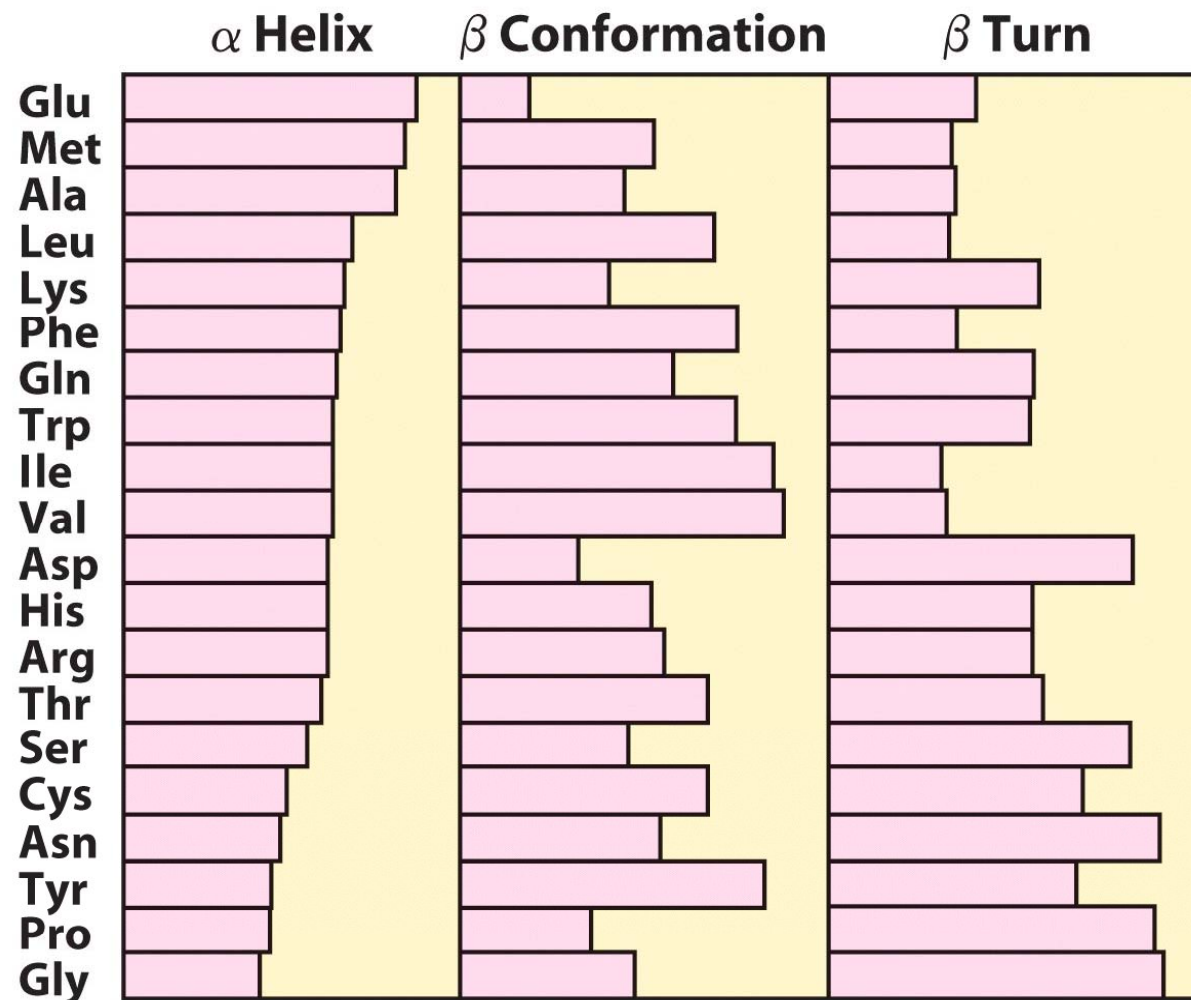
Secondary Structure

- Turns / Loops
 - ✓ Gly & Pro

(a) β Turns



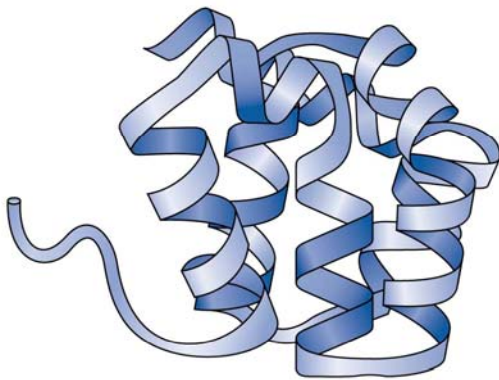
Amino acids occurrence in secondary structures



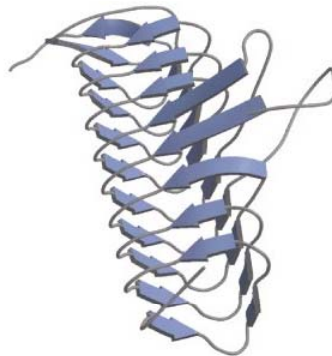
Ultrasecondary Structures

- Motifs / Folds – refers to particularly stable arrangements of several elements of secondary structure and the connections between them.
- Structural Classification

All α

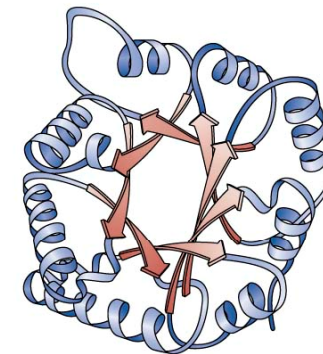


All β

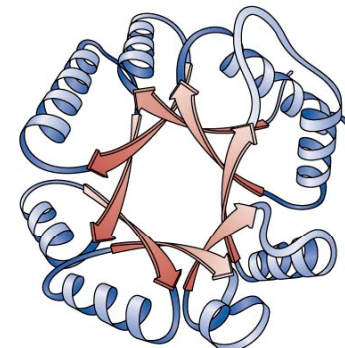


1LXA
Single-stranded left-handed β helix
Trimeric LpxA-like enzymes
UDP *N*-acetylglucosamine acyltransferase
UDP *N*-acetylglucosamine acyltransferase
Escherichia coli

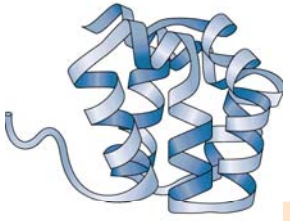
α, β



Triose Phosphate Isomerase

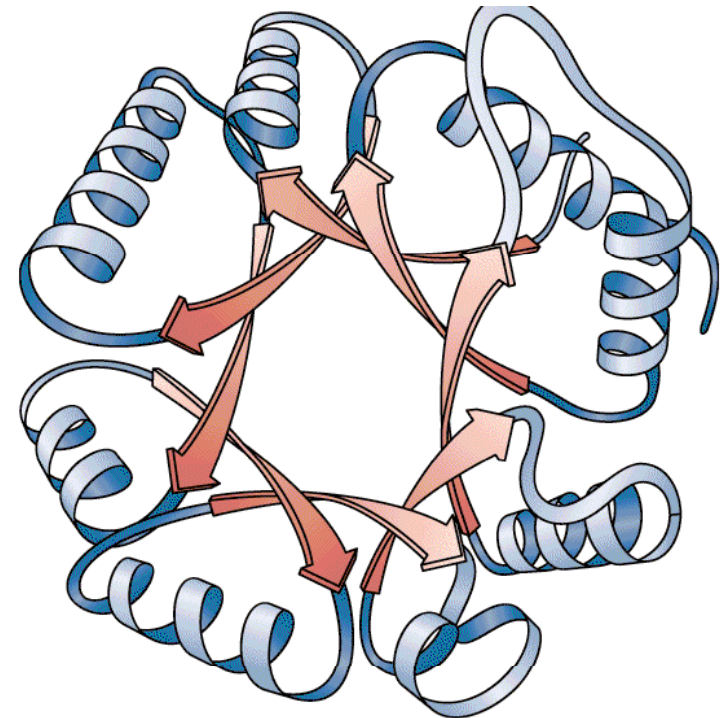


Pyruvate Kinase domain 1



Tertiary Structure

- Refers to 3D conformation (location of each atom in space)
- Stabilized by:
 - ✓ Disulfide bonds (covalente)
 - ✓ Non-covalent interactions
 - Hydrophobic
 - H-bonds
 - Ionic



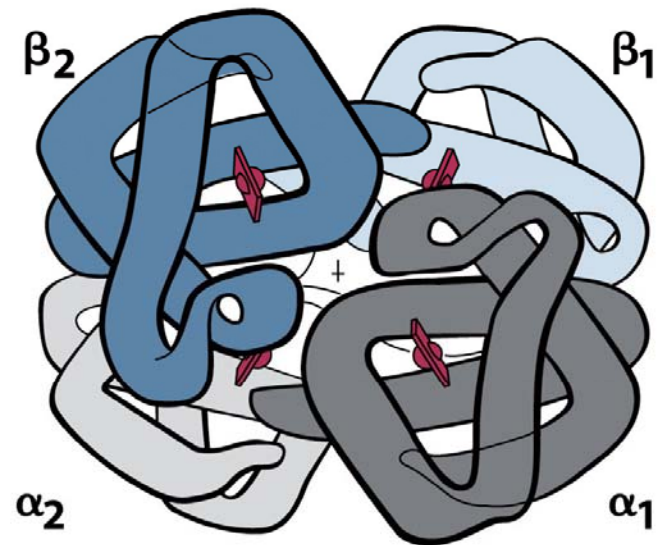
Pyruvate Kinase domain 1

Quaternary Structure

- Non-covalent assemblies of two or more monomer subunits.
- Sub-units may work independently or cooperatively

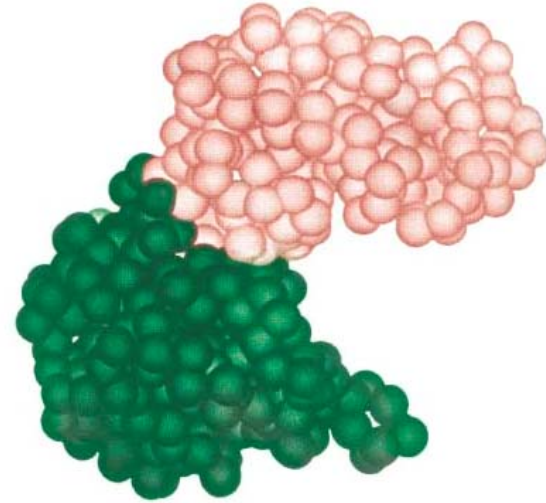


Collagen, a fibrous protein

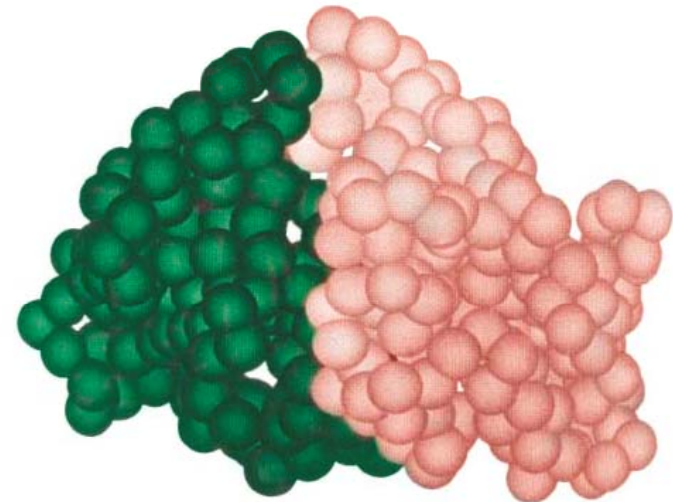


Protein Domains

- Domains = Globular units within proteins
 - ✓ Different domains have different functions
 - ✓ Small proteins usually have one domain

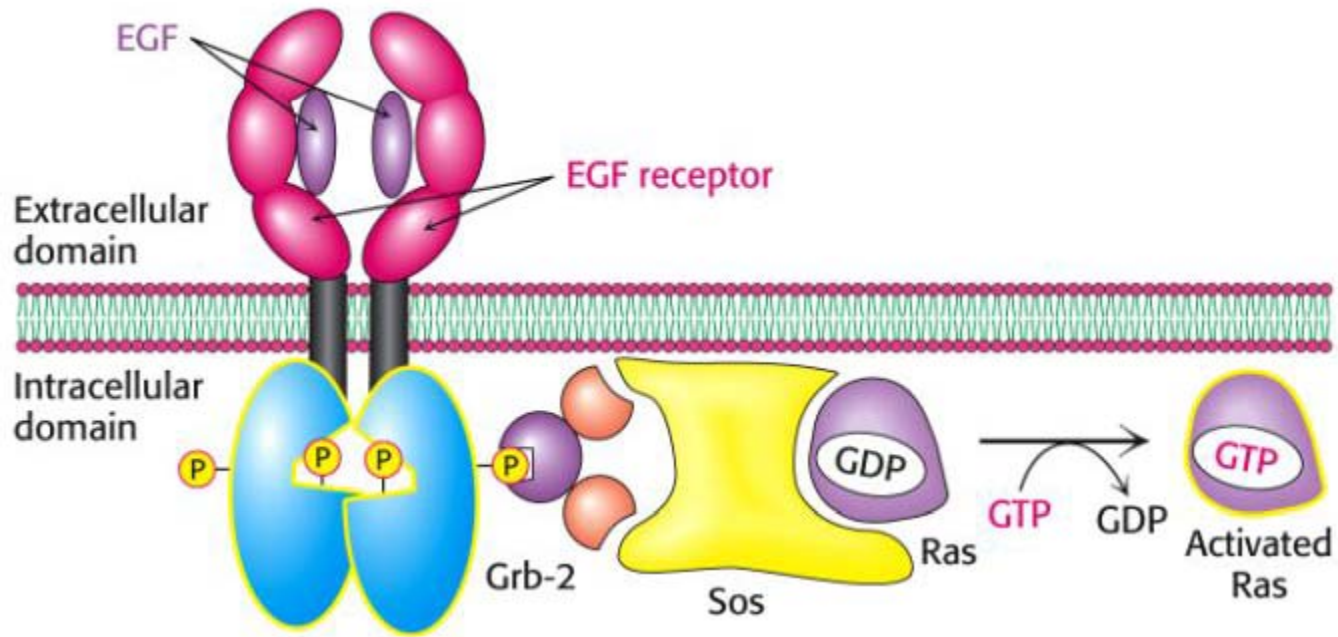


(a)

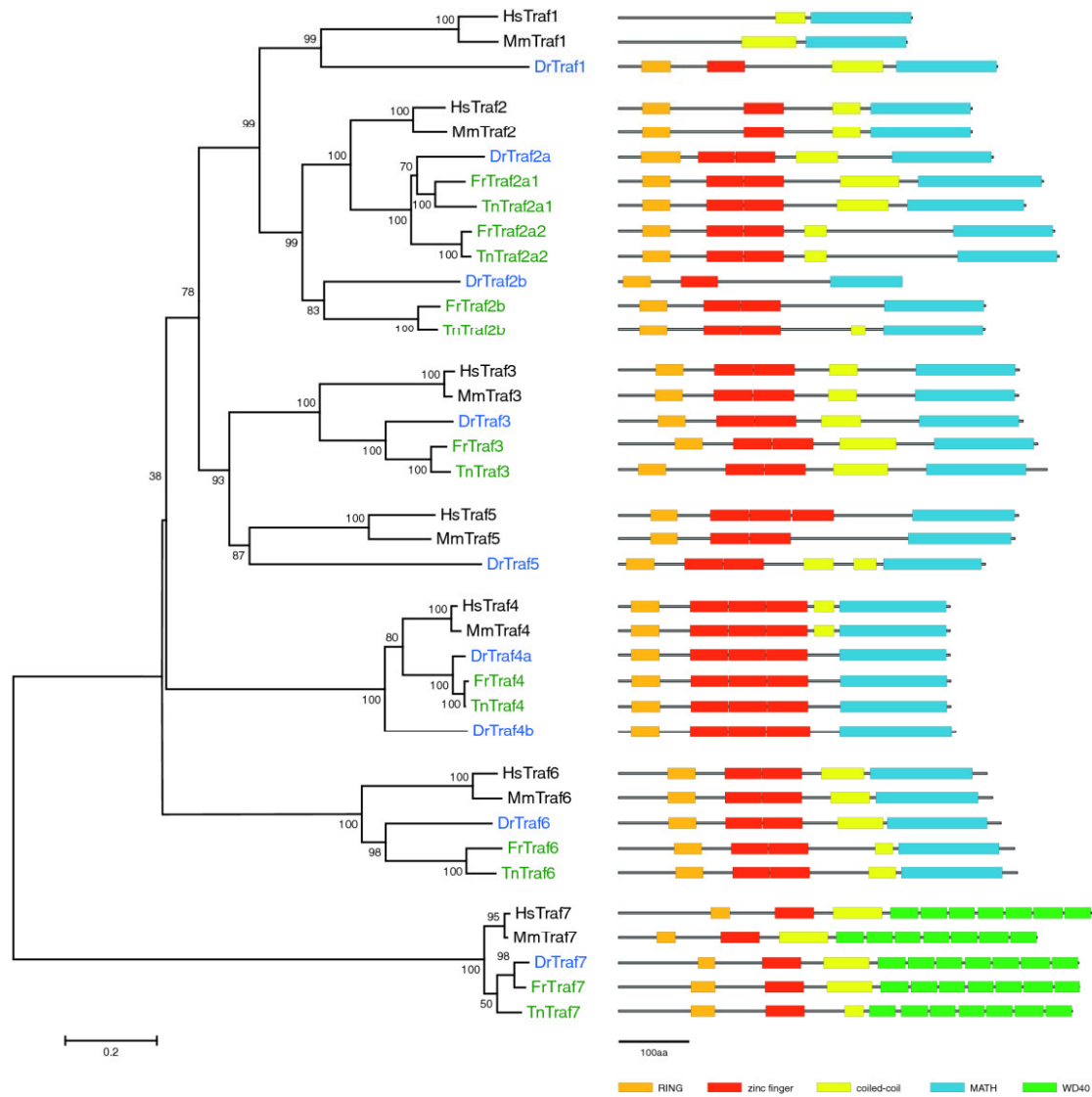


(b)

Domains

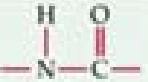

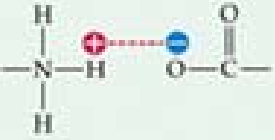




Domains



Forces that stabilize protein structure

2.1 Chemical Bonds and Interactions

NAME	BASIS OF INTERACTION	STRUCTURE	BOND ENERGY* (KCAL/MOL)
Covalent bond	Sharing of electron pairs		50–110
*Hydrogen bond	Sharing of H atom		3–7
Ionic interaction	Attraction of opposite charges		3–7
*van der Waals interaction	Interaction of electron clouds		1
Hydrophobic interaction	Interaction of nonpolar substances		1–2

*Bond energy is the amount of energy needed to separate two bonded or interacting atoms under physiological conditions.

Hydrogen bonds

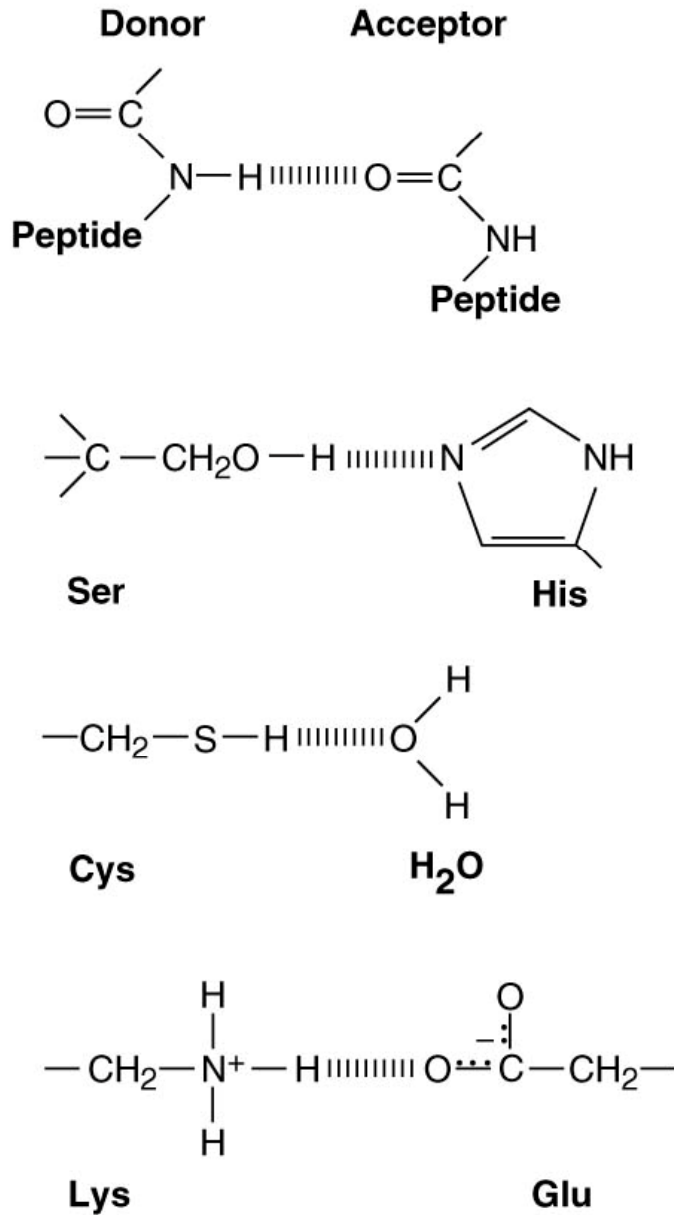
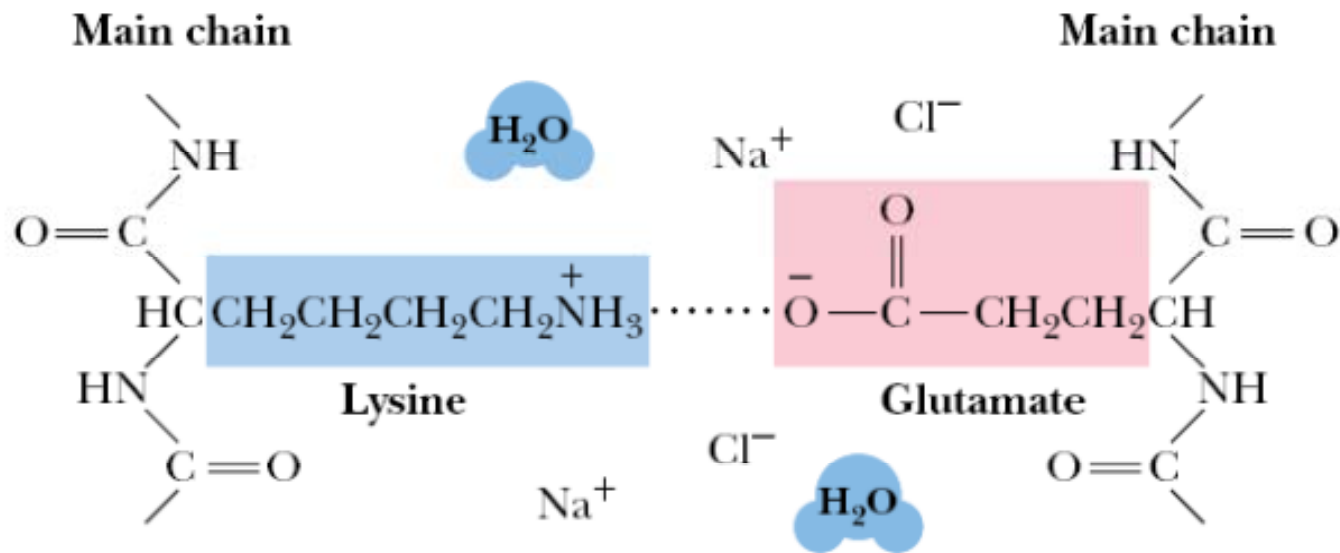
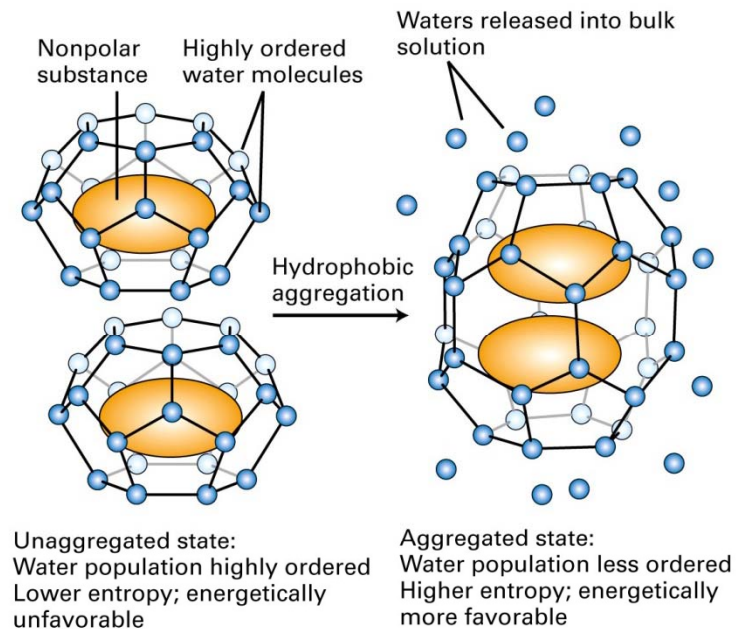
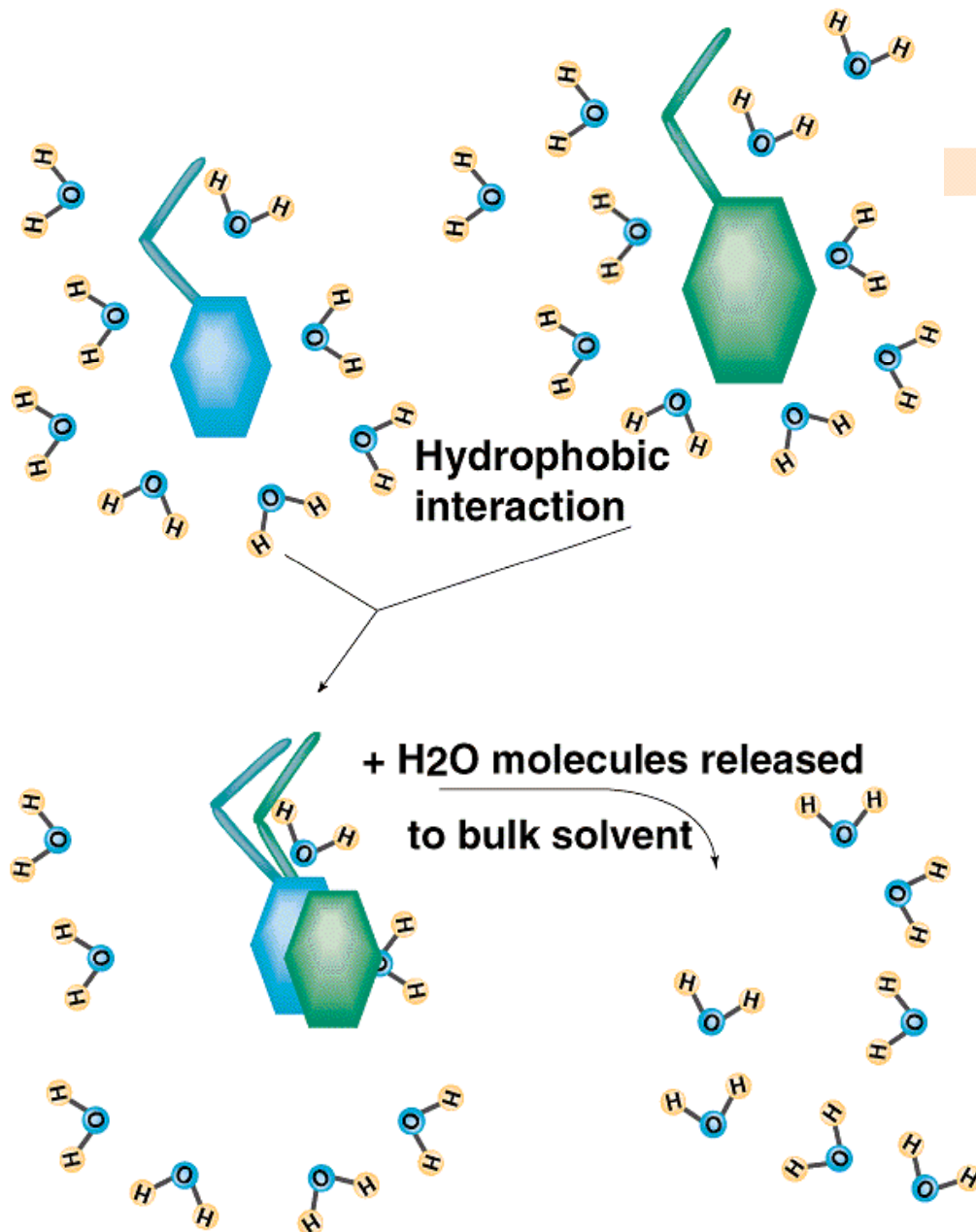


Figure 3.48. Some common hydrogen bonds found in proteins.

Ionic Interaction



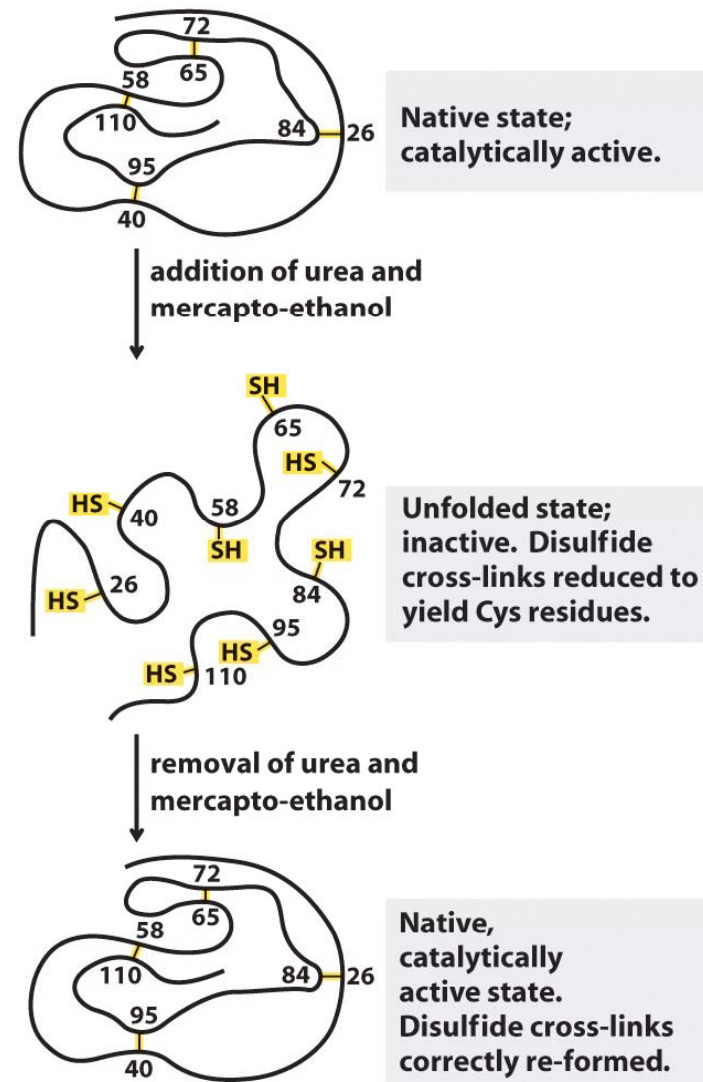
Hydrophobic Interactions



- Increases entropy of H₂O

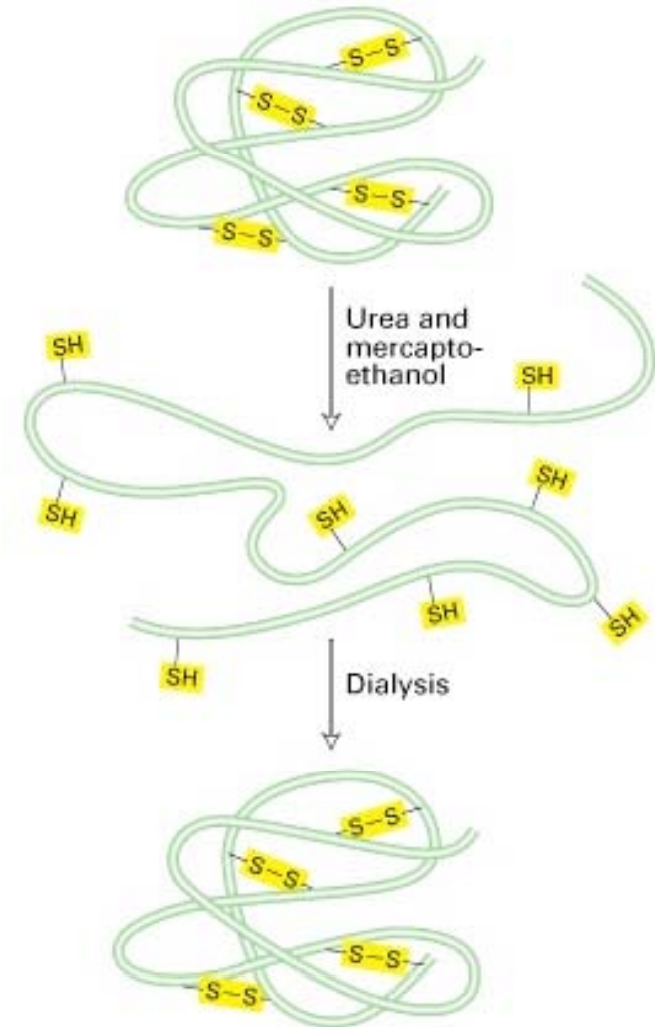
Protein Denaturation

- Anfisen
 - ✓ Denatured ribonuclease using urea and β -ME
 - ✓ Removed denaturing agents – protein refolded into its native conformation
- Evidence that 3^o structure of a globular protein is determined by its aa sequence



Protein Denaturation

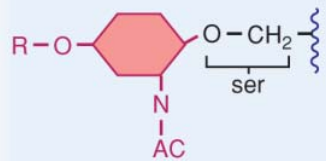
- Temperature (heat)
 - ✓ Affects weak interaction - hydrophobic
 - pH (Acids and Bases)
 - ✓ Alters net charge, cause electrostatic repulsion and disruption of H bonds
 - Organic Solvents
 - Detergents
 - Urea
 - Heavy metal ions
 - ✓ Lead
 - ✓ Mercury
- } Disrupt hydrophobic interactions



Post-translational modifications

Carbohydrate addition

O-glycosylation: OH of ser, thr, tyr,

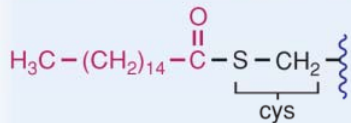


N-glycosylation: NH₂ of asn

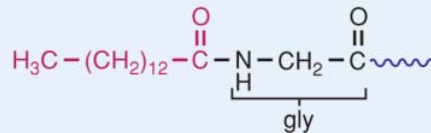


Lipid addition

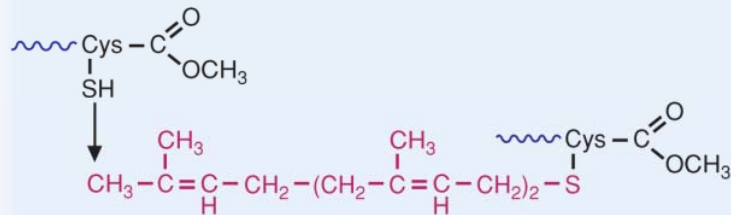
Palmitoylation: Internal SH of cys



Myristoylation: NH of N-terminal gly

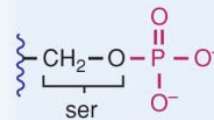


Prenylation: SH of cys

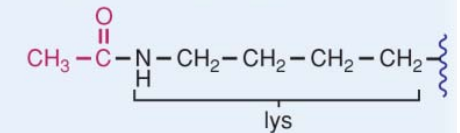


Regulation

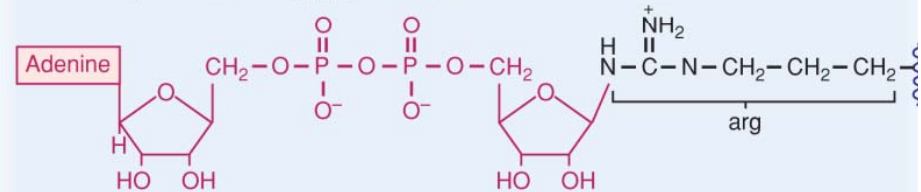
Phosphorylation: OH of ser, thr, tyr



Acetylation: NH₂ of lys, terminus

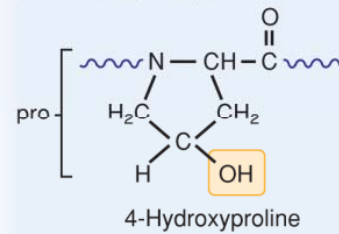


ADP-ribosylation: N of arg, gln; S of cys

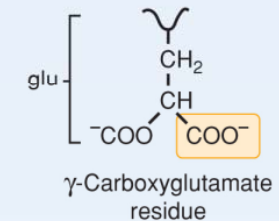


Modified amino acids

Oxidation: pro, lys

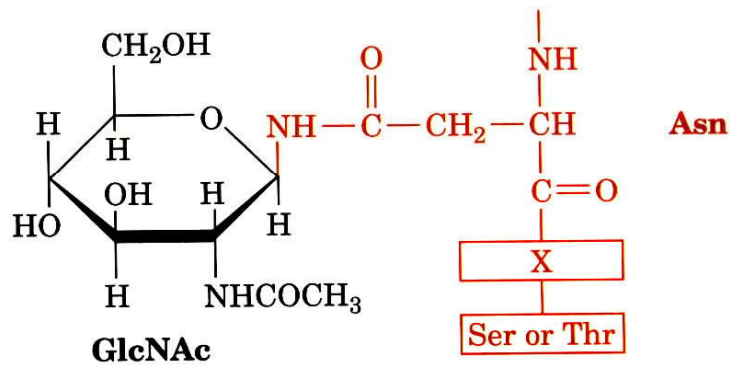


Carboxylation: glu

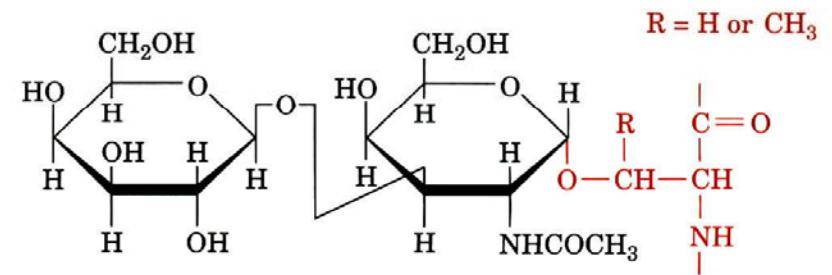


Post-translational modifications

- Glycosylation
 - ✓ Protection against proteases



N-linked

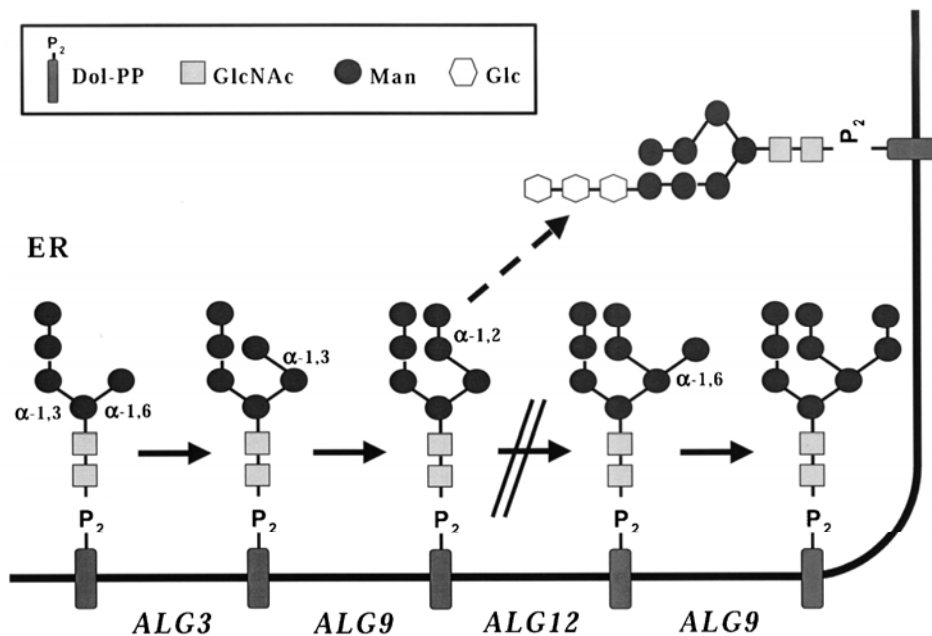


O-linked

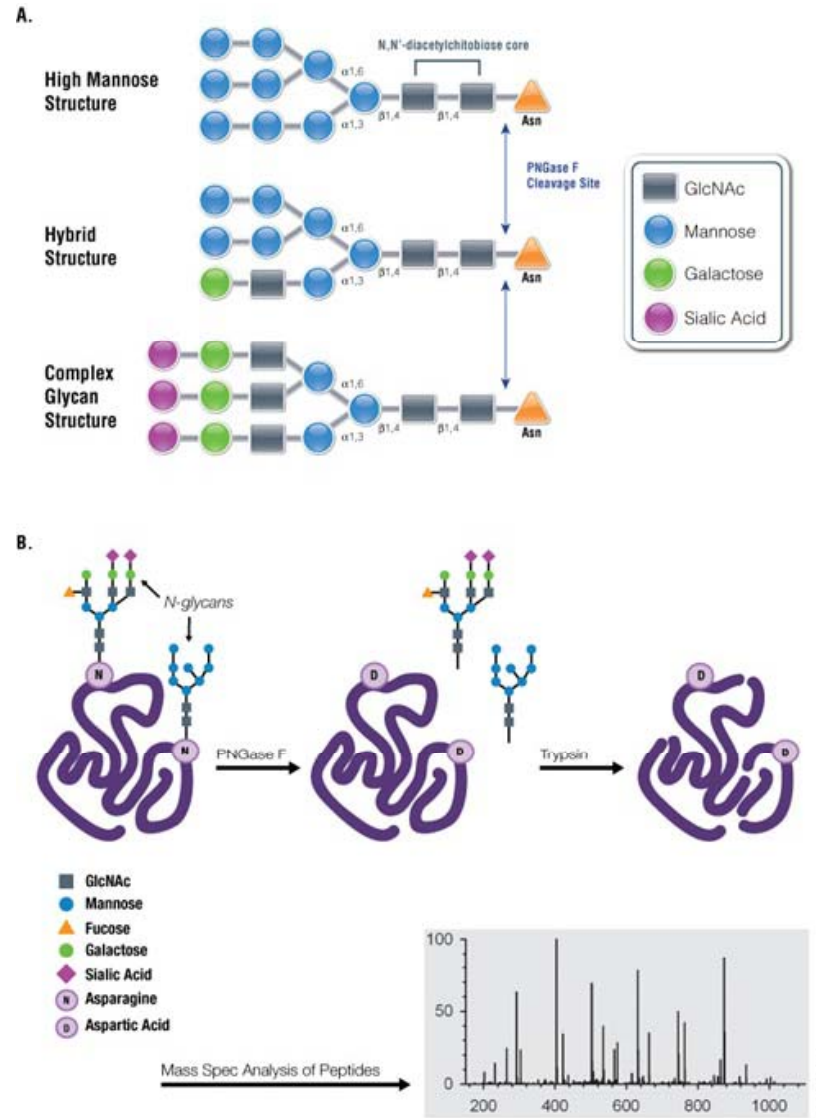
Post-translational modifications

■ Glycosilation

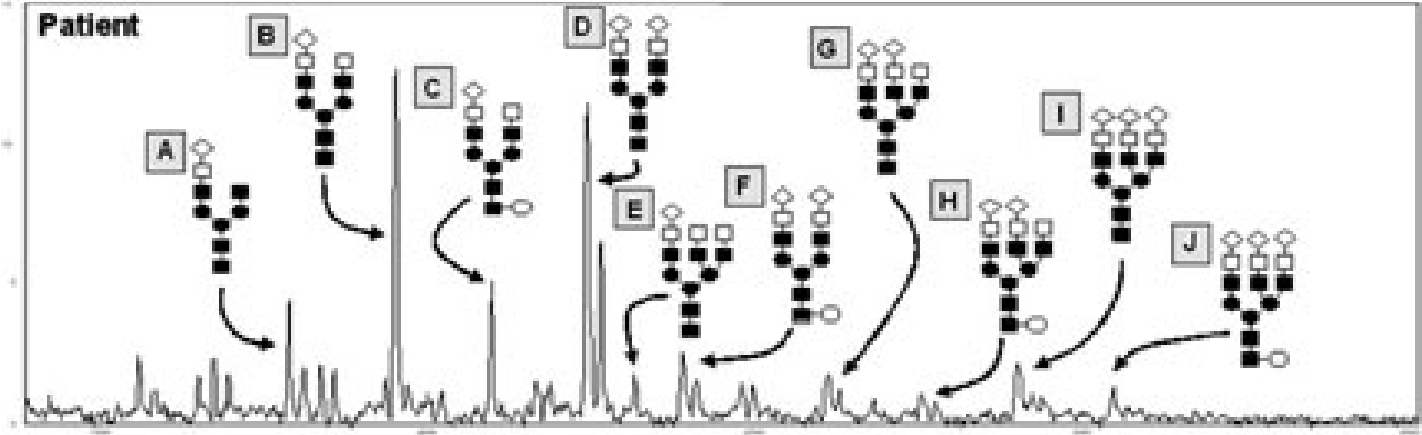
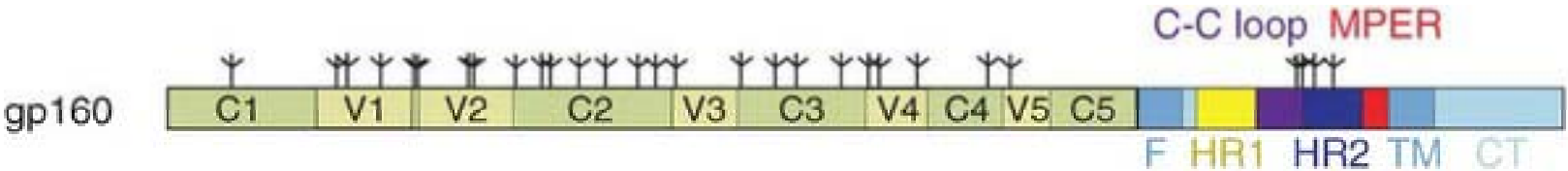
- ✓ Protection against proteases
- ✓ For the detection of unfolded proteins in cells
- ✓ Changes in glycosylation patterns in malignant cells
- ✓ Increased branching pattern associated with malignancy



Glycosylation of the HIV Env Protein

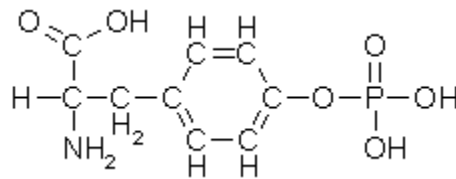


Glycosylation of the HIV Env Protein



Post-translational modifications

- Phosphorylation - regulation
 - ✓ Serine, threonine, tyrosine



Conjugated Proteins

- Apoprotein + prosthetic group = Prot conjugated

TABLE 3-4 Conjugated Proteins

<i>Class</i>	<i>Prosthetic group</i>	<i>Example</i>
Lipoproteins	Lipids	β_1 -Lipoprotein of blood
Glycoproteins	Carbohydrates	Immunoglobulin G
Phosphoproteins	Phosphate groups	Casein of milk
Hemoproteins	Heme (iron porphyrin)	Hemoglobin
Flavoproteins	Flavin nucleotides	Succinate dehydrogenase
Metalloproteins	Iron	Ferritin
	Zinc	Alcohol dehydrogenase
	Calcium	Calmodulin
	Molybdenum	Dinitrogenase
	Copper	Plastocyanin

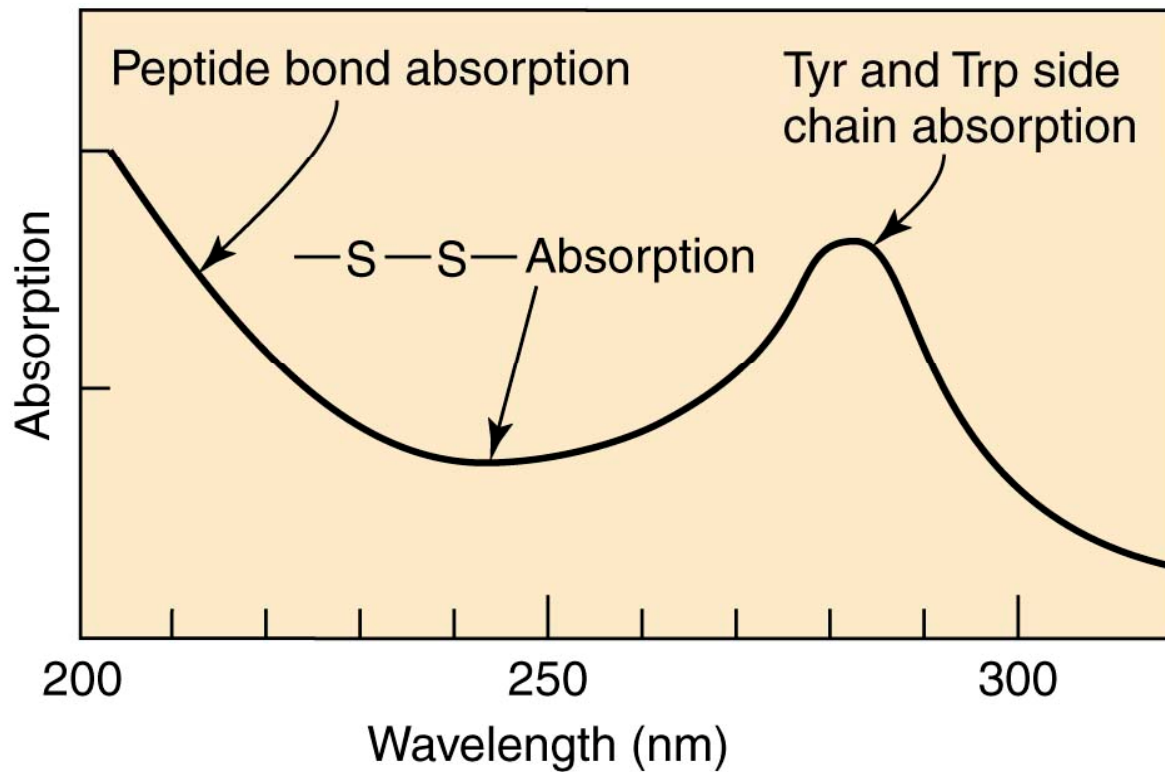
Chemical Properties of Proteins



- UV Light
- Fluorescence
- Optical Rotatory Dispersion & Circular Dichroism

Spectroscopic Properties

■ UV



Spectroscopic Properties

■ Fluorescence

- ✓ Most relevant = Trp
- ✓ Used to study structural and conformational changes

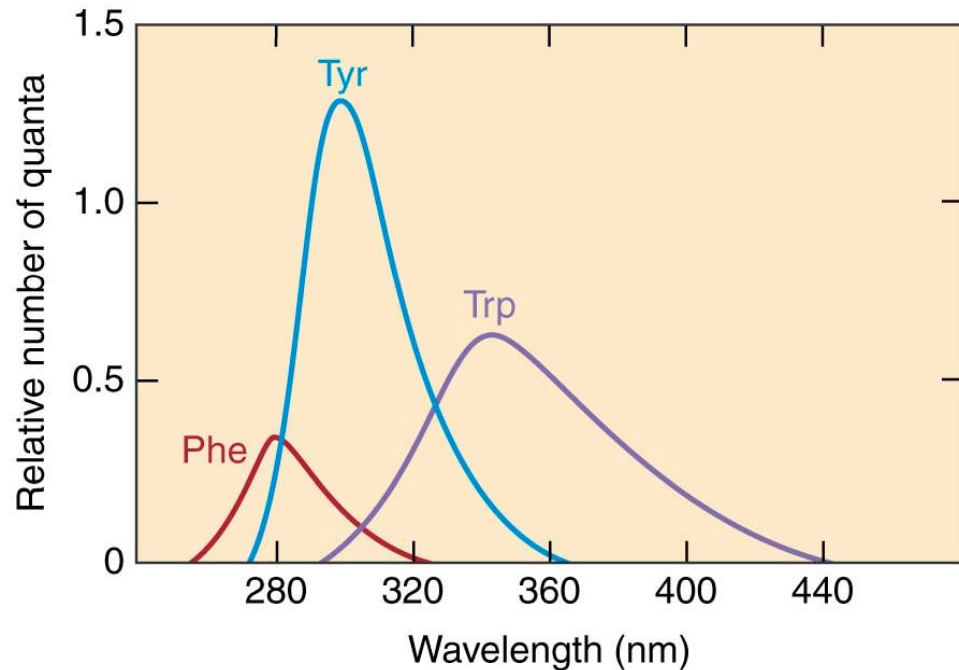
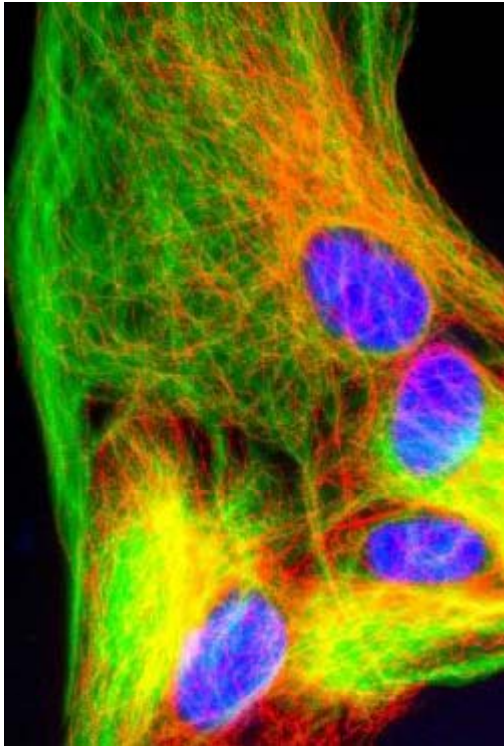


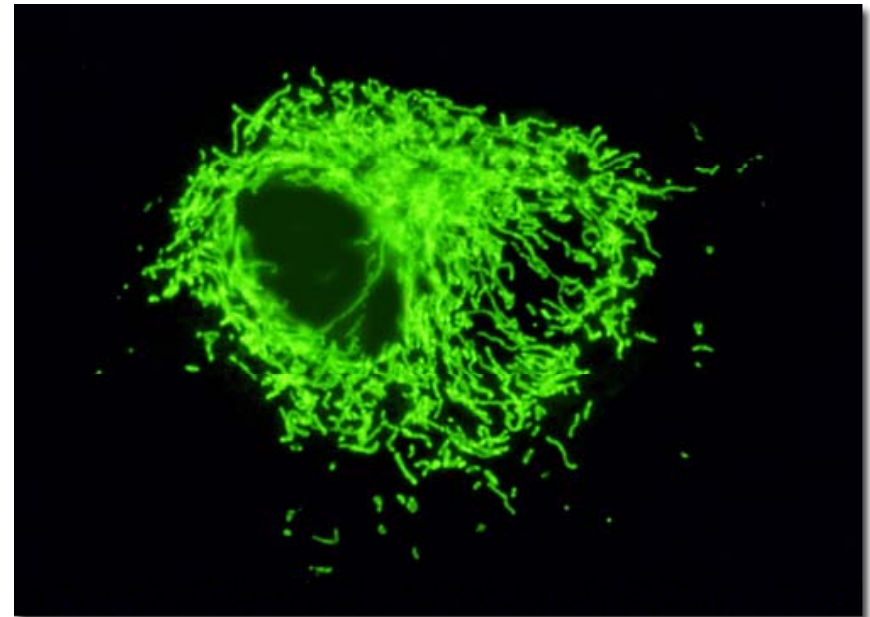
Figure 3.74. Characteristic fluorescence of aromatic groups in proteins. Redrawn from d'

Spectroscopic Properties

Fluorescence is used to visualize proteins inside the cell and determine their localization



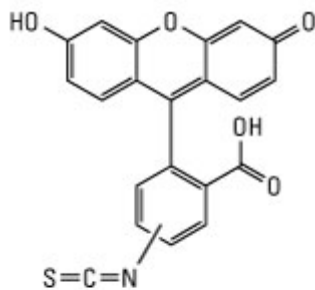
Nucleus



Mitochondria

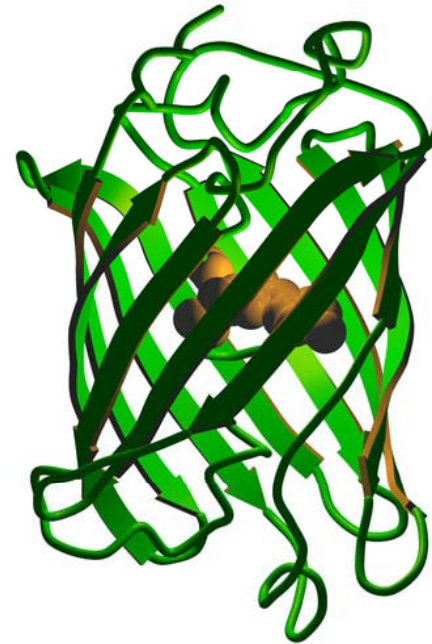
Spectroscopic Properties

Fluorescence is used to visualize proteins inside the cell and determine their localization



FITC
5/6-Fluorescein-isothiocyanate
MW 389.38

Protein Labeling



Green Fluorescent Protein
from jellyfish *Aequorea victoria*

Protein Analysis



- Separation of Proteins
- General Approach to Protein Purification
- Determination of Amino Acid Composition
- Spectroscopic Methods
- Determination of 3D Structure

Protein Analysis

(by Techniques)

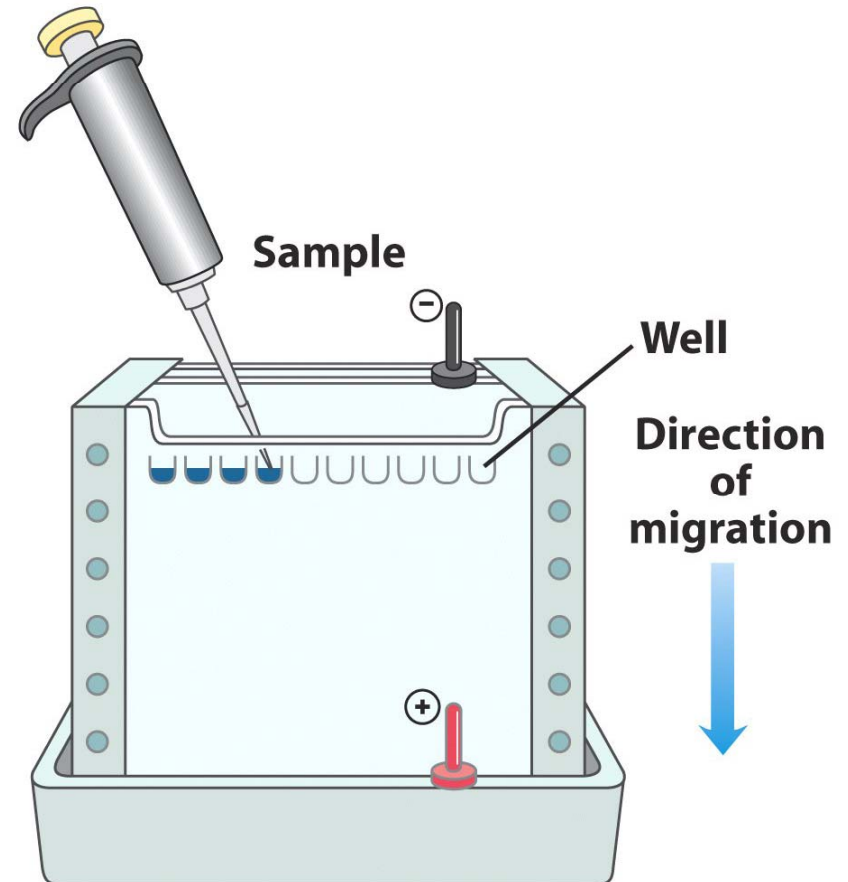
- Precipitation
- Ultracentrifugation
- Electrophoresis
 - ✓ Isoelectric focusing
 - ✓ SDS-PAGE Gel Electrophoresis
 - ✓ 2D gel electrophoresis
- Chromatography
 - ✓ Ion Exchange
 - ✓ Gel Filtration
 - ✓ Affinity
- Others

Electrophoresis

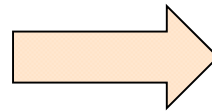
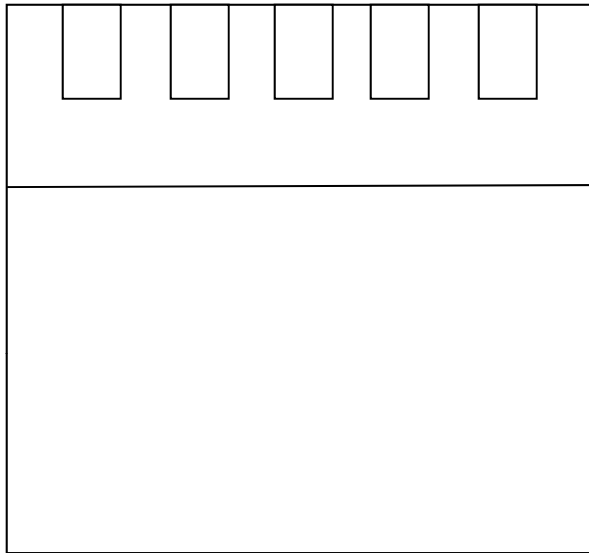
- Based on the movement of charged molecules when an electric field is applied
- Protein migrates according to its mass/charge ratio
- Use inert matrix:
 - ✓ Acrylamide
 - Small pore gels
 - used to separate most proteins
 - 5,000 to 200,000 Da
 - ✓ Agarose

Electrophoresis

- Molecules negatively charged will move towards the anode

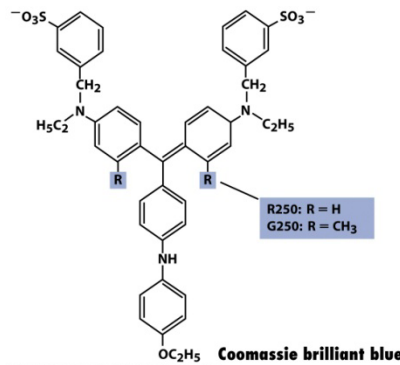
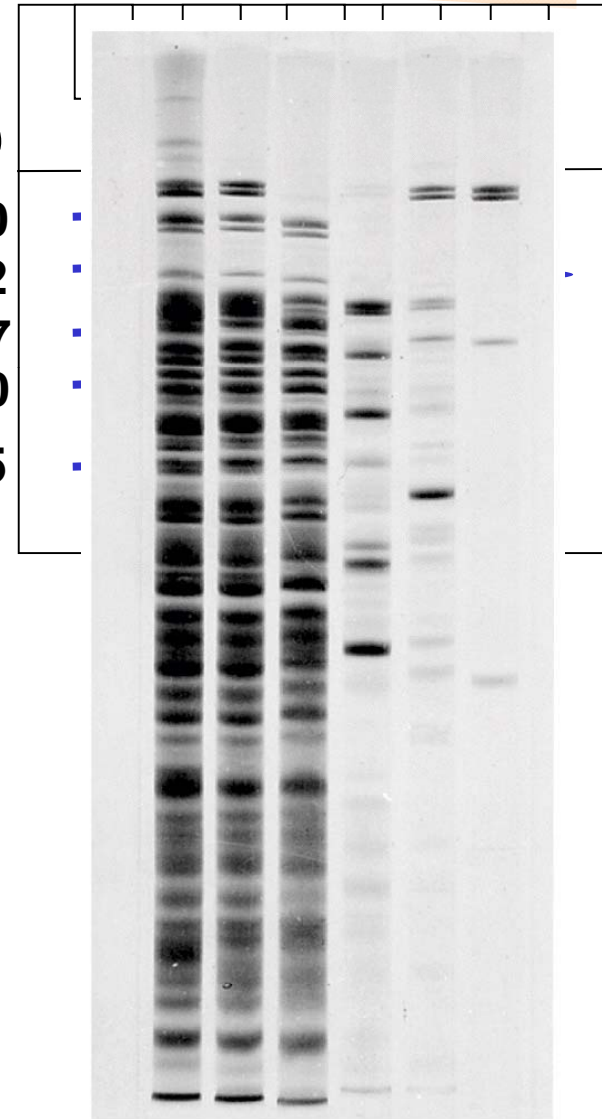


Electrophoresis



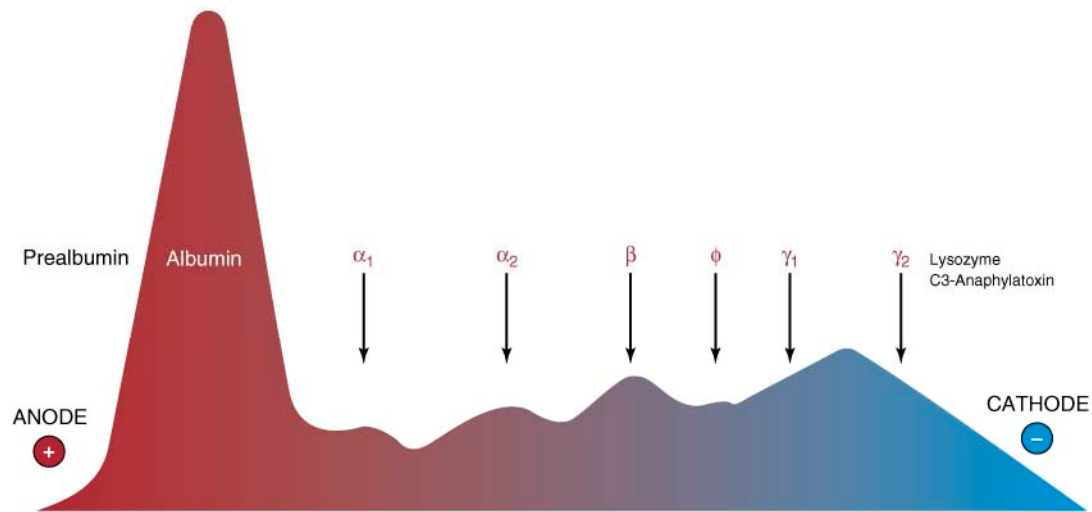
**Coomassie
Blue
Staining**

kD
170
112
87
50
25



- Other Stains:
 - ✓ Silver Stain

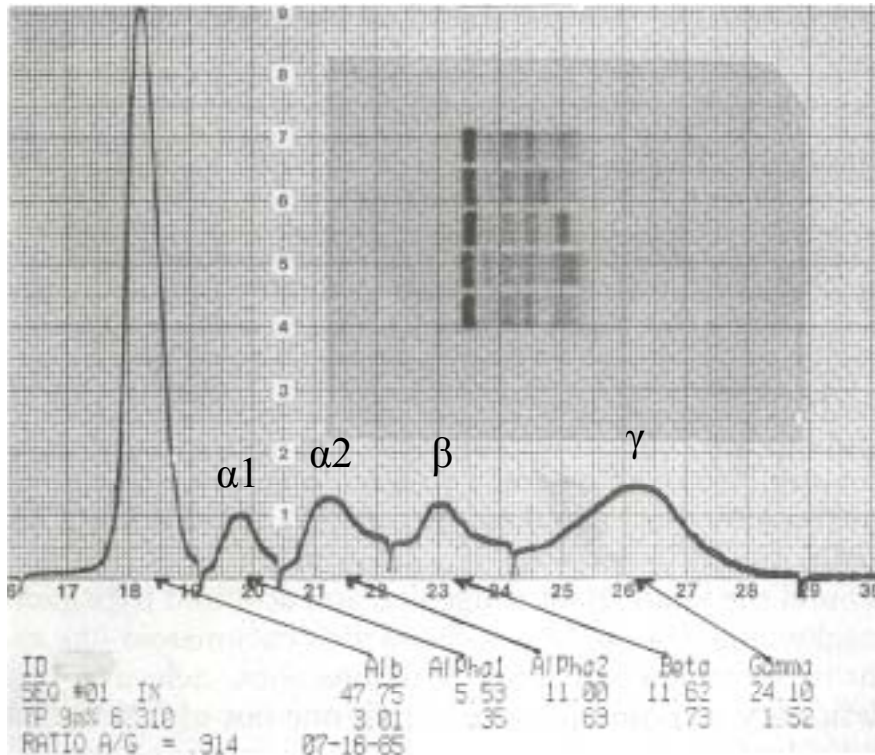
Electrophoresis Pattern for Serum Proteins



<p><u>α_1</u></p> <p>α_1-Acid glycoprotein α_1-T Glycoprotein α_1-Antitrypsin Transcortin α_1-Antichymotrypsin α_1-B Glycoprotein α_1-glycoprotein Vitamin D-binding protein α_1-Lipoproteins</p>	<p><u>α_1/α_2</u></p> <p>Thyroxine-binding globulin Zn-α_2-glycoprotein Gc globulin Ceruloplasmin Inter-α-trypsin inhibitor Antithrombin III Factor X (Stuart-Prower) Transcobalamin I C9</p>	<p><u>α_2</u></p> <p>Retinol-binding protein α_2 HS glycoprotein Histidine-rich 3,8 S-α_2-glycoprotein Haptoglobin Pregnancy zone protein α_2-Macroglobulin Prothrombin Antihemophilic factor C1 inactivator C1s</p>	<p><u>α_2/β_1</u></p> <p>Serum cholinesterase α_3-glycoprotein α_2, β_1-glycoprotein Transcobalamin III</p>
<p><u>β_1</u></p> <p>Hemopexin Steroid-binding β-globulin Transferrin Pregnancy-specific β_1-glycoprotein Cold insoluble globulin Factor V (Accelerin) Factor VII (Proconvertin) Factor IX (Christmas) Plasminogen C3 Proactivator Transcobalamin II β-Lipoproteins C1r, C2 C4, C5</p>	<p><u>β_2</u></p> <p>β_2-Microglobulin β_2-Glycoprotein III β_2-Glycoprotein I Fibrinogen Factor XI (PTA) Factor XII (Hageman) Factor XIII (FSF) C3, C6, C7 C3 Activator (β_2II)</p>	<p><u>γ_1</u></p> <p>IgG IgA IgD IgE IgM Amylase</p>	<p><u>γ_2</u></p> <p>IgG Ciq Properdin</p>

Electrophoresis Pattern for Serum Proteins

Albumin



In response to infections there is an increase in the intensity of the α_2 and γ fractions

Hypo-gamma-globulinemia is a virtual lack of gamma-globulins and is symptomatic of immunosuppression.

Hyper-gamma-globulinemia used to diagnose liver disease and lupus erythematosus

In hepatic cirrhosis there is a decrease in total albumin together with an increase in the gamma fractions.

α_1 band indicative of antitrypsin deficiency

Electrophoresis Pattern for Serum Proteins

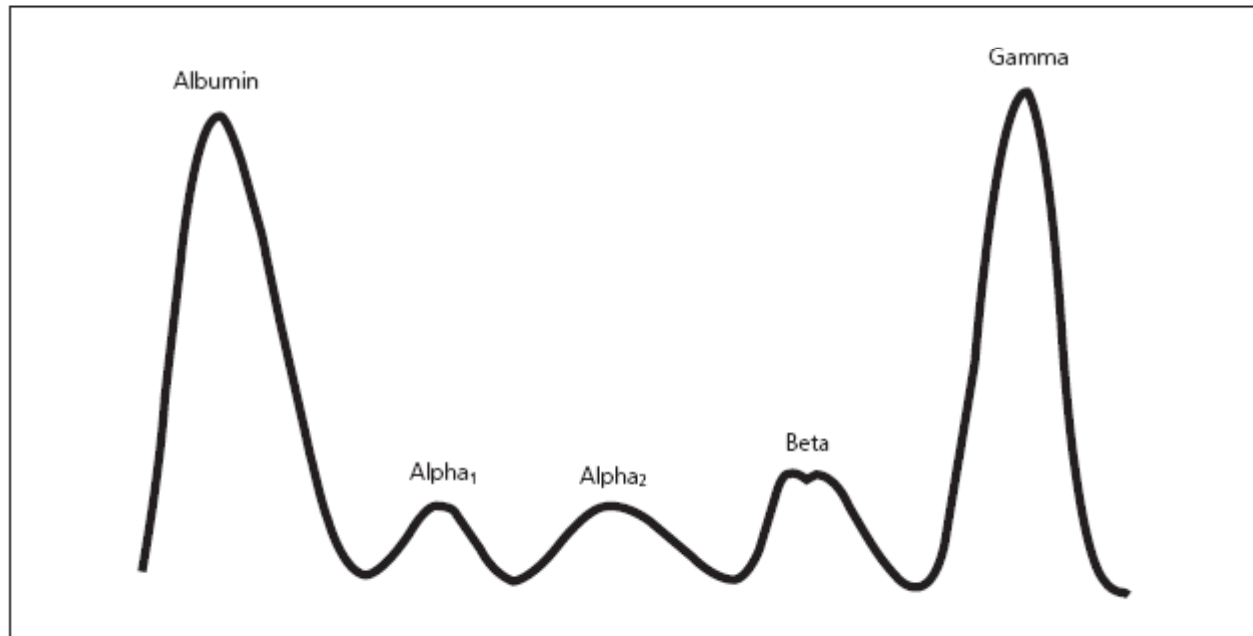
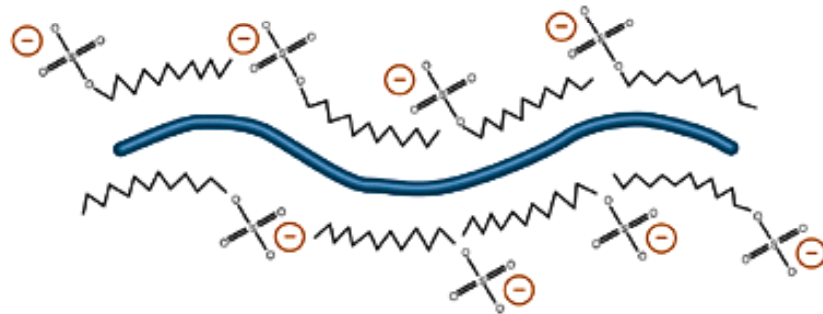


Figure 2. Abnormal serum protein electrophoresis pattern in a patient with multiple myeloma. Note the large spike in the gamma region.

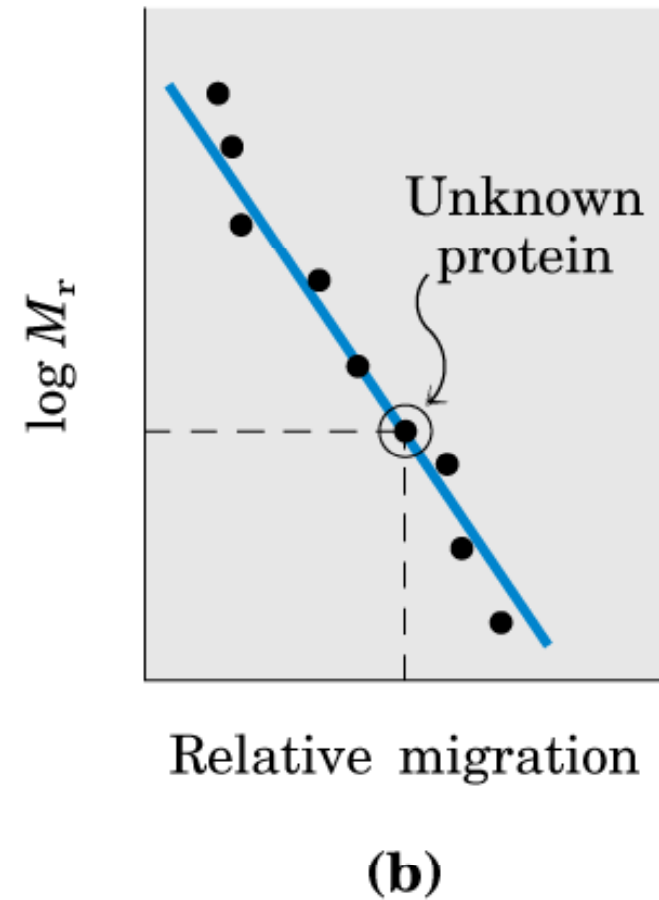
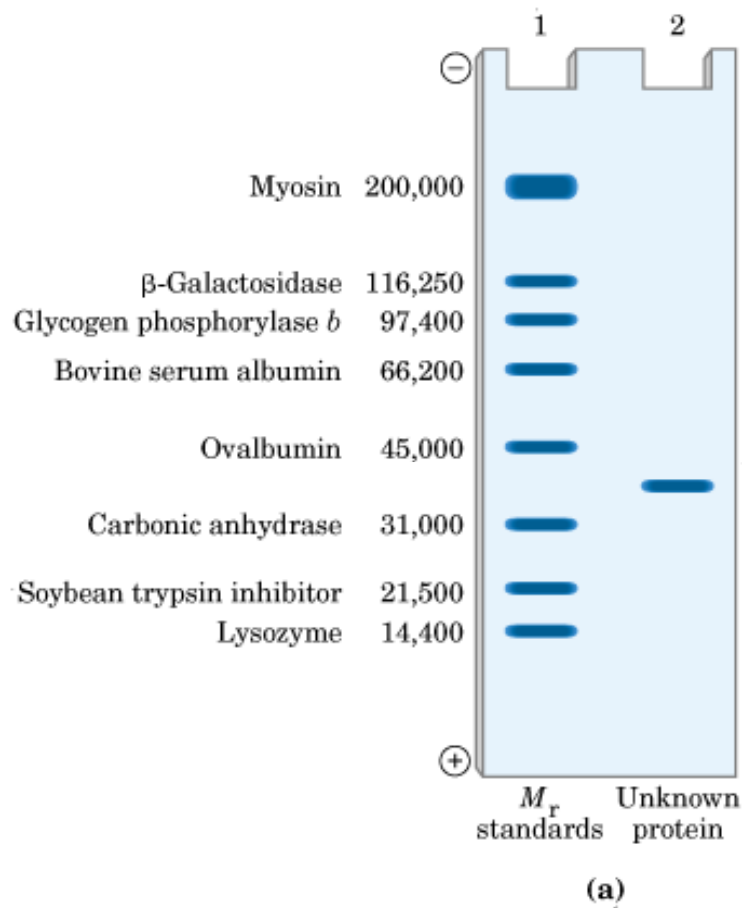
Gamma fraction spikes correlate with multiple myeloma and Hodgkin's disease
O'Connell *et al.* (2005) American Family Physician

SDS-PAGE

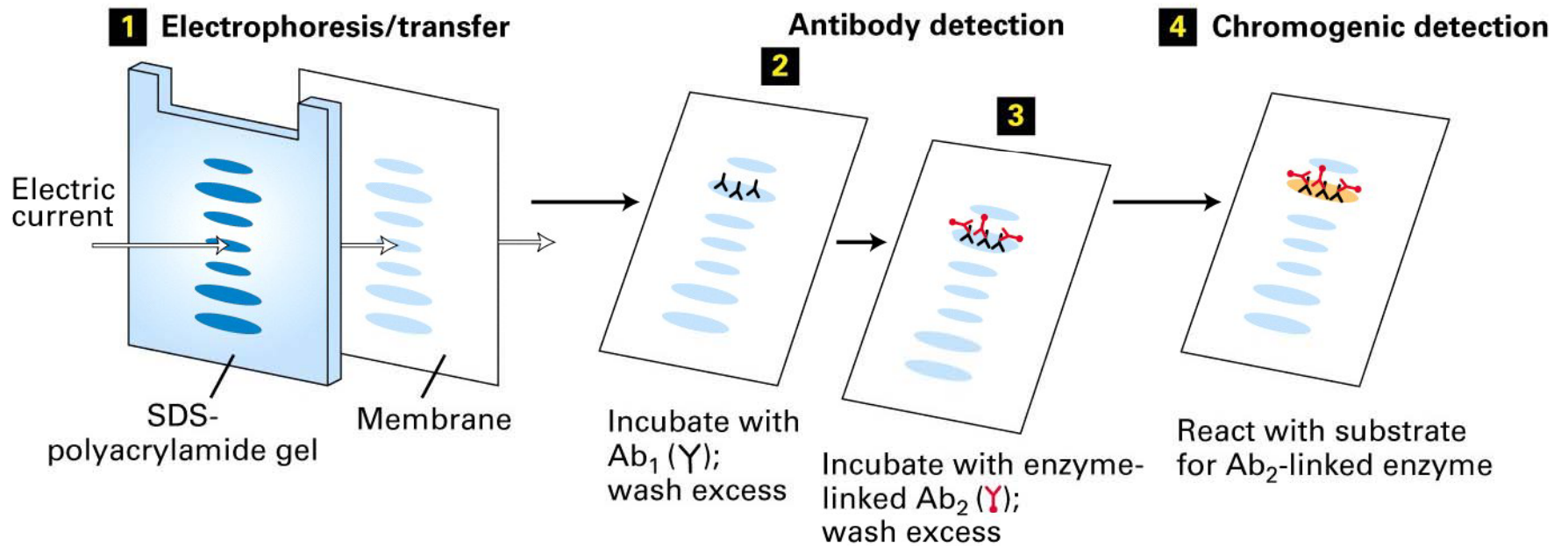
- PAGE = PolyAcrilamide Gel Electrophoresis
- SDS = sodium dodecyl sulfate
- Separate proteins based on size
- All proteins will be negatively charged and will migrate based on size (not charge/size)



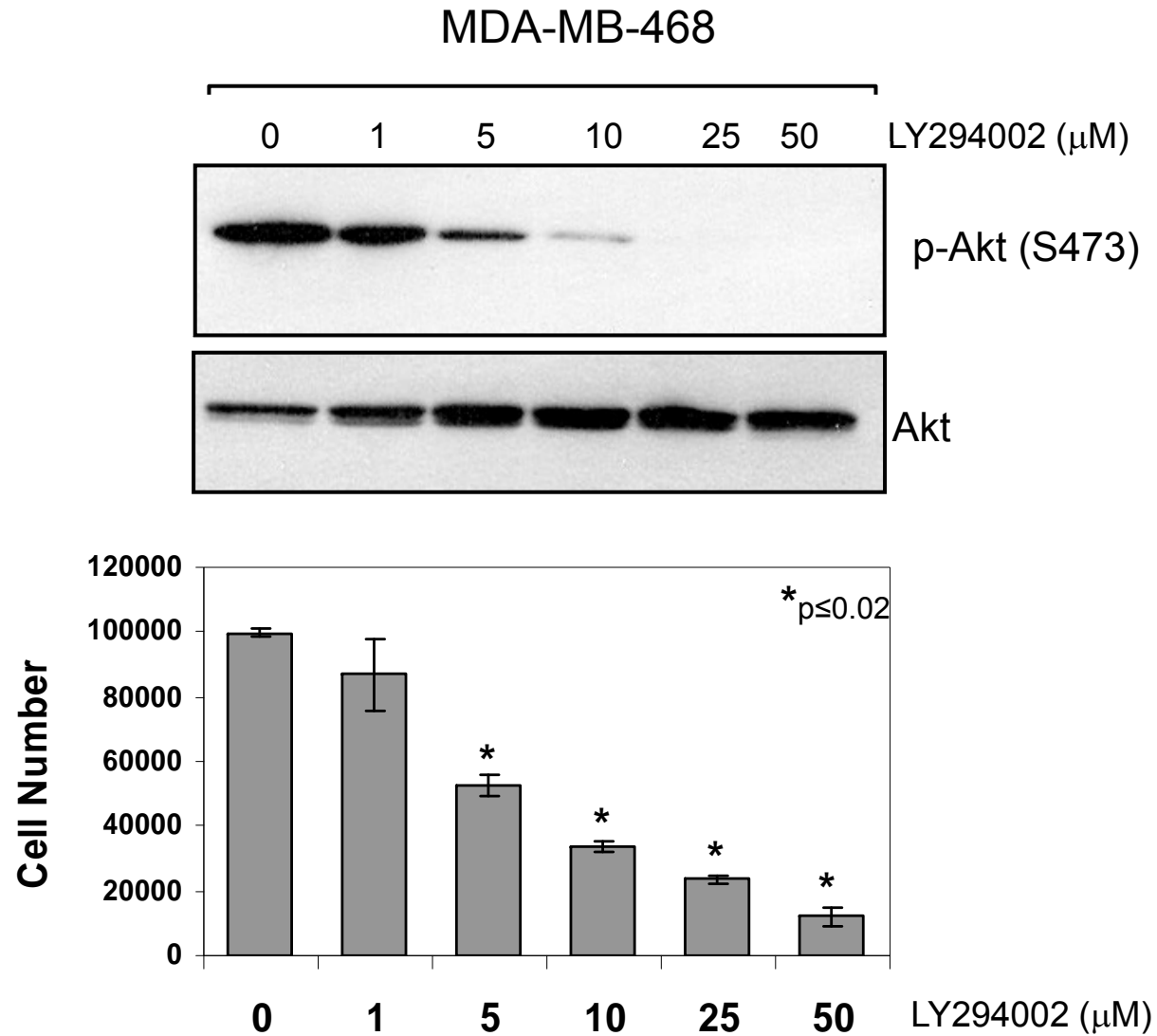
SDS - PAGE



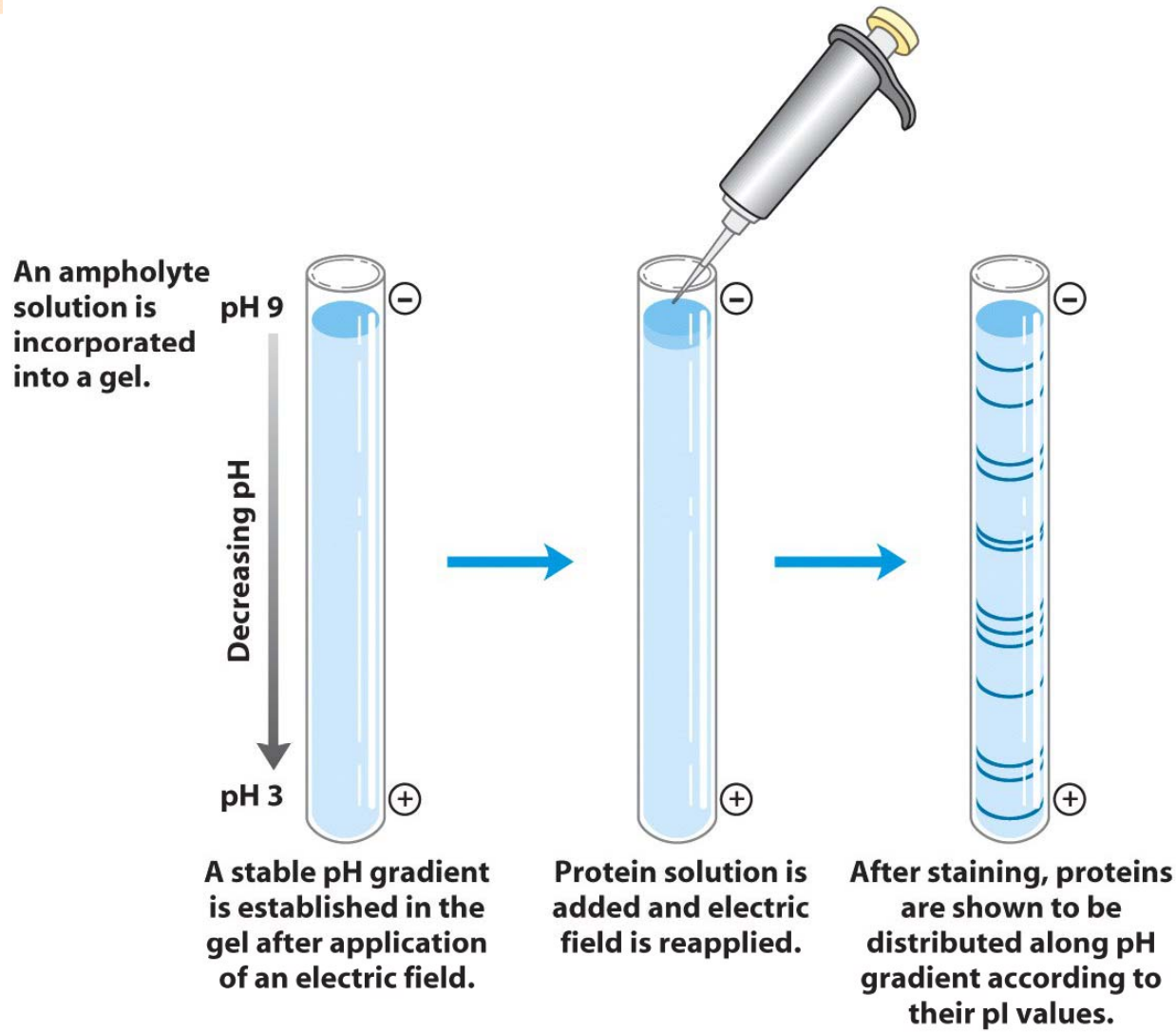
Western Blot



Western Blot - Example



Isoelectric focusing



pI

Table 5-2 Isoelectric Points of Several Common Proteins

Protein	pI
Pepsin	<1.0
Ovalbumin (hen)	4.6
Serum albumin (human)	4.9
Tropomyosin	5.1
Insulin (bovine)	5.4
Fibrinogen (human)	5.8
γ -Globulin (human)	6.6
Collagen	6.6
Myoglobin (horse)	7.0
Hemoglobin (human)	7.1
Ribonuclease A (bovine)	9.4
Cytochrome <i>c</i> (horse)	10.6
Histone (bovine)	10.8
Lysozyme (hen)	11.0
Salmine (salmon)	12.1

pH > pI, then protein charge negative
pH < pI, then protein charge positive

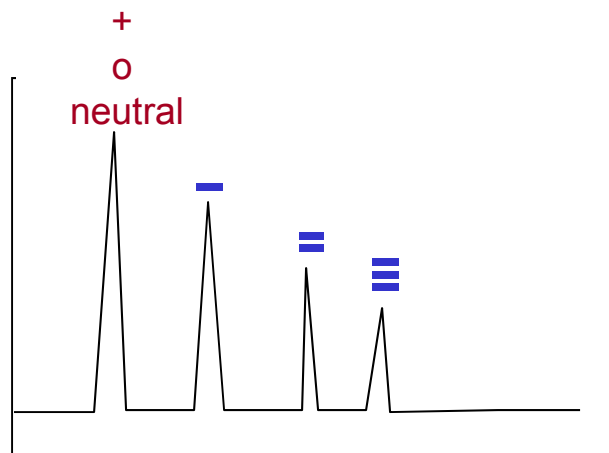
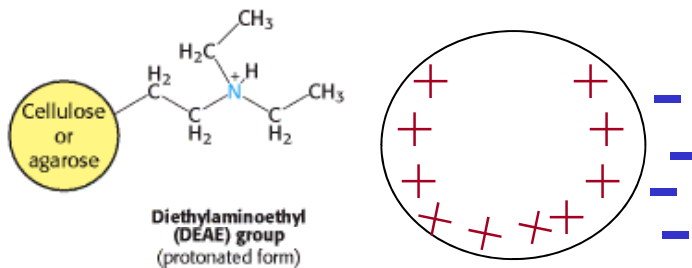
Chromatography

- Group of techniques based in the separation of the components of a mixture and its detection
- Chromatographic techniques are diverse, but all consist of a **mobile phase** (gas, liquid) that moves the sample along a **stationary phase**.
- Each component of the sample will interact in a different manner with the mobile and stationary phases.

Ion Exchange Chromatography

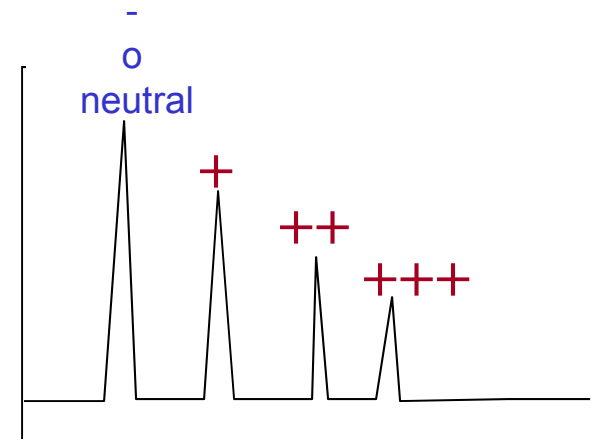
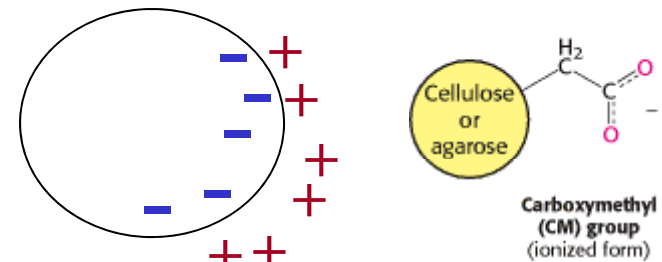
■ Anionic Exchange

- ✓ resin - positive (+)
- ✓ Attracts anions (-)

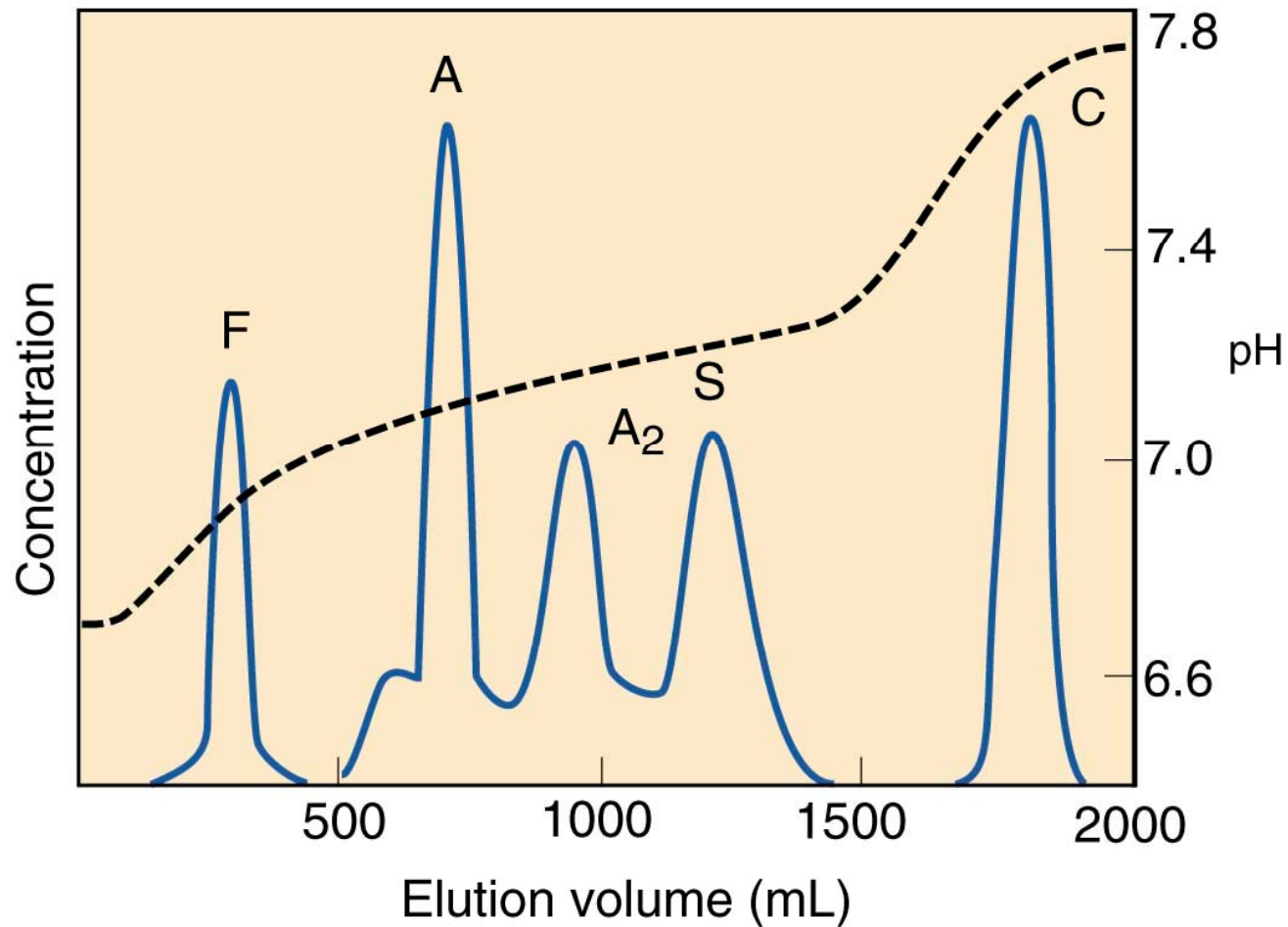


■ Cationic Exchange

- ✓ resin - negative (-)
- ✓ Attracts cations (+)

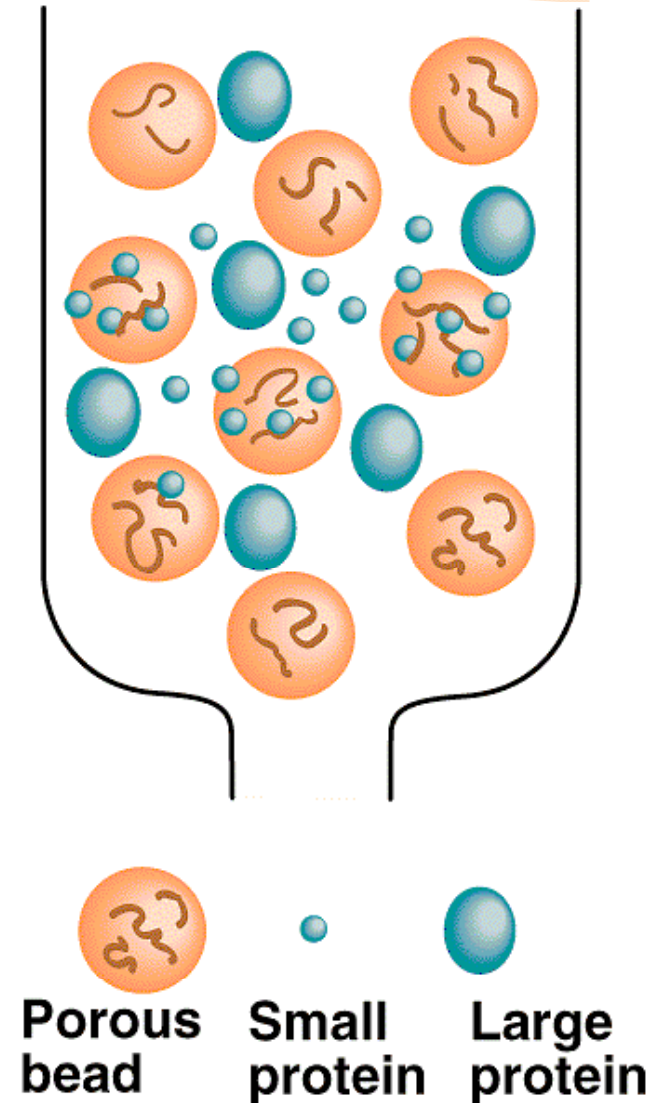


Ion Exchange Chromatography

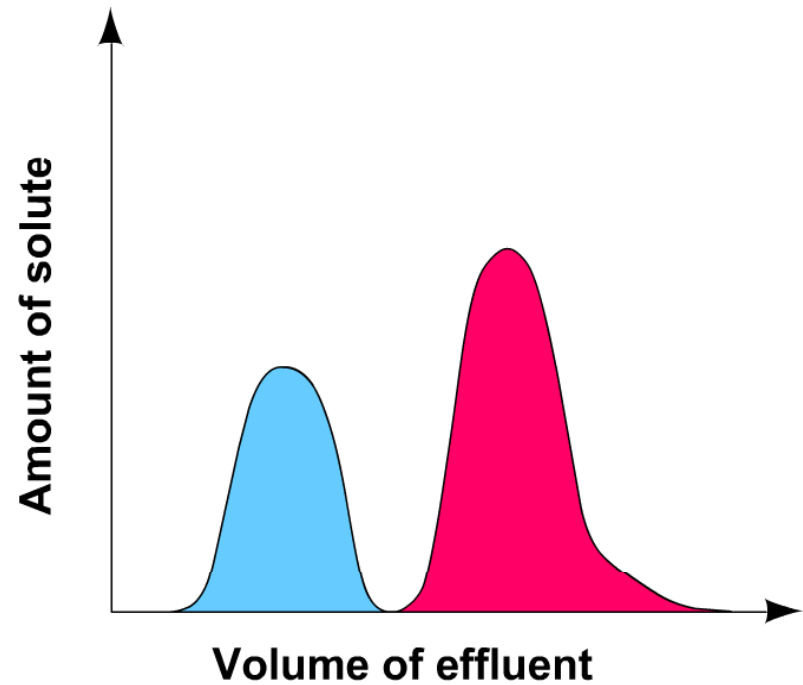
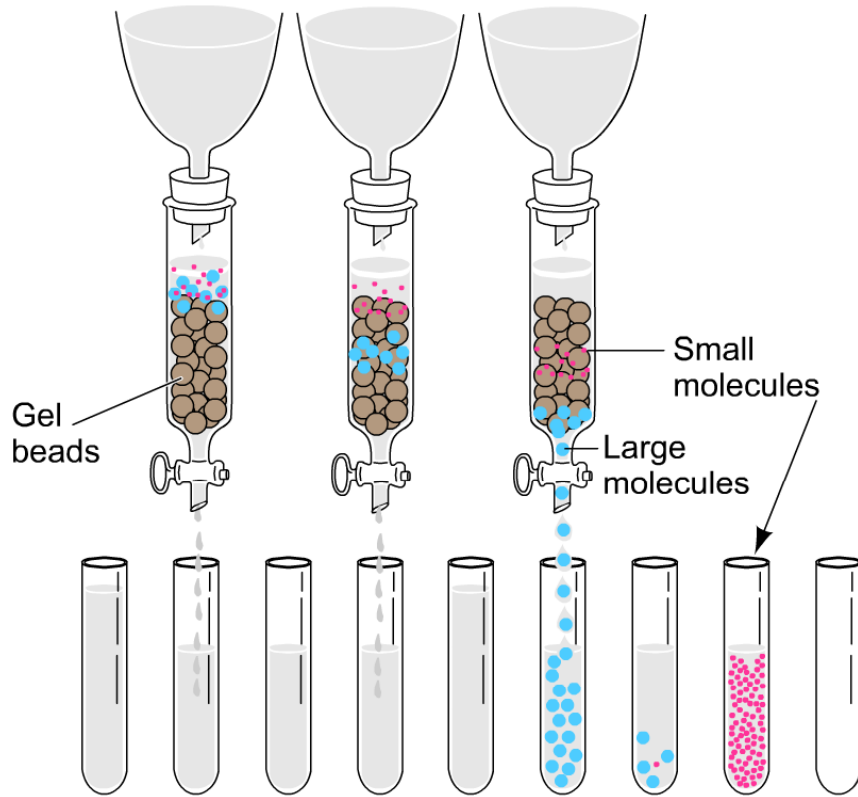


Size-Exclusion Chromatography

- aka: Gel Filtration Chromatography
- Separates by size
- Small proteins penetrate the pores of the gel and have a larger solvent volume through which to travel in the column than larger proteins



Size-Exclusion Chromatography



Other Separation Techniques:

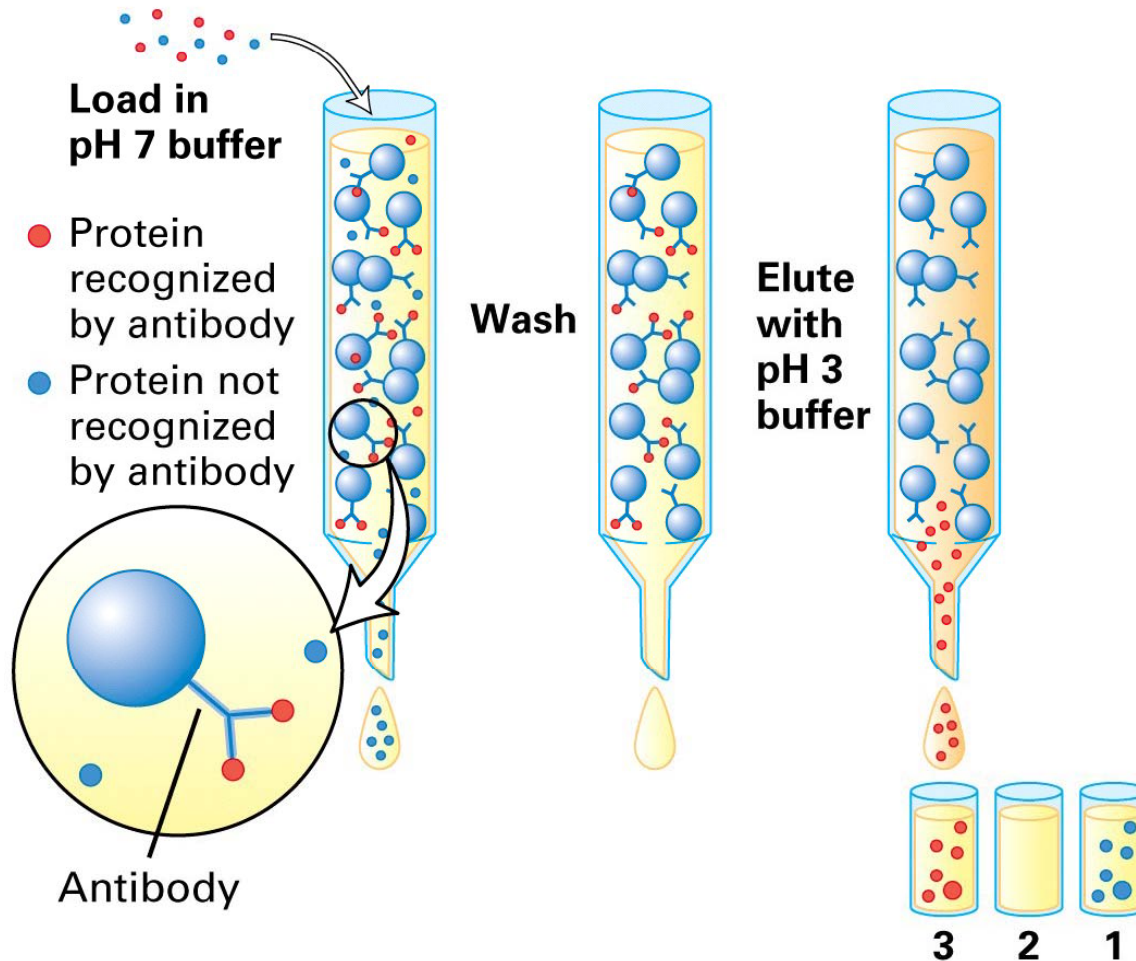


Affinity Chromatography

- Take advantage of specific interactions of proteins
- Examples:
 - ✓ enzyme - substrate
 - ✓ antigen - antibody
 - ✓ Any other biological pair

Affinity Chromatography

(c) Antibody-affinity chromatography

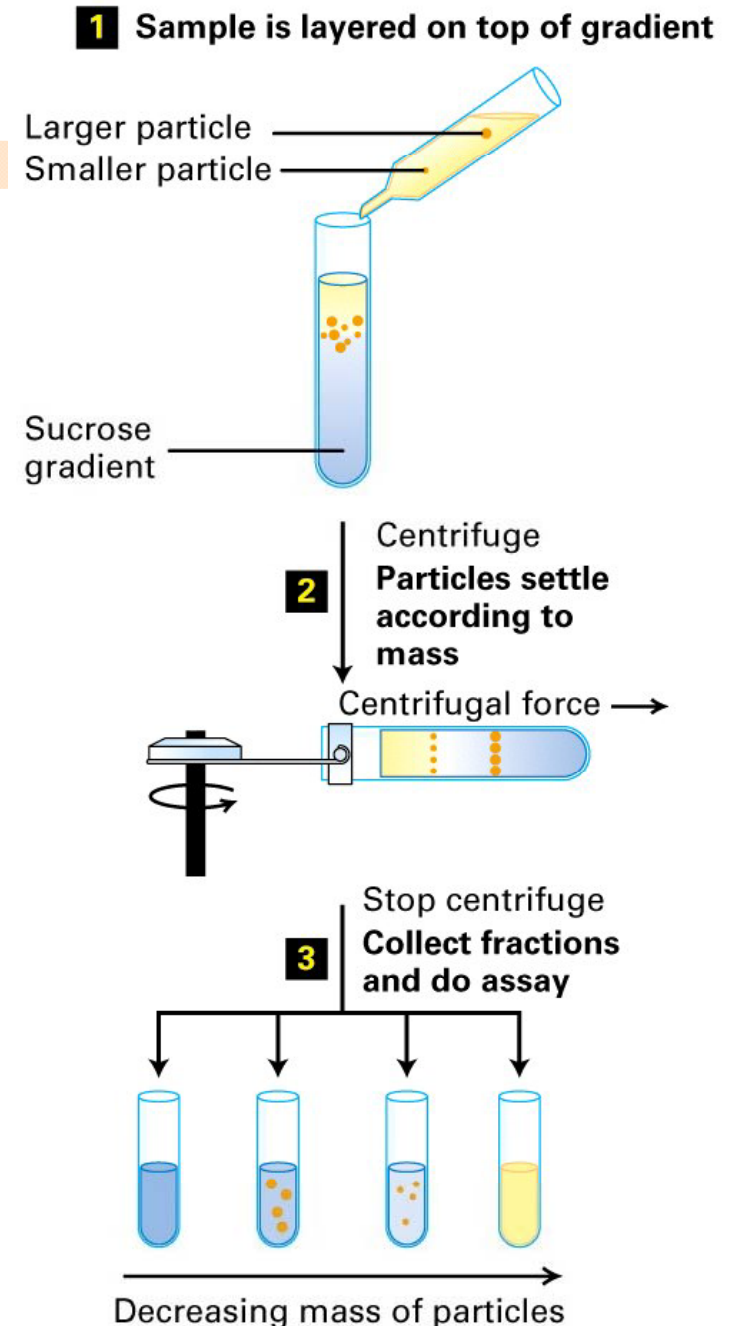


Ultracentrifugation

- Measures sedimentation coefficient
 - ✓ Svedver units (10^{-13} sec)
 - ✓ Method to determine MW
- Use a gradient
 - ✓ Sucrose
 - ✓ Cesium Chloride
- Protein will migrate at a rate controlled by factors that affect sedimentation constant

$$s = \frac{v}{\omega^2 r}$$

Equation for calculation of the Svedberg coefficient.



Ultracentrifugation

Table 3.17 Svedberg Coefficient for some Plasma Proteins

Protein	Svedberg Coefficient	Mol. Weight
Lysozyme	2.19	15,000 – 16,000
Albumin	4.6	69,000
Immunoglobulin G	6.6 – 7.2	153,000
Fibrinogen	7.63	341,000
C1q	11.1	410,000

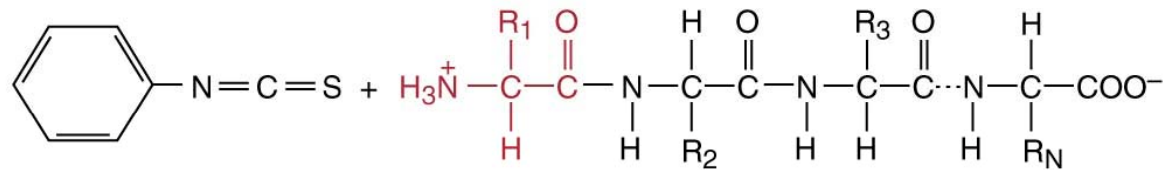
Determination of Amino Acid Sequence



- Classic Method – Edman Rxn.
 - ✓ Edman Reaction
 - ✓ Enzymatic Fragmentation
 - ✓ Chemical Fragmentation
- DNA Sequencing
- MS (Proteomics)

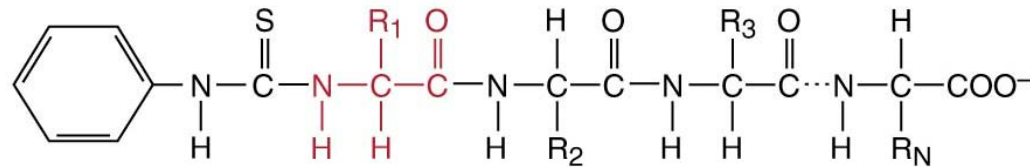
Determination of Amino Acid Sequence

Edman Rxn

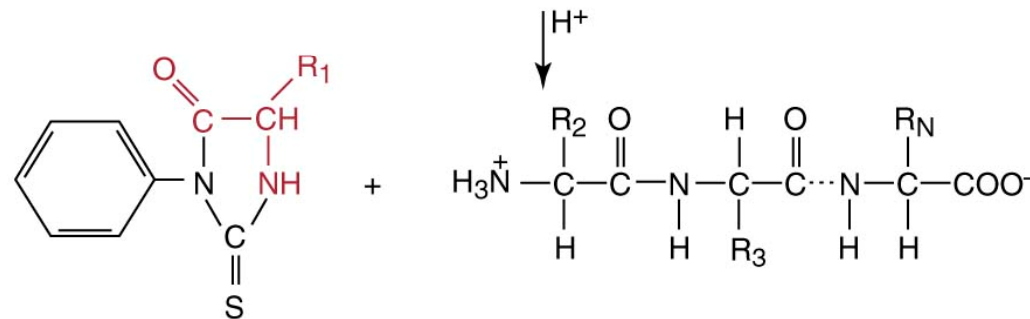


Phenylisothiocyanate

Polypeptide chain



Phenylthiocarbamoyl (PTC) peptide (or protein)



Phenylthiohydantoin

Polypeptide chain (minus original NH_2 - terminal amino acid)

Determination of Amino Acid Sequence

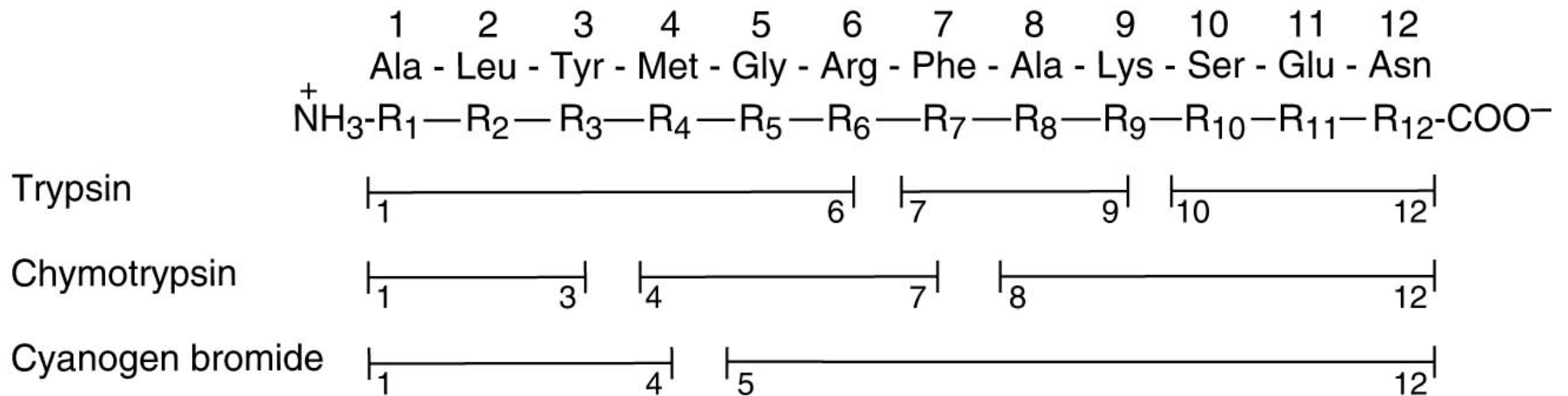


Figure 3.67. Ordering of peptide fragments from overlapping sequences produced by specific proteolysis of a peptide.

In the Post-Genome Era

DNA \longrightarrow Transcription \longrightarrow Translation \longrightarrow Protein

Amino acid

sequence (protein)

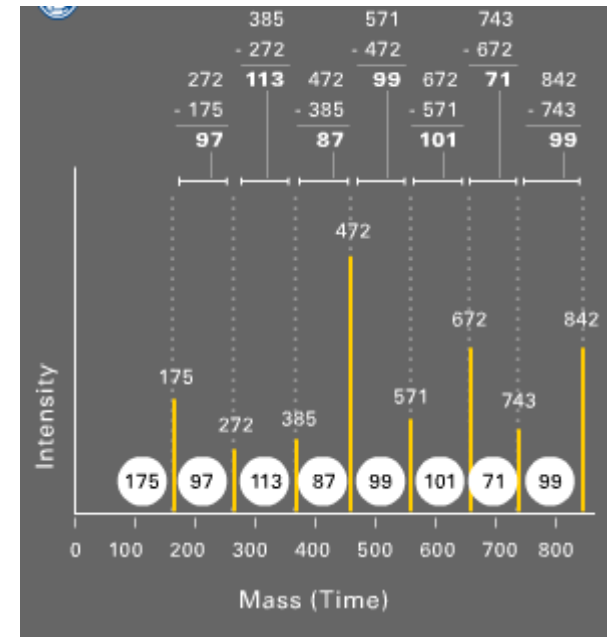
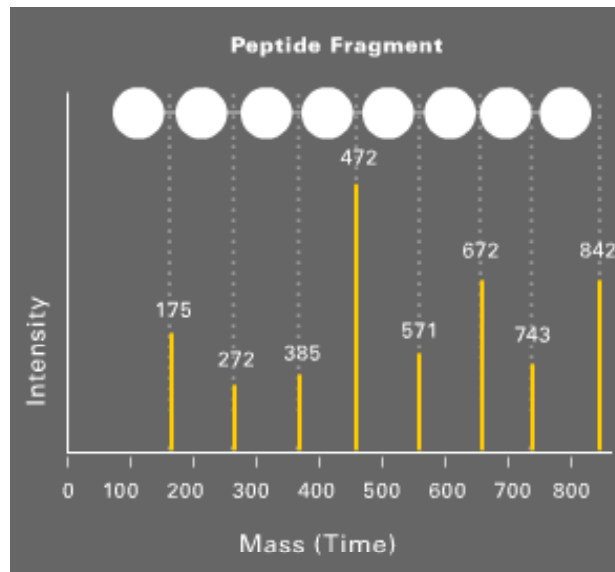
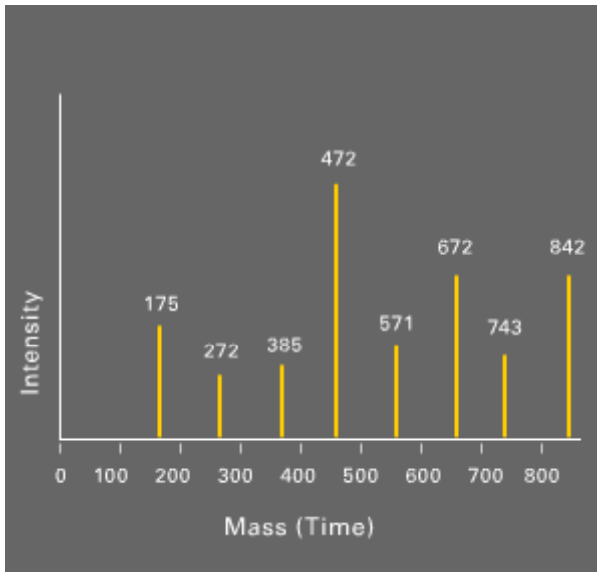
Gln – Tyr – Pro – Thr – Ile – Trp

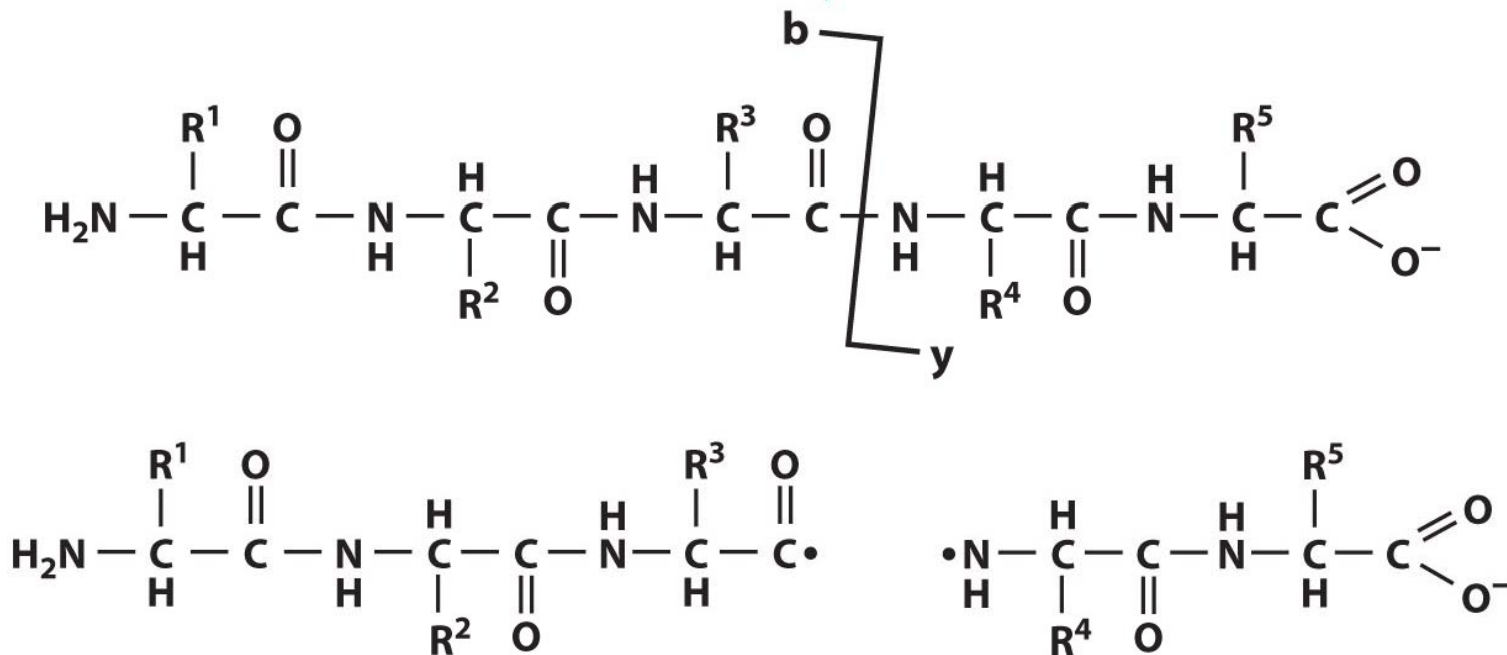
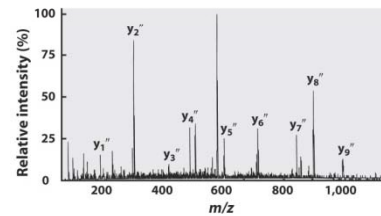
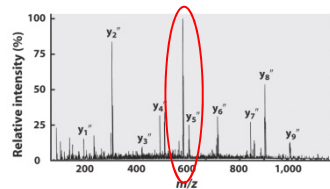
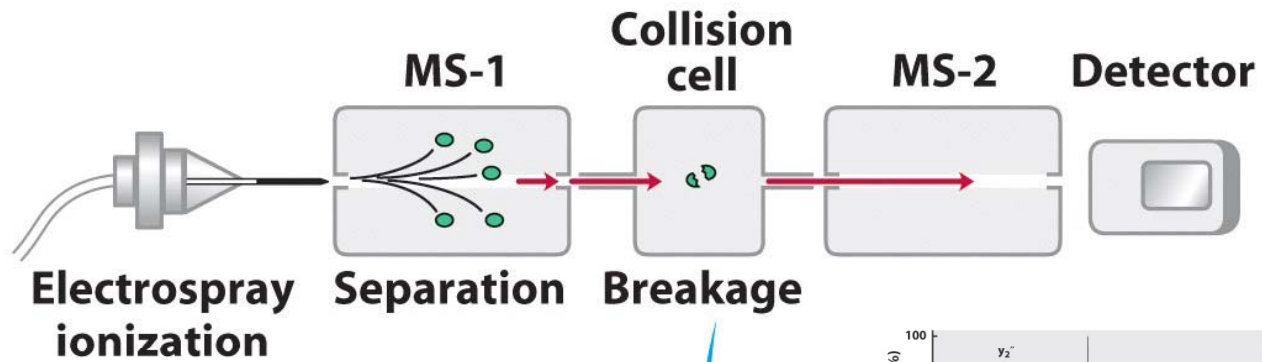
DNA sequence (gene)

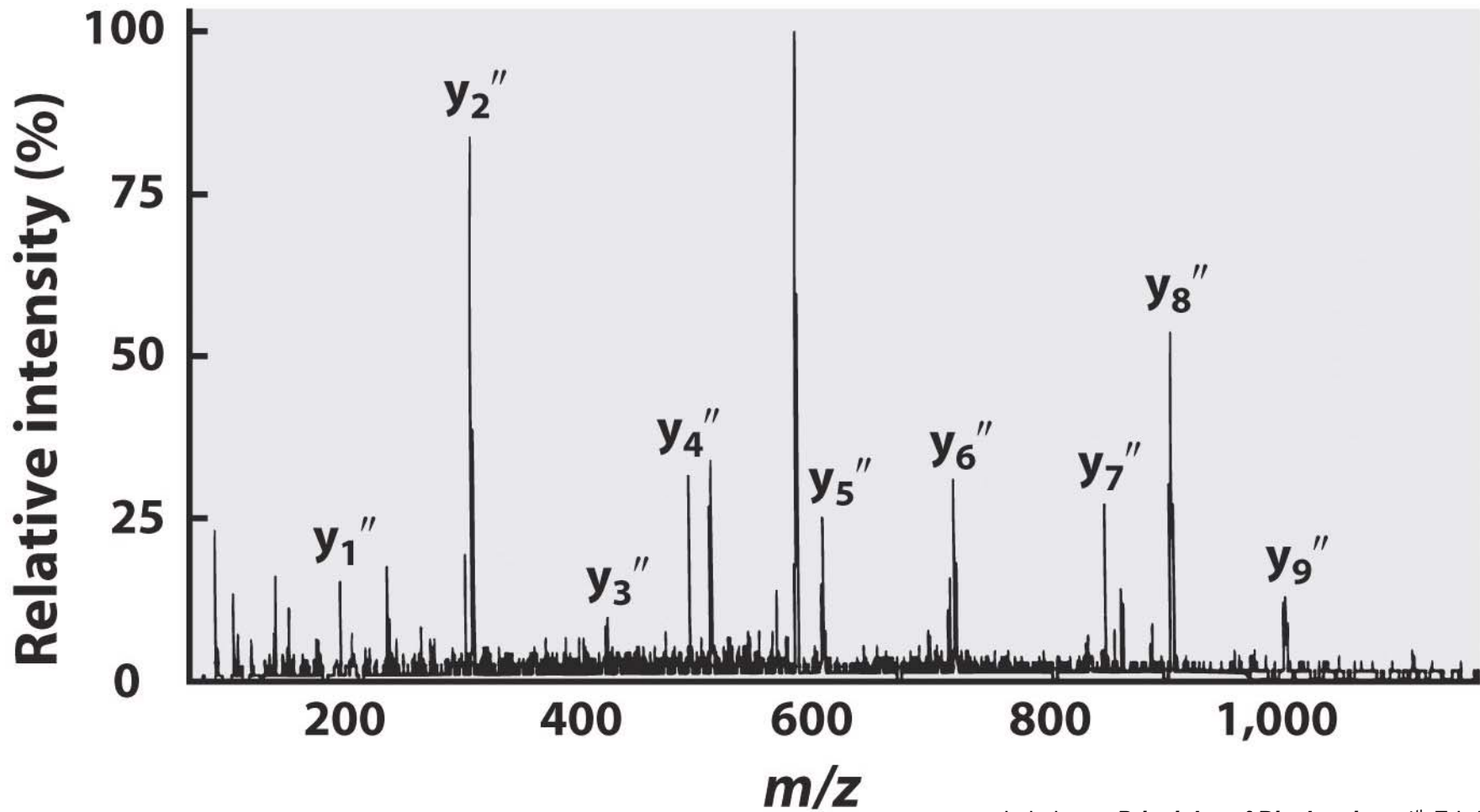
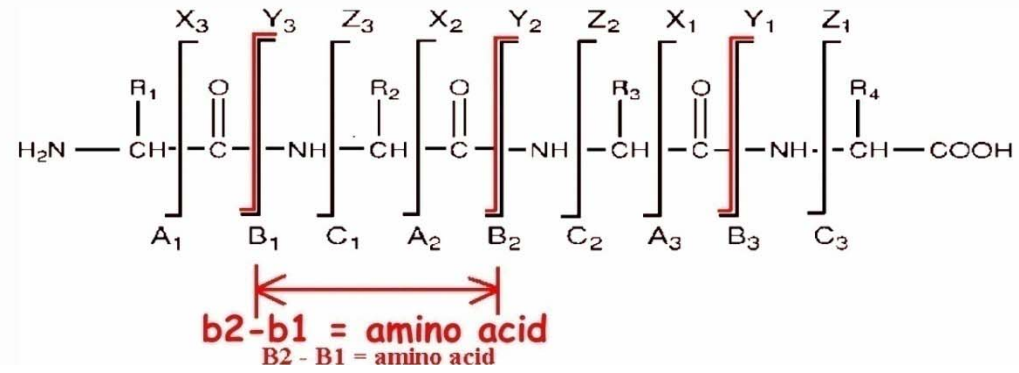
CAGTATCCTACGATTTCG

- Method to predict the amino acid sequence of a protein
- Disadvantage:
 - ✓ Do not predict the position of disulfide bonds
 - ✓ Do not identify modified aa (post-translation)

Mass Spectrometry for Protein Sequence Determination



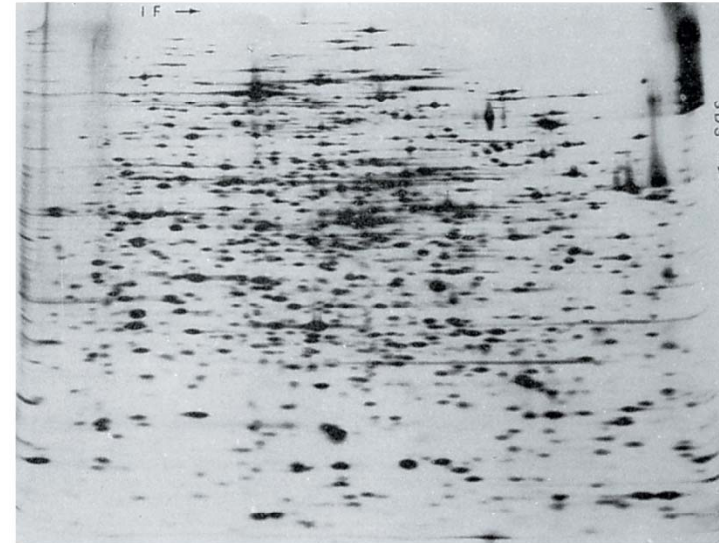
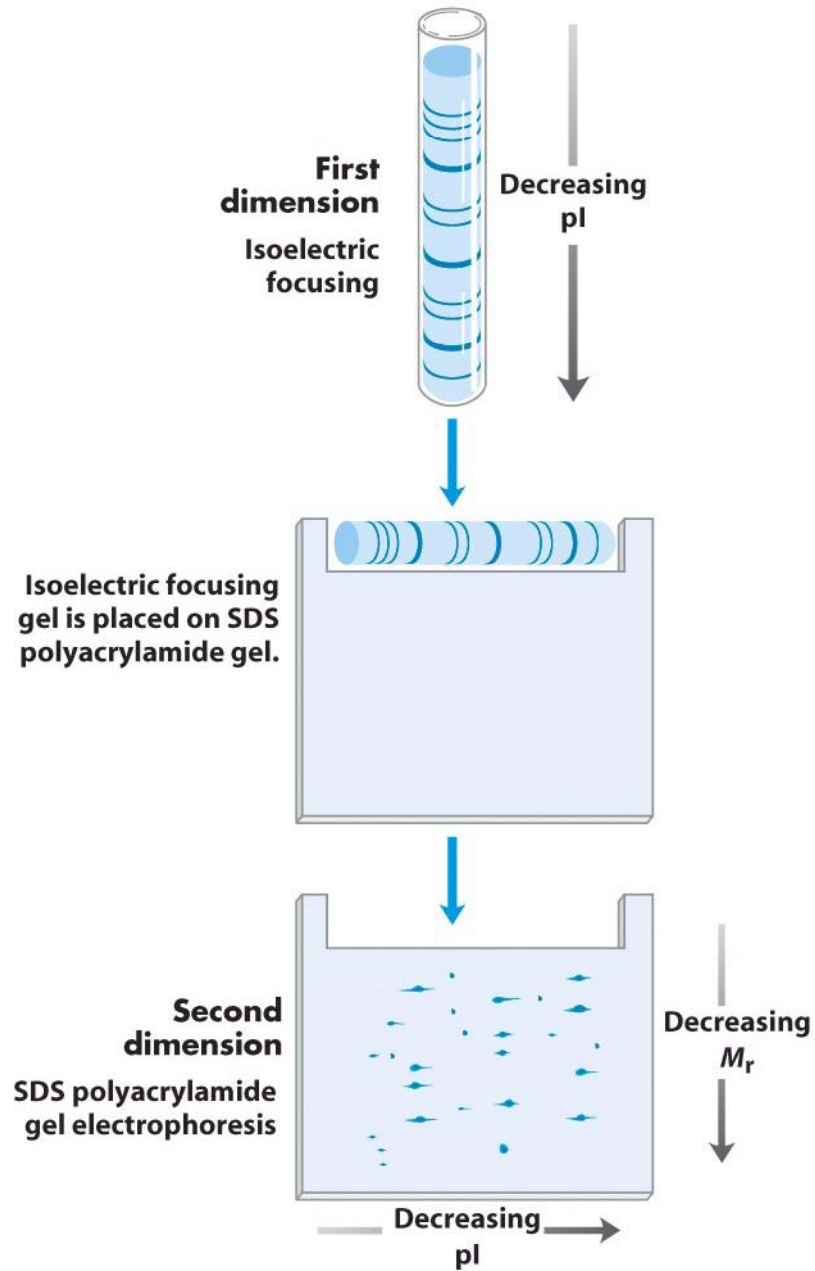




Proteomics

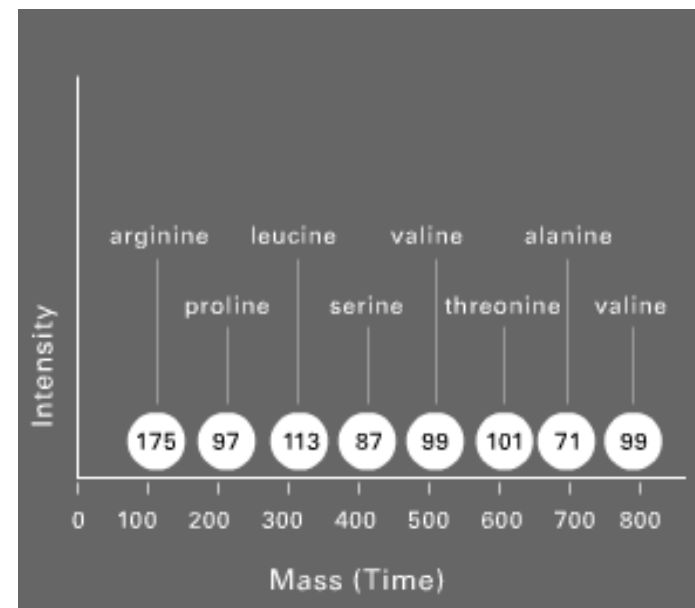
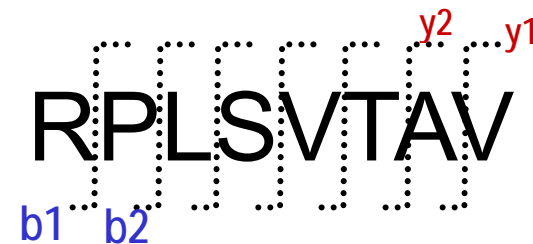
- Systematic study of the amounts, modifications, interactions, localization and function of all or a subsets of proteins at the whole-organism, tissue, cellular and sub-cellular levels.
- Can address questions as:
 - ✓ In a given sample, what fraction of the proteoma is expressed?
 - ✓ Of those present – relative abundance?
 - ✓ Chemically modified?
 - ✓ Protein profile change due to a condition? Cancer changes the profile of serum proteins? Response to tx alter profile?

2D Electrophoresis

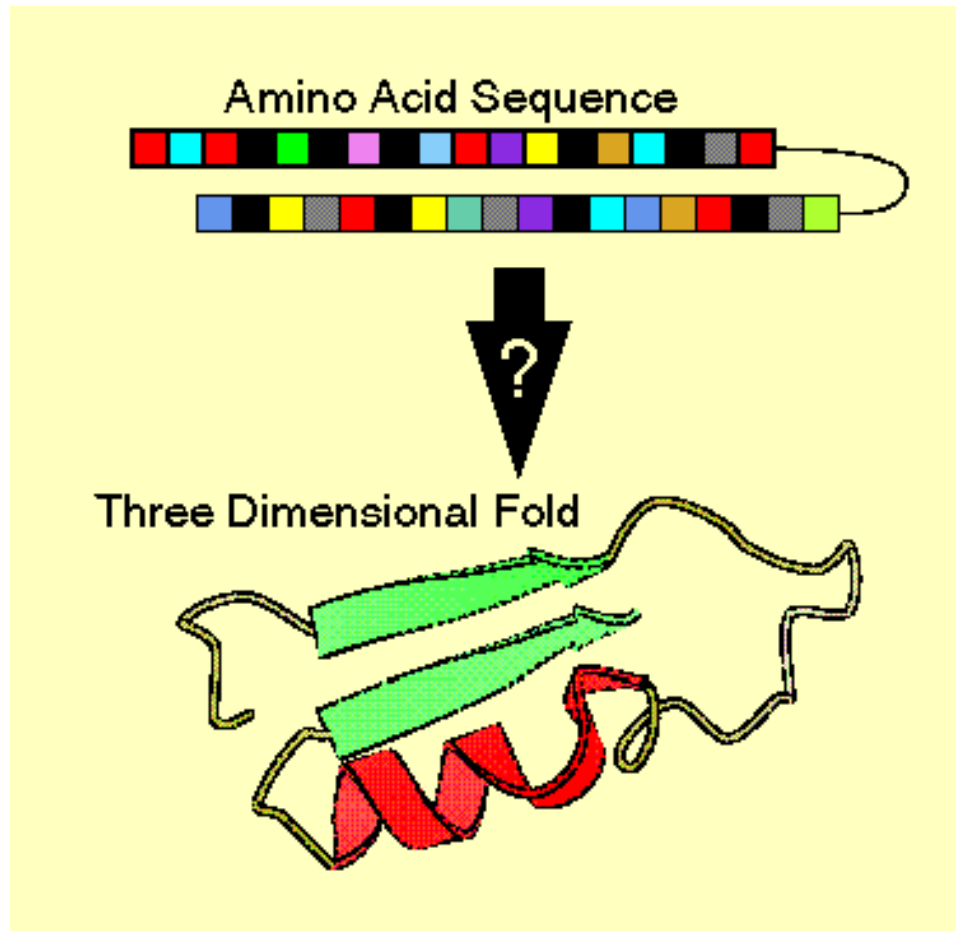


Symbol	Structure	Mass (Da)
Ala A	-NH.CH.(CH ₃).CO-	71.0
Arg R	-NH.CH.[(CH ₂) ₃ .NH.C(NH).NH ₂].CO-	156.1
Asn N	-NH.CH.(CH ₂ CONH ₂).CO-	114.0
Asp D	-NH.CH.(CH ₂ COOH).CO-	115.0
Cys C	-NH.CH.(CH ₂ SH).CO-	103.0
Gln Q	-NH.CH.(CH ₂ CH ₂ CONH ₂).CO-	128.1
Glu E	-NH.CH.(CH ₂ CH ₂ COOH).CO-	129.0
Gly G	-NH.CH ₂ .CO-	57.0
His H	-NH.CH.(CH ₂ C ₃ H ₃ N ₂).CO-	137.1
Ile I	-NH.CH.[CH.(CH ₃)CH ₂ .CH ₃].CO-	113.1
Leu	-NH.CH.[CH ₂ CH(CH ₃) ₂].CO-	113.1
Lys K	-NH.CH.[(CH ₂) ₄ NH ₂].CO-	128.1
Met M	-NH.CH.[(CH ₂) ₂ .SCH ₃].CO-	131.0
Phe F	-NH.CH.(CH ₂ Ph).CO-	147.1
Pro P	-NH.(CH ₂) ₃ .CH.CO-	97.1
Ser S	-NH.CH.(CH ₂ OH).CO-	87.0
Thr T	-NH.CH.[CH(OH)CH ₃].CO-	101.0
Trp W	-NH.CH.[CH ₂ .C ₈ H ₆ N].CO-	186.1
Tyr Y	-NH.CH.[(CH ₂).C ₆ H ₄ .OH].CO-	163.1
Val V	-NH.CH.[CH(CH ₃) ₂].CO-	99.1

Proteomics

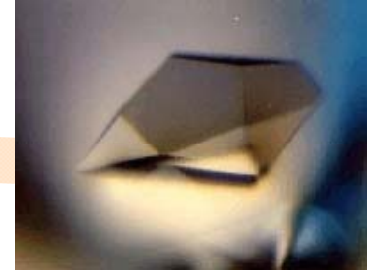


Determination 3D Structure

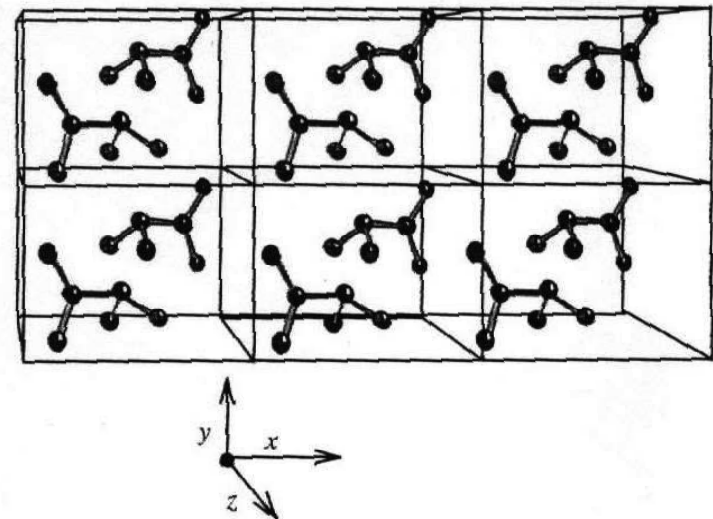
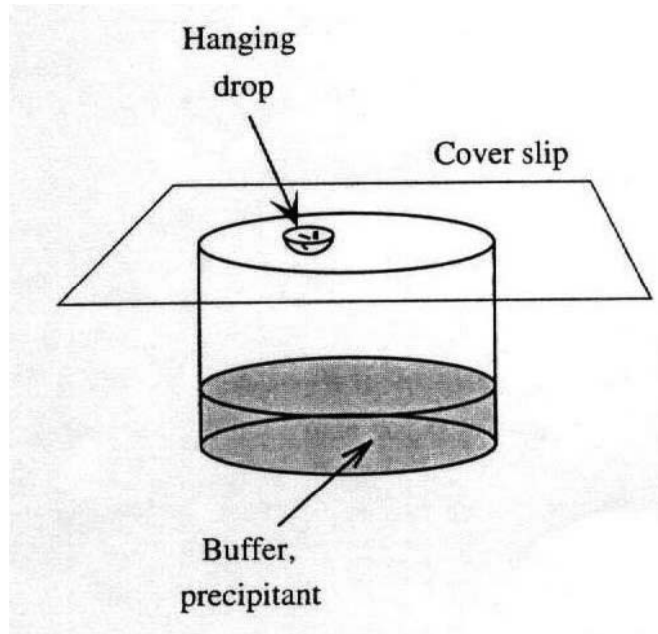


- X-ray Difraction
- NMR

Crystallography

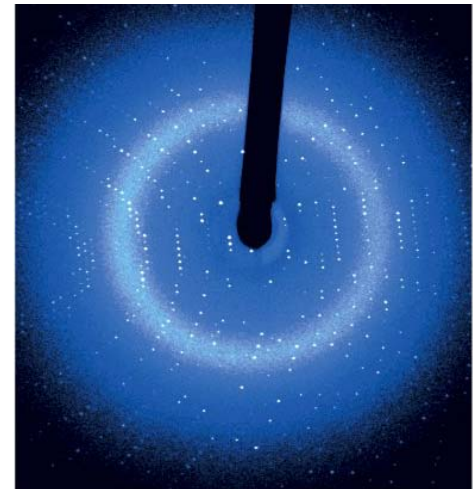
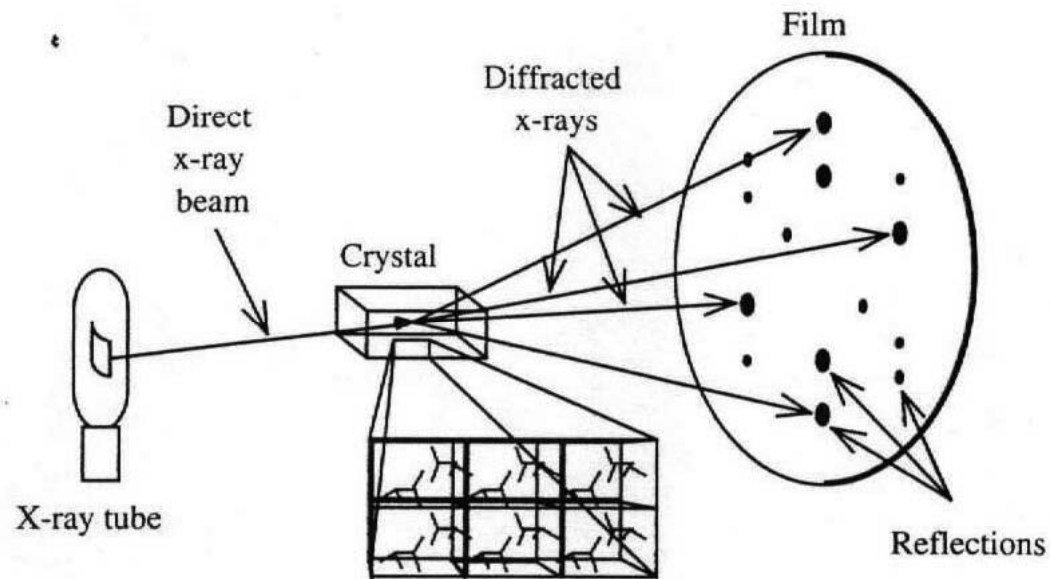


- Crystals
 - ✓ Method: “salting out”

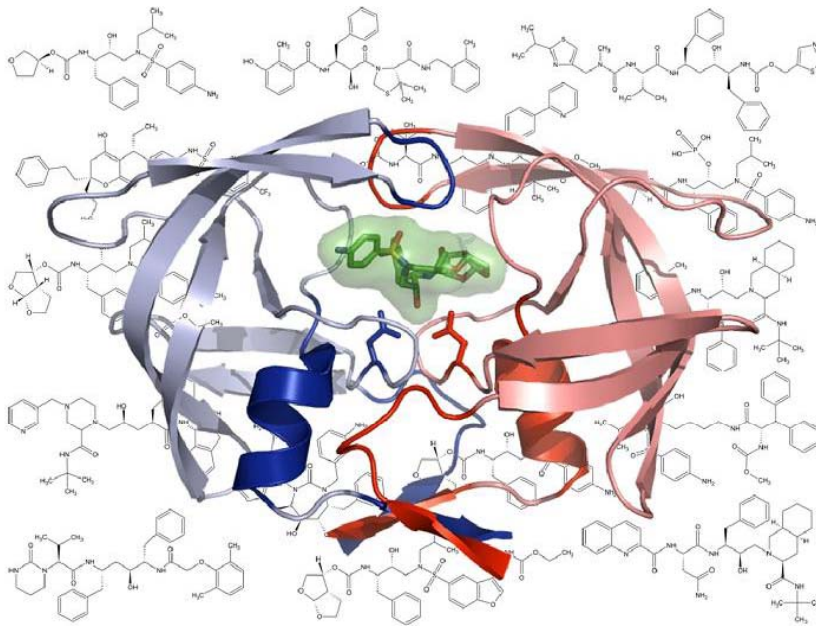


Crystallography – Data Collection

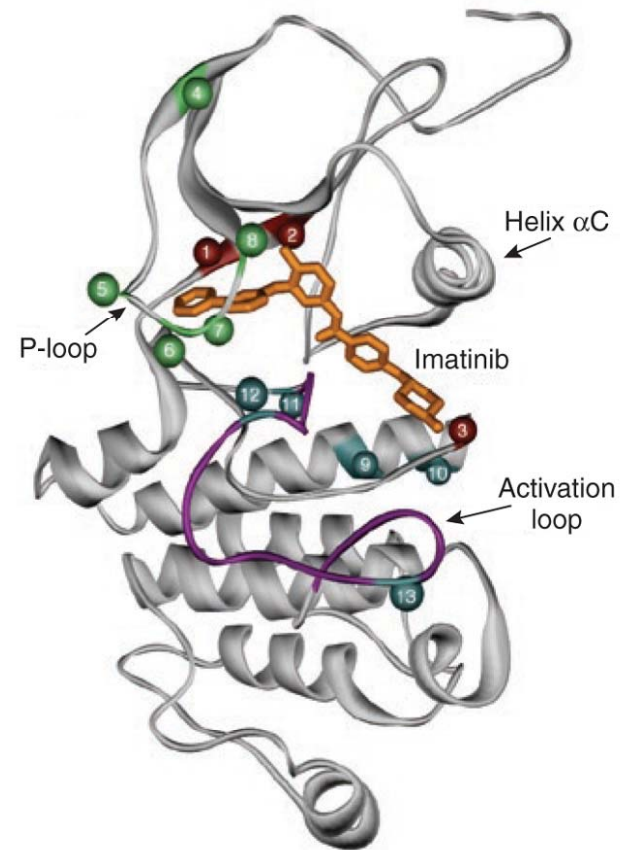
- ✓ Expose crystal to X-rays



Protein Structures in Medicine

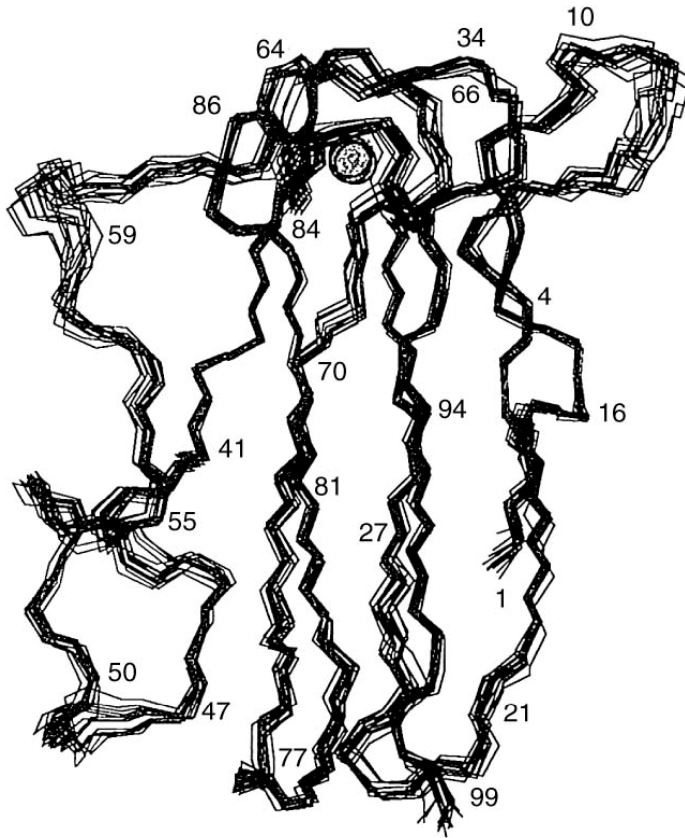


HIV protease
Essential for the
maturation of HIV



cAbl kinase
Constitutively active in
Chronic myelogenous leukemia

NMR



- Advantage:
 - ✓ No crystal needed
- Disadvantage:
 - ✓ limited to small proteins 20kDa

Figure 3.76. NMR structure of plastocyanin from the French bean. From Moore, J.M., Lepre, C. A., Gippert, G. P., Chazin, W. J., Case, D. A., and Wright, P. E. *J. Mol. Biol.* 221:533, 1991. Figure generously supplied by P. E. Wright.